

**XLVI Congresso Nazionale AMCLI**  
Rimini, 11-14 Novembre 2017

*Update sui test di resistenza ai farmaci antvirali*

# **Cosa ci ha insegnato l'esperienza su HIV**

M. Zazzi – Siena



*ui test di resistenza ai farmaci antivirali*  
ha insegnato l'esperienza su HIV

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Nazionale AMCLI

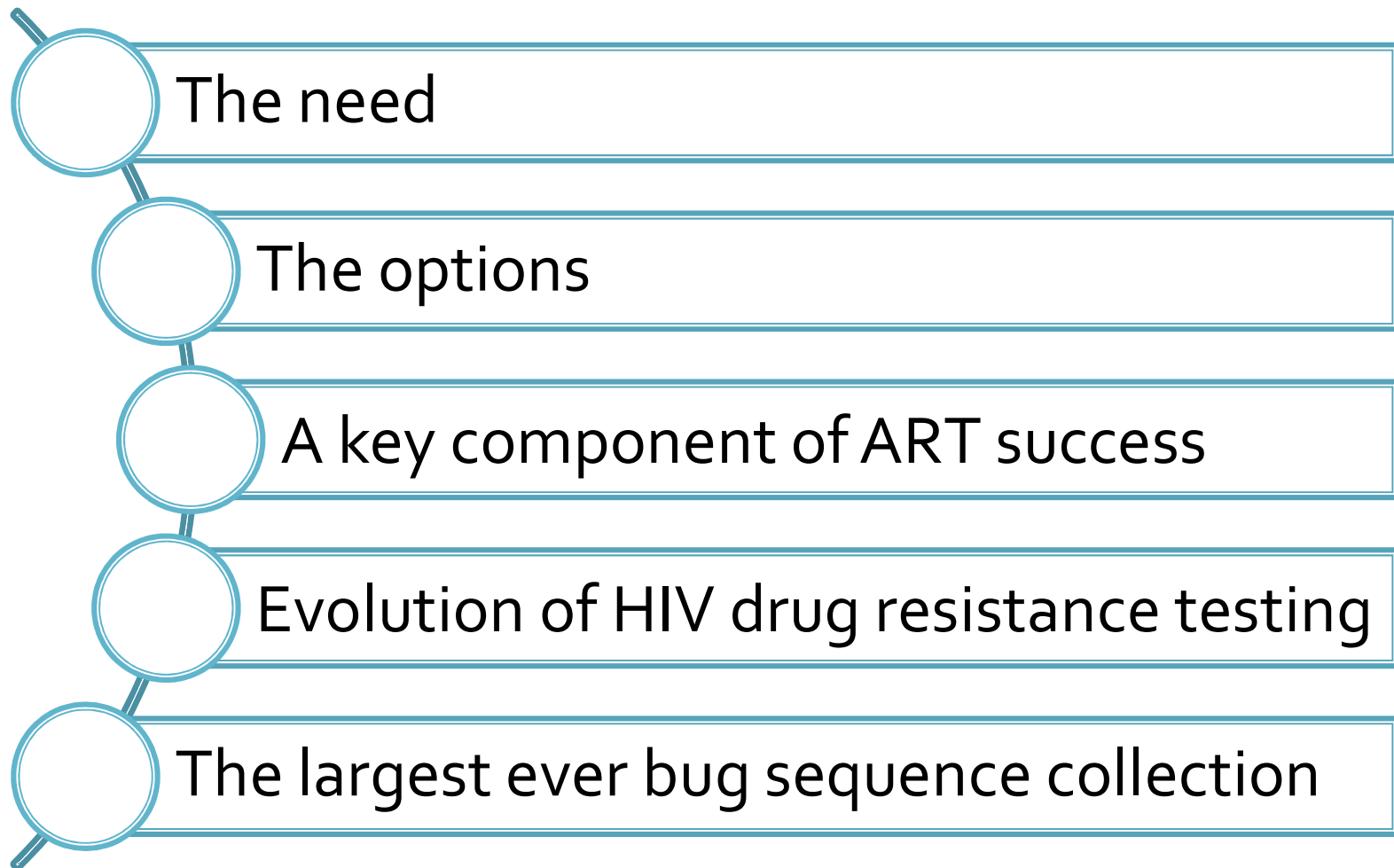
11 - 14 Novembre 2017

Palacongressi di Rimini

# Outline

- The need for HIV drug resistance testing
- The options for HIV drug resistance testing
  - Genotyping replacing phenotyping
- The science of HIV genotype interpretation
  - The multidisciplinary approach and the expert systems
- Drug resistance testing as a key component of the success of antiretroviral therapy
  - The driver for cART
  - The driver for novel drug classes
  - The focus on genetic barrier, adherence and STR
- Evolution of HIV drug resistance testing
  - NGS to replace Sanger?
- Opportunities from the largest ever bug sequence collection
  - Research platforms
  - Molecular epidemiology and surveillance

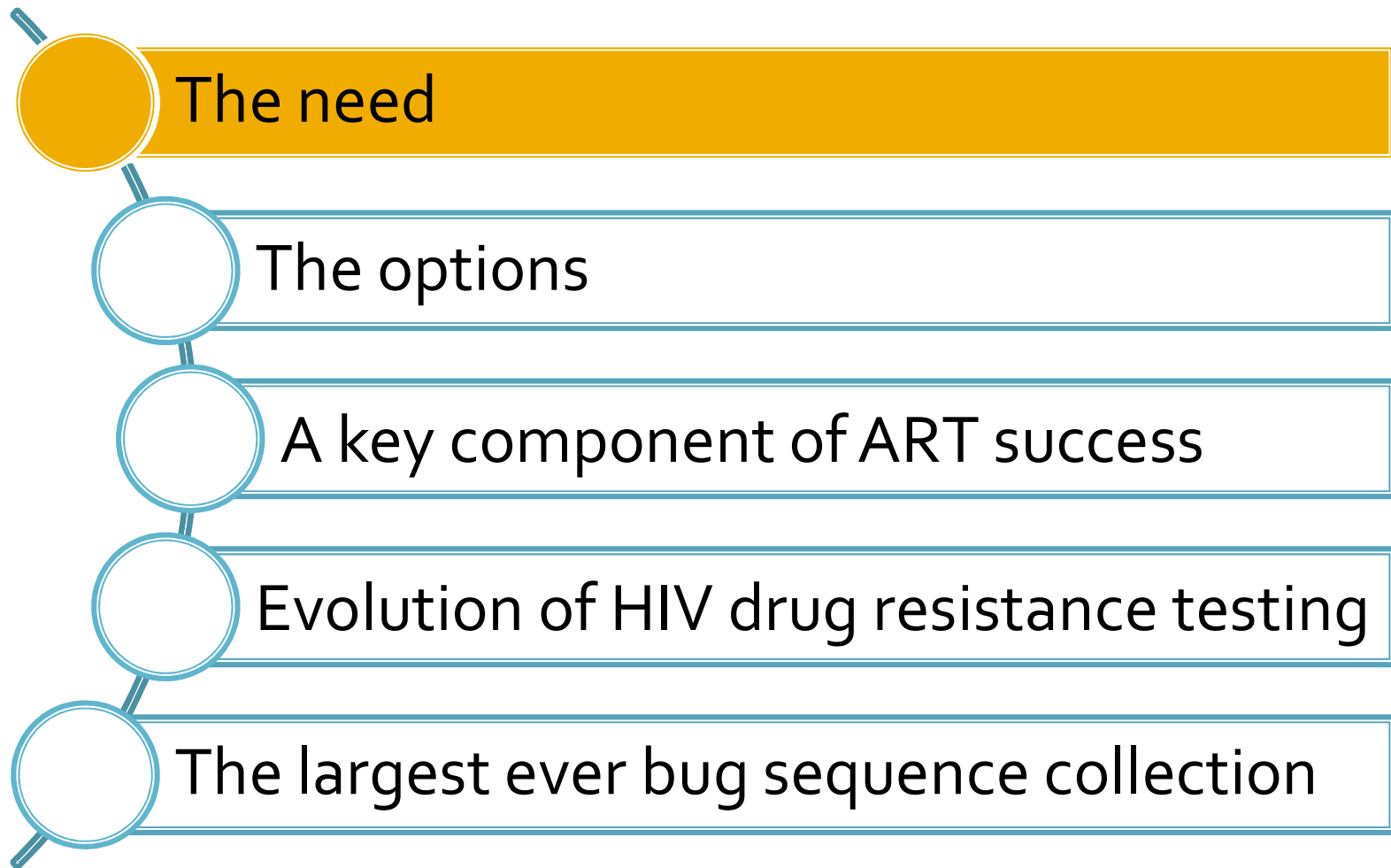
# HIV drug resistance testing





The need

# HIV drug resistance testing



# First drug approved to treat HIV infection



OFFERING INFORMATION ON HIV/AIDS  
TREATMENT, PREVENTION, AND RESEARCH

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## HIV/AIDS News

[Home](#) > [HIV/AIDS News](#) > [Approval of AZT](#)

### Approval of AZT

Date: March 20, 1987

Source: Department of Health and Human Services (DHHS)

Author: Public Health Service (PHS)

Robert E. Windom, M.D., assistant secretary for health, today announced that the Food and Drug Administration has approved the drug zidovudine, commonly known as azidothymidine, or AZT, to help certain patients with Acquired Immunodeficiency Syndrome (AIDS) and advanced AIDS-Related Complex (ARC).

**AZT could offer about a year of prolonged life  
in those early days.**

# First evidence of HIV drug resistance

## HIV with Reduced Sensitivity to Zidovudine (AZT) Isolated During Prolonged Therapy

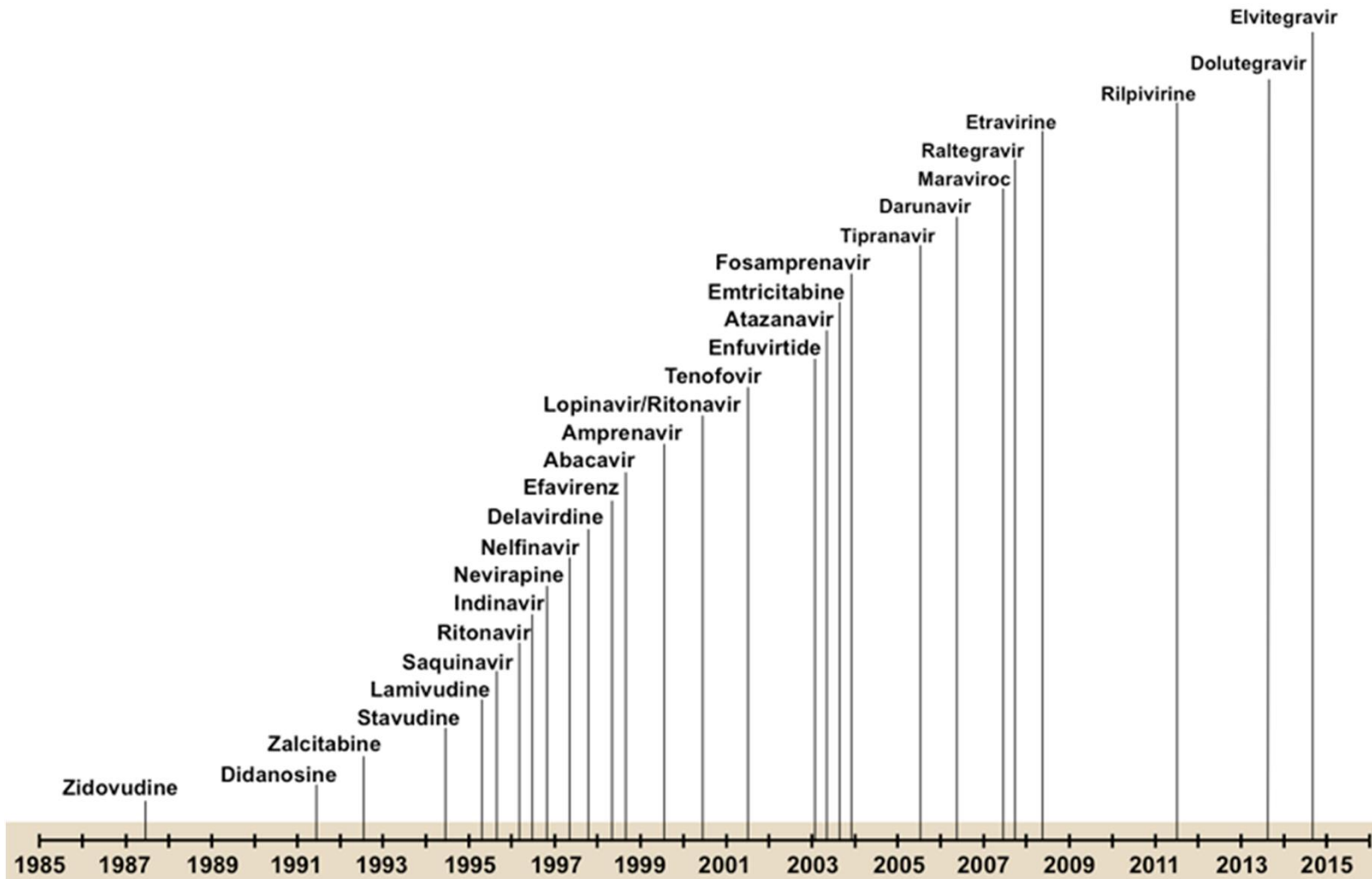
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BRENDAN A. LARDER, GRAHAM DARBY, DOUGLAS D. RICHMAN

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The drug sensitivities of human immunodeficiency virus (HIV) isolates from a group of patients with acquired immunodeficiency syndrome (AIDS) or AIDS-related complex (ARC) who were receiving zidovudine (3'-azido-3'-deoxythymidine, AZT) therapy were tested by means of a newly developed plaque assay in CD4<sup>+</sup> HeLa cells. Fifty percent inhibitory dose (ID<sub>50</sub>) values of 18 isolates from untreated individuals ranged between 0.01  $\mu$ M and 0.05  $\mu$ M. In contrast, most isolates from patients who had received zidovudine for 6 months or more exhibited decreased sensitivity characterized by changes in ID<sub>50</sub> or ID<sub>95</sub> values (or both), with isolates from several patients (5/15) showing 100-fold increases in ID<sub>50</sub>. The latter isolates were also insensitive to 3'-azido-2',3'-dideoxyuridine; however, the isolates were still sensitive to 2',3'-dideoxycytidine, 2',3'-dideoxy-2',3'-didehydrothymidine, or phosphonoformate. It cannot be determined from this small sample of patients whether development of a less sensitive virus phenotype results in clinical resistance. Appearance of such variants was not associated with a consistent increase in viral p24 concentrations in patient plasma and did not herald any sudden deterioration in clinical status. More extensive studies are required to determine the clinical significance. Thus, it would be premature to alter any treatment protocols for HIV-infected individuals at present.

# HIV drug development timeline



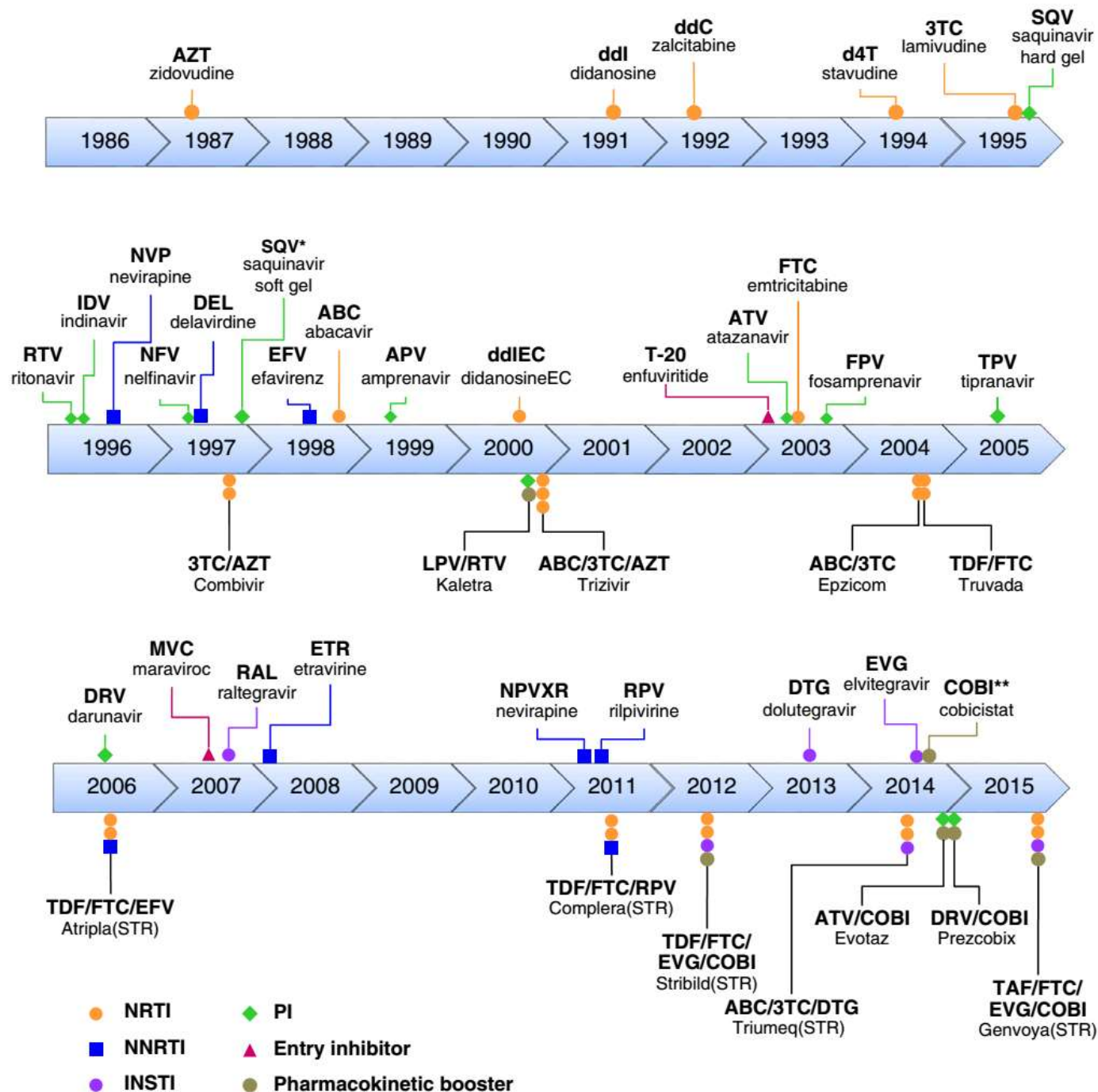
# Anti-HIV compounds & HIV resistance

*"An antiretroviral drug is something to which HIV becomes resistant"*

(Douglas Richman)



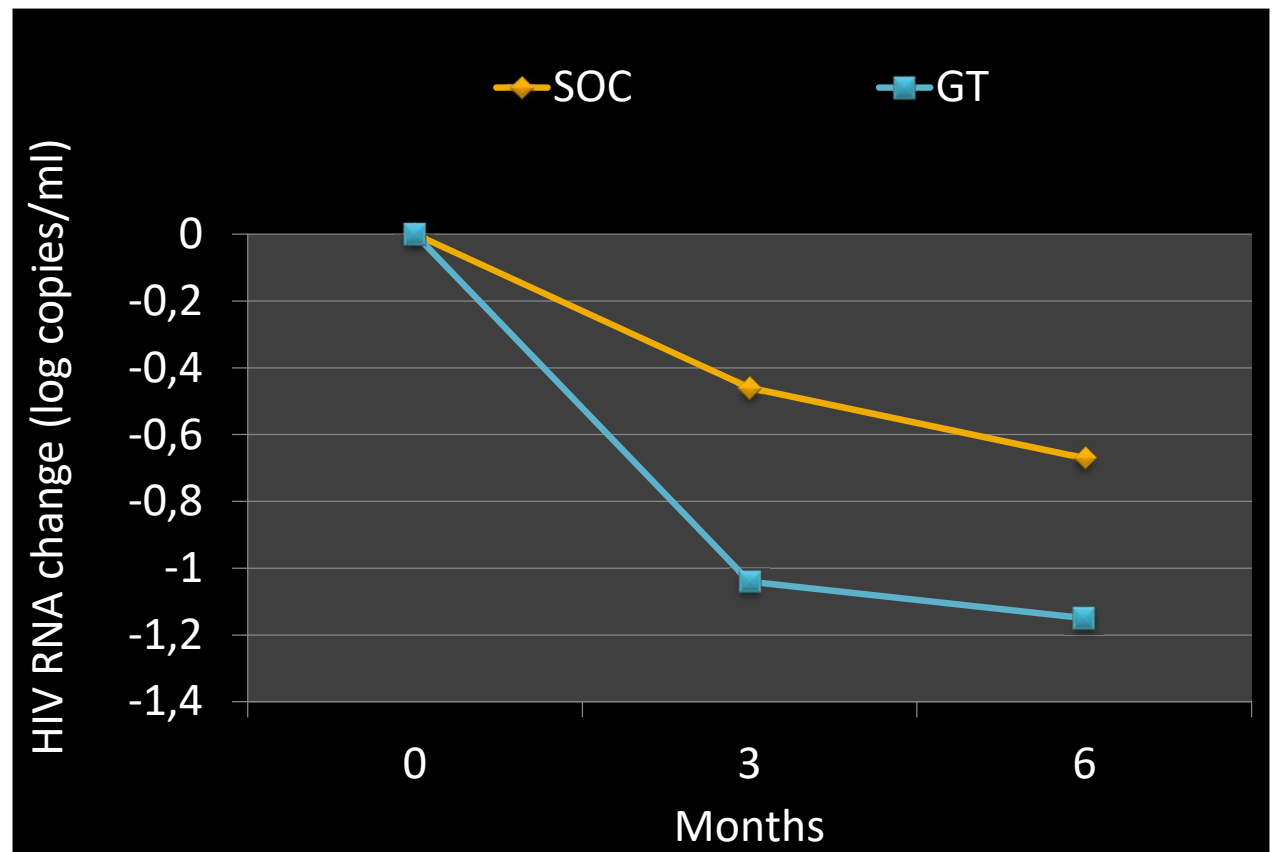
# ART progress



- Only **NRTIs** up to 1995
- 1995 → **PIs**
- 1996 → **NNRTIs**
- Drug resistance regularly documented for each drug
- Also, much cross-resistance within each class

# Drug-resistance genotyping in HIV-1 therapy: the VIRADAPT randomised controlled trial

- Prospective, open, randomised, controlled study
- Patients on treatment failure randomly assigned to standard care (SOC, n=43) or treatment according to the resistance mutations in protease and reverse-transcriptase genes (genotypic group, n=65)
- Endpoint: change in HIV-1 RNA viral load by intention to treat



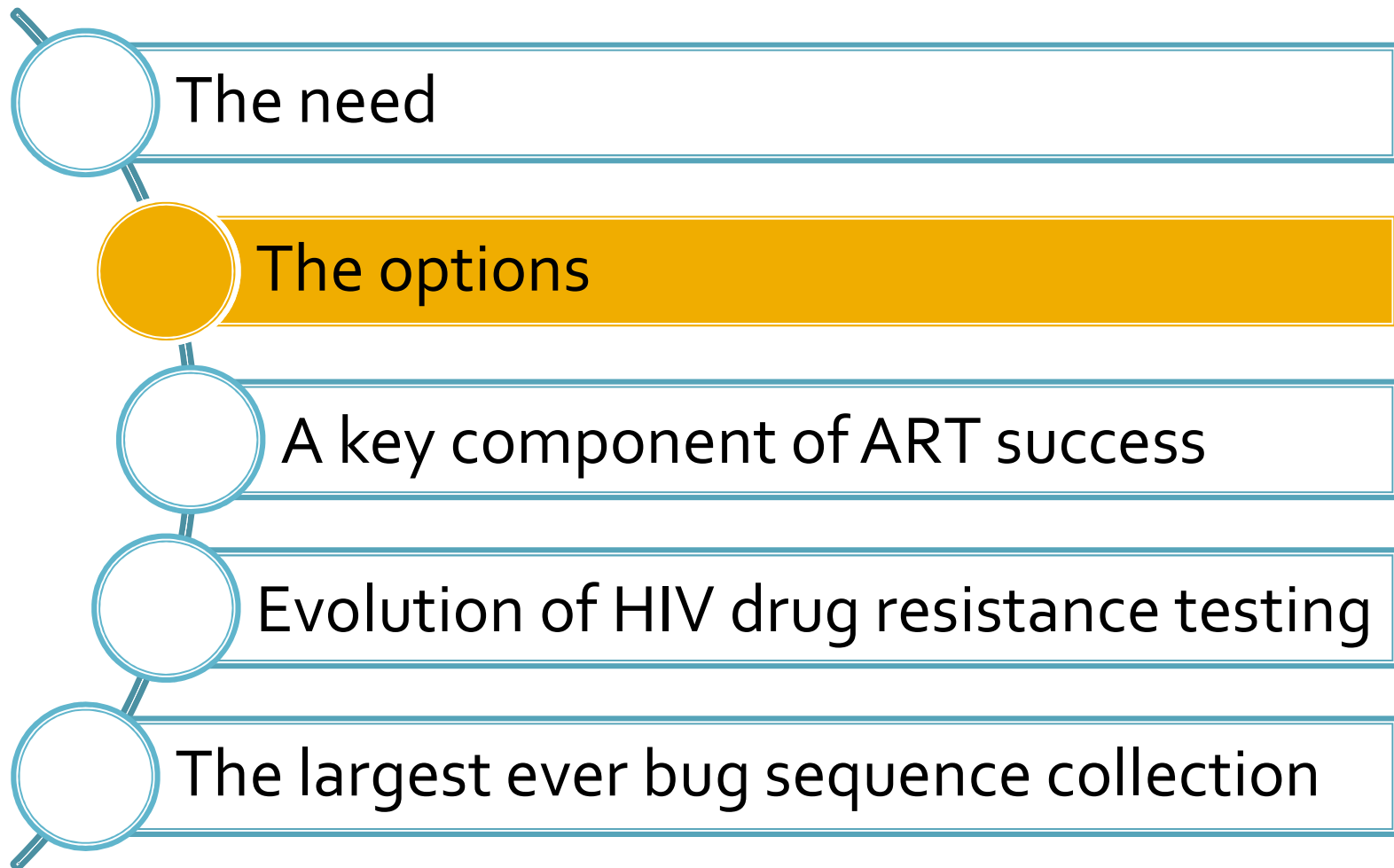
# Pivotal studies supporting the value of drug resistance testing at treatment failure

Study	Patients	Outcome
VIRADAPT (Durant 1999)	108	GENO >SOC
CPCRA 046 (Baxter 2000)	153	GENO > SOC
HAVANA (Tural 2002)	326	GENO+EXPERT OPINION > SOC
CCTG 575 (Haubrich 2005)	256	PHENO = SOC
ARGENTA (Cingolani 2002)	174	GENO > SOC
VIRA3001 (Cohen 2002)	221	PHENO > SOC

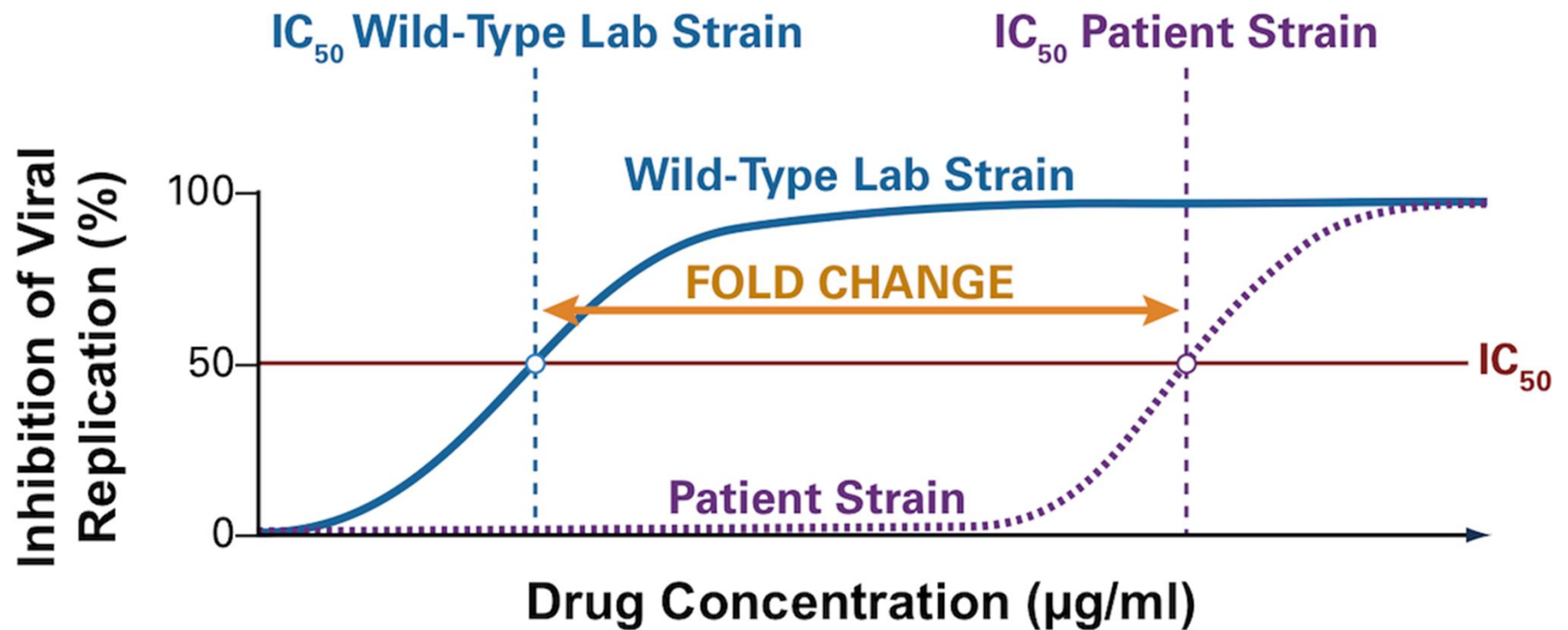
GENO = Genotype; PHENO = Phenotype; SOC = 'standard of care'

# The options

# HIV drug resistance testing



# Measurement of HIV drug resistance in vitro



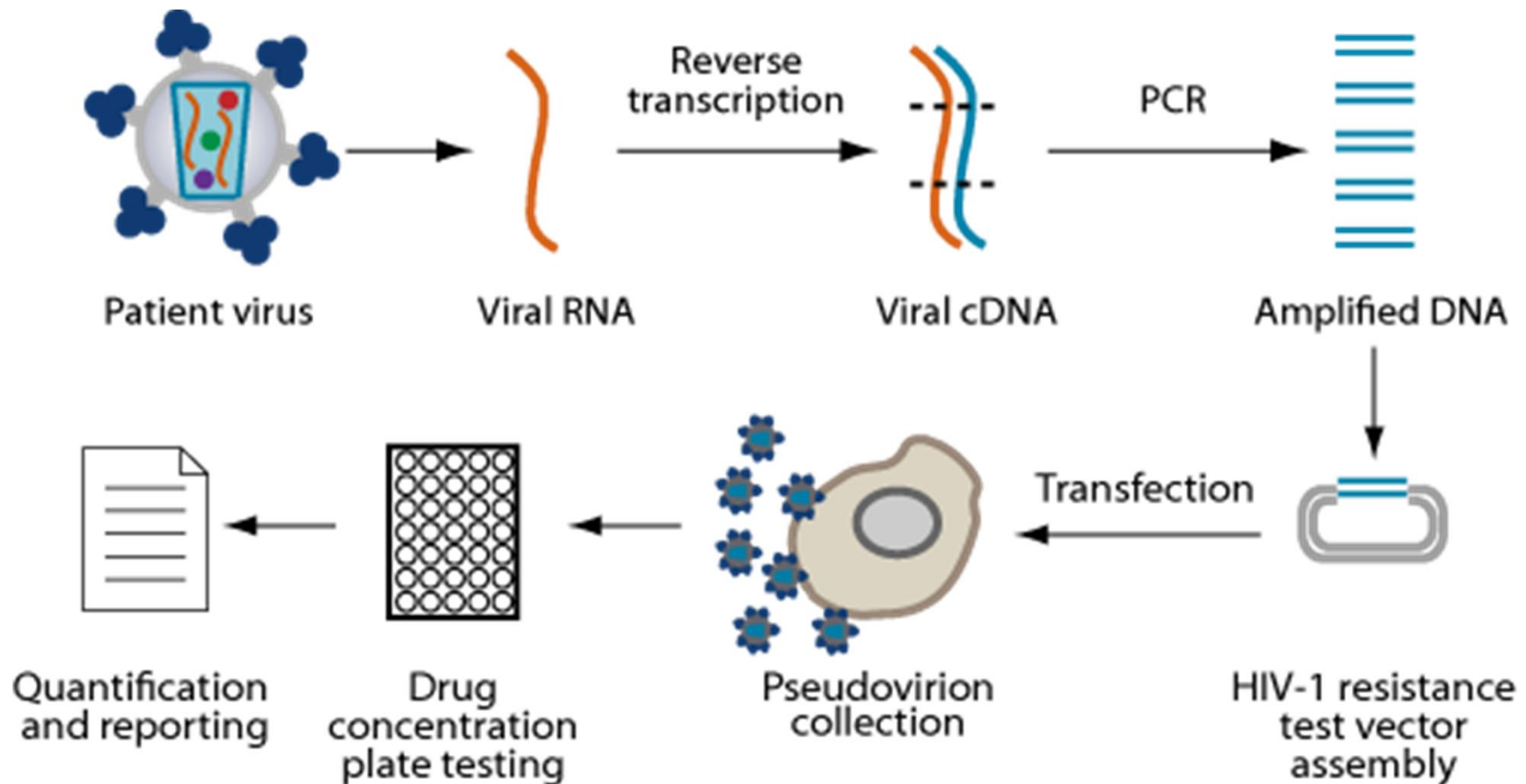


# Measurement of HIV drug resistance in vitro

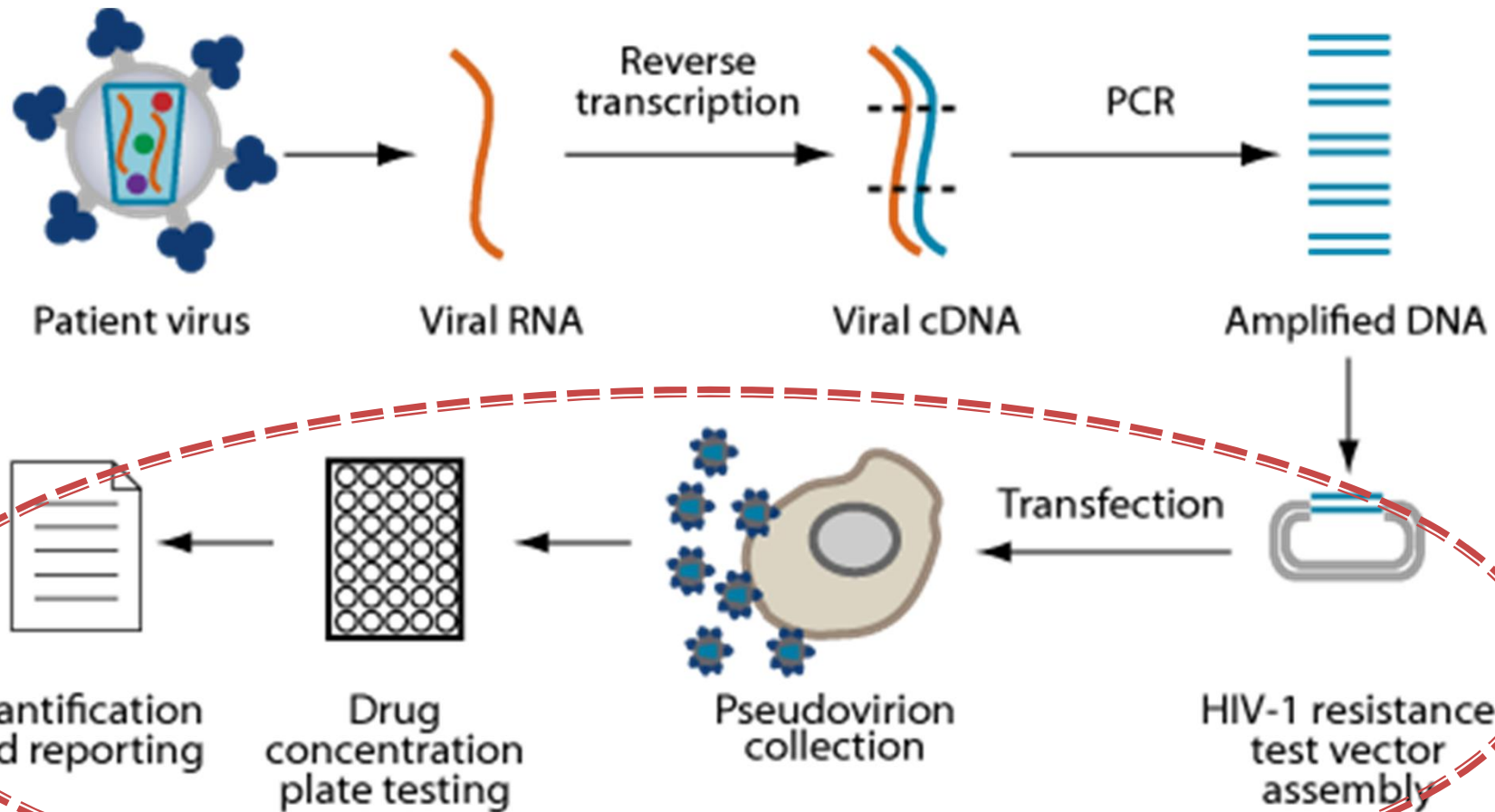
## *A complicated story*

- Most primary isolates do not grow well in cell lines
- Co-culture with (CD8 cell depleted) patient PBMCs and HIV-negative donor PBMCs is required
- An HIV isolate may grow with a different kinetics in different donor PBMCs

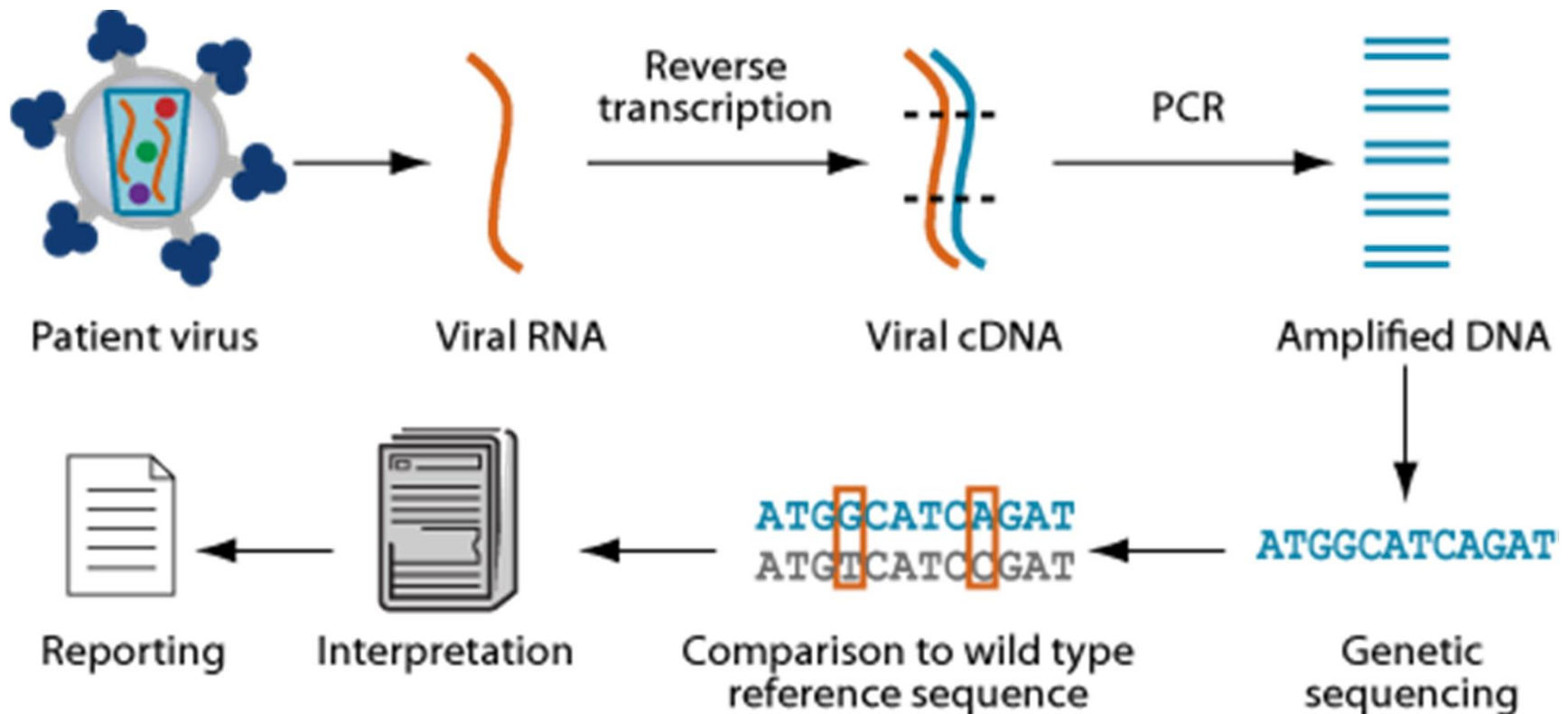
# Phenotypic HIV drug resistance testing



# Phenotypic HIV drug resistance testing



# Genotypic HIV drug resistance testing



# IVD CE marked and FDA approved systems for HIV drug resistance genotyping

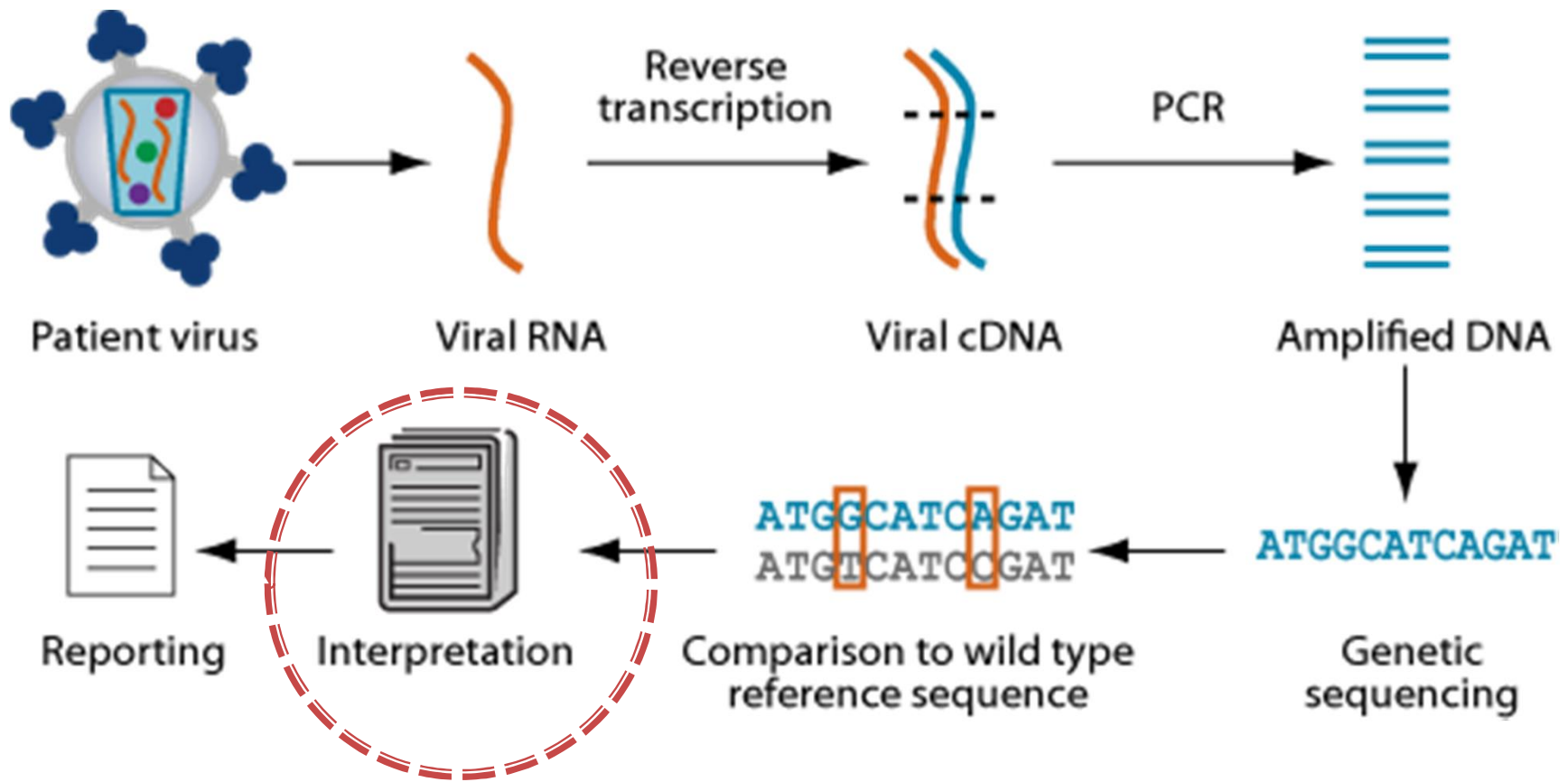


- First DNA sequencing IVD products in infectious diseases
  - Goal: to have HIV genotyping available at every lab with minimal molecular diagnostics background
  - Result: most cases successful but also some failures
- Yet, “expert” labs continues to use homebrew technology

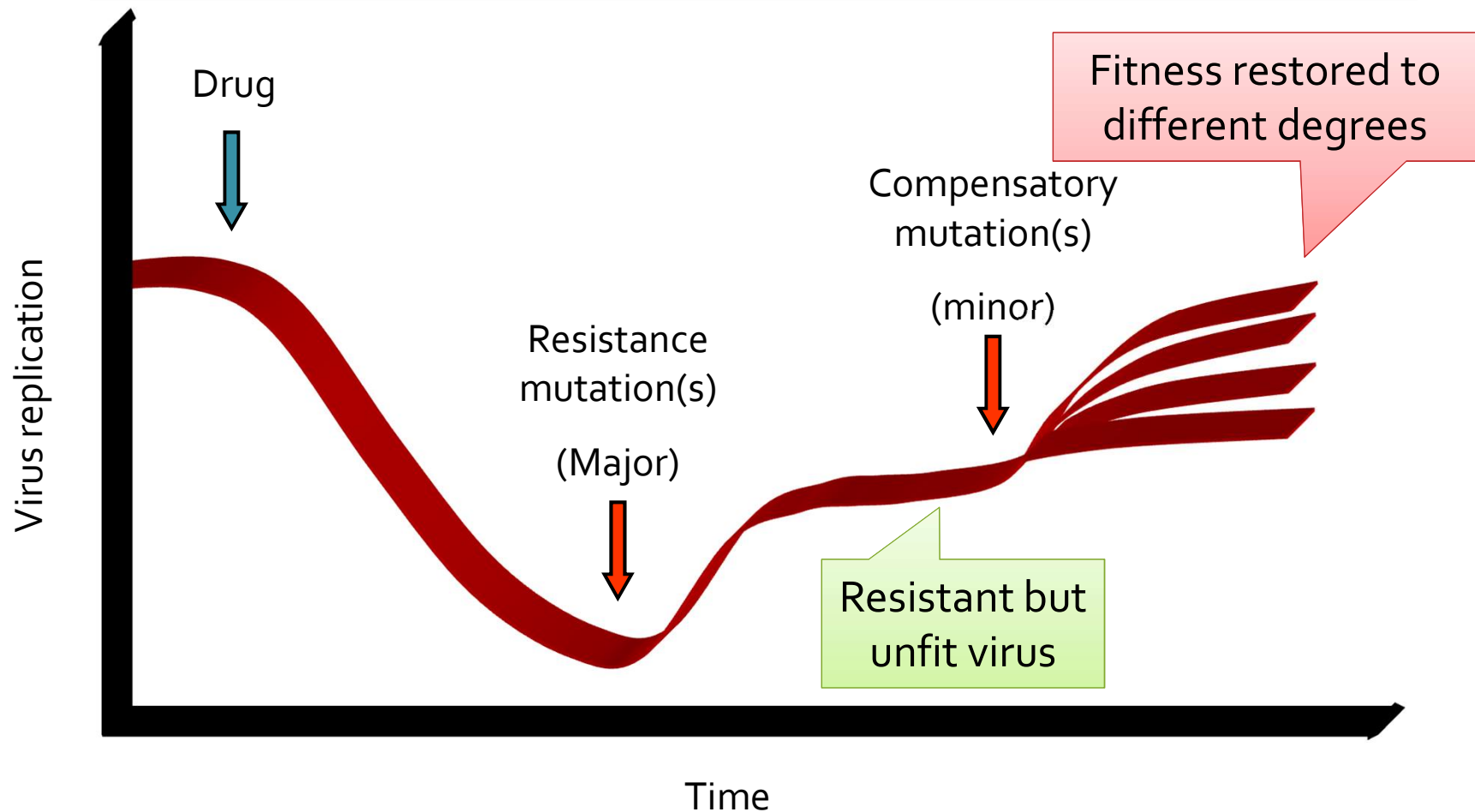
GISs



# Genotypic HIV drug resistance testing

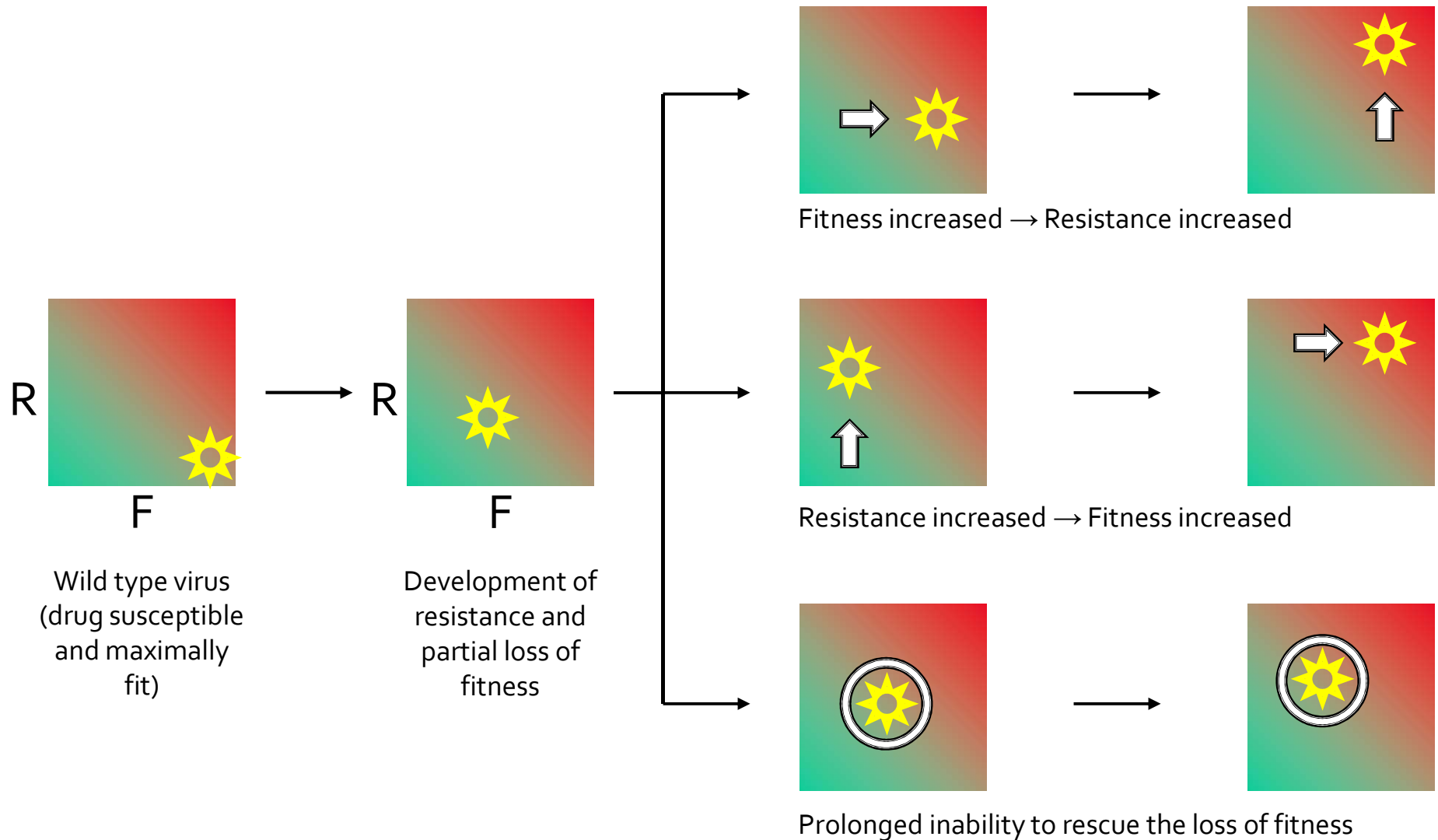


# Resistance and compensatory mutations

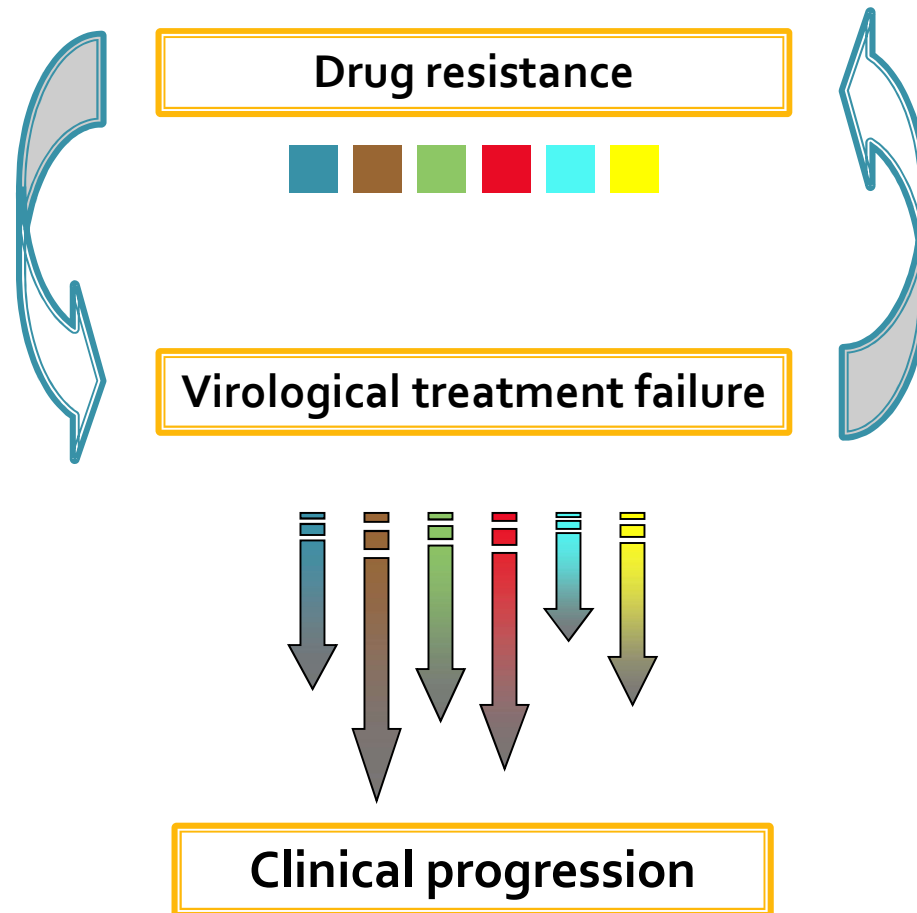


- **Major** mutation → Confers resistance on its own, may often decrease fitness
- **Minor** mutation → Does not confer resistance on its own but may modulate resistance and/or (partially) restore fitness which was decreased by a major mutation

# Drug resistance (R) vs. fitness (F)



# Drug resistance and clinical progression

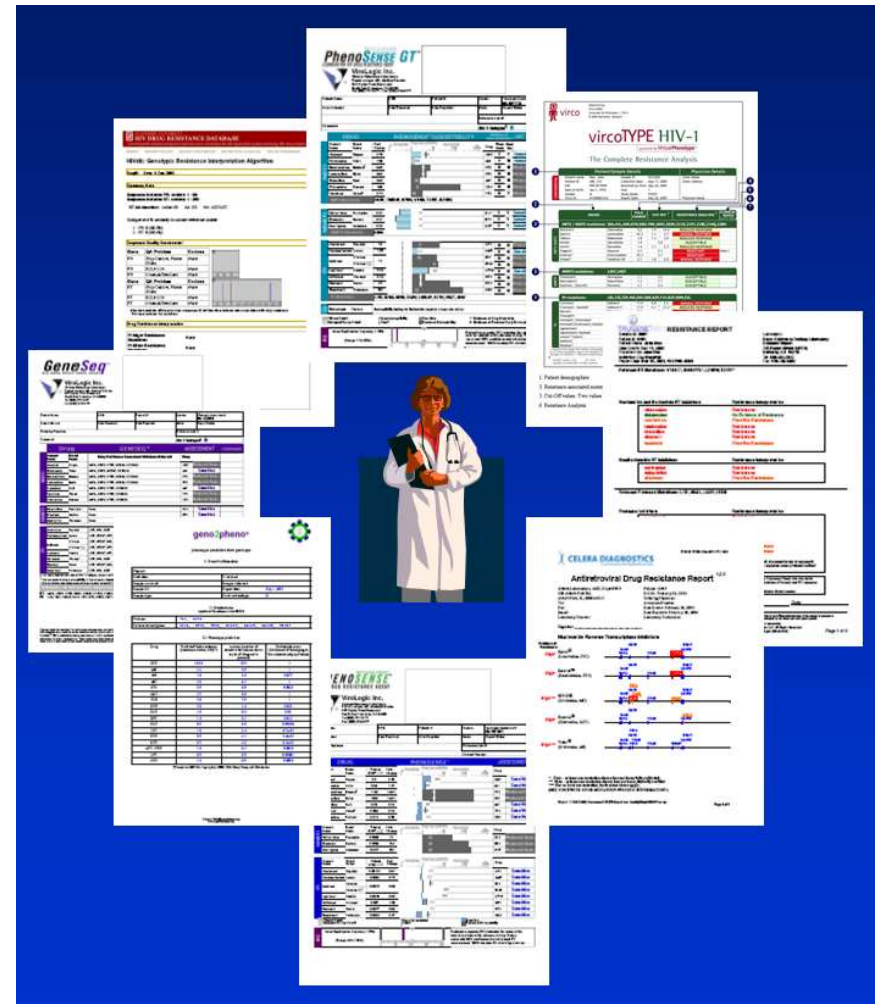


Virological failure is not always / immediately associated with clinical progression

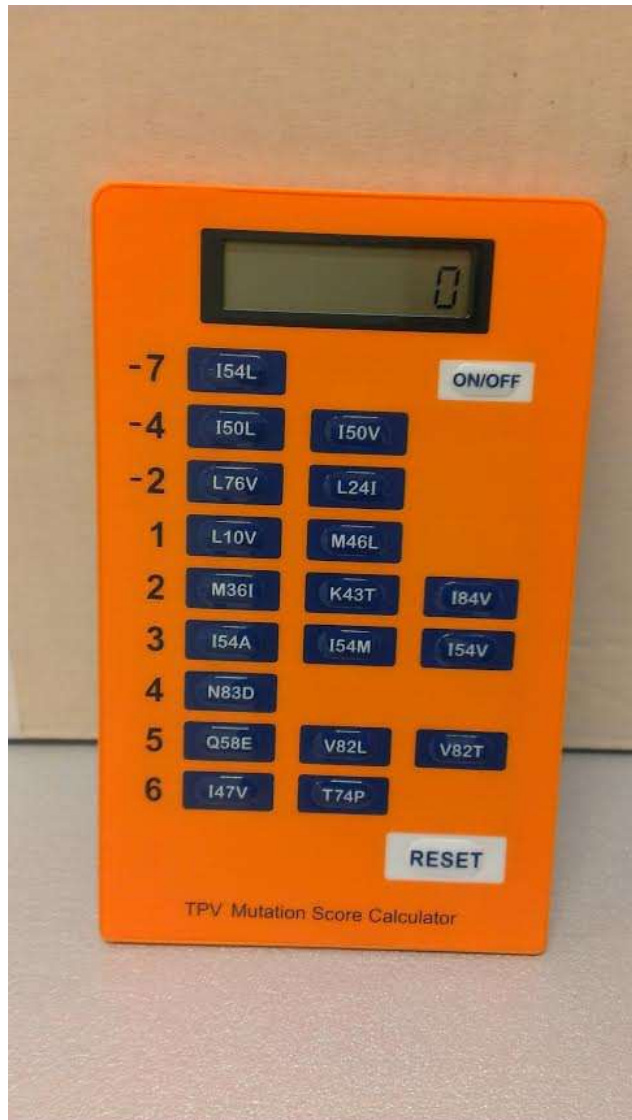
Most drug-resistant variants have reduced pathogenicity

# How to infer drug resistance from HIV sequence analysis

- Multiple options
  - Mutation lists
  - Computer programs
  - Commercial and academic systems
- Some discordances
  - High genetic barrier drugs
- Nevertheless excellent support
  - A must for proper management of treatment choices



# How to infer drug resistance from HIV sequence analysis





# Data sources for HIV genotype interpretation



Stanford University

## HIV DRUG RESISTANCE DATABASE

*A curated public database to represent, store and analyze HIV drug resistance data.*

### Query Pages

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#### Genotype-treatment

Retrieve sequences (and/or mutations) from persons receiving selected HIV drugs

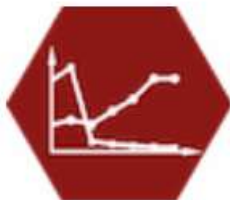
Retrieve sequences and treatments from viruses with specific mutations



#### Genotype-phenotype

Retrieve drug susceptibility data for isolates with selected mutations

Download genotype-phenotype research datasets



#### Genotype-clinical

Summaries of genotype-clinical outcome studies

Genotype-clinical outcome datasets (download)



#### References

Published drug resistance studies in HIVDB

Published studies by Stanford database group

# Most widely used HIV genotype interpretation systems

## HIValg Program

Comparison of Genotypic Resistance Algorithms

HIVdb version 8.4 (last updated 2017-06-16)

HIValg compares HIVdb results to those of 2 other algorithms: i. Rega Institute ([rules](#)), and ii. Agence Nationale de Recherches sur le SIDA (ANRS [rules](#)). However, it does not provide the complete HIVdb report. HIValg also allows users to interpret sequences using any algorithm created using the [Algorithm Specification Interface \(ASI\)](#). A detailed description of the program as well as all updates can be found in the [Release Notes](#).

Protease, RT, and integrase mutations can be entered using either the text box or auto-suggestion boxes. To use the text box, type each mutation separated by one or more spaces. The consensus wildtype and separating commas are optional. If there is a mixture of more than one amino acid at a position, write both amino acids (an intervening slash is optional). Insertions should be indicated by “Insertion” and deletions by “Deletion”.

**Select algorithms**

Select two or more previously published algorithms and/or upload one or more ASI-encoded interpretation algorithms from your computer using the file selection box below.

☒ HIVdb ☒ ANRS ☒ rega [Choose Local ASI2 File\(s\)](#)

Input mutations

Input sequences

Reverse Transcriptase

Input mutation(s)

Protease

Input mutation(s)

Integrase

Input mutation(s)

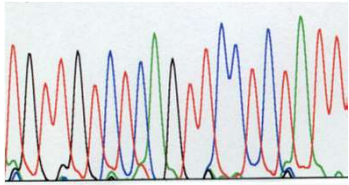
<https://hivdb.stanford.edu/hivalg>

# HIV genotype is a necessary guide to treatment choices

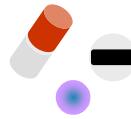
- Best example of successful introduction of **DNA sequence based diagnostics** in the routine management of an infectious disease
- Recommended in all HIV **treatment guidelines** since 2000
  - At treatment failure to detect **acquired resistance**
  - At treatment initiation to detect **transmitted resistance**
  - At pregnancy to adjust treatment and minimize **mother-to-child HIV transmission**

TOSs

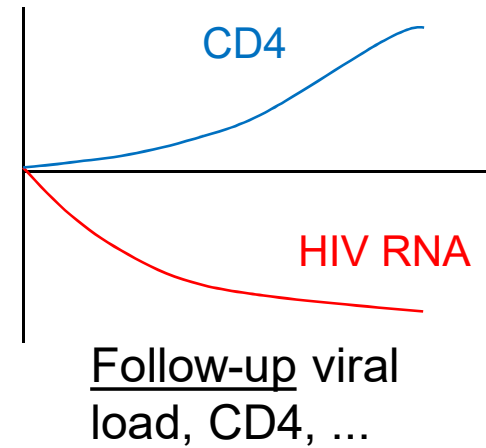
# From HIV genotype to response to treatment



Baseline HIV  
genotype, viral load,  
CD4, previous  
genotypes and drug  
exposure...



Treatment switch



Model training

# From HIV genotype to response to treatment

## **Model training**

Case-based reasoning

Generative-Discriminative Hybrid method

Graph theoretical methods

Fuzzy logic

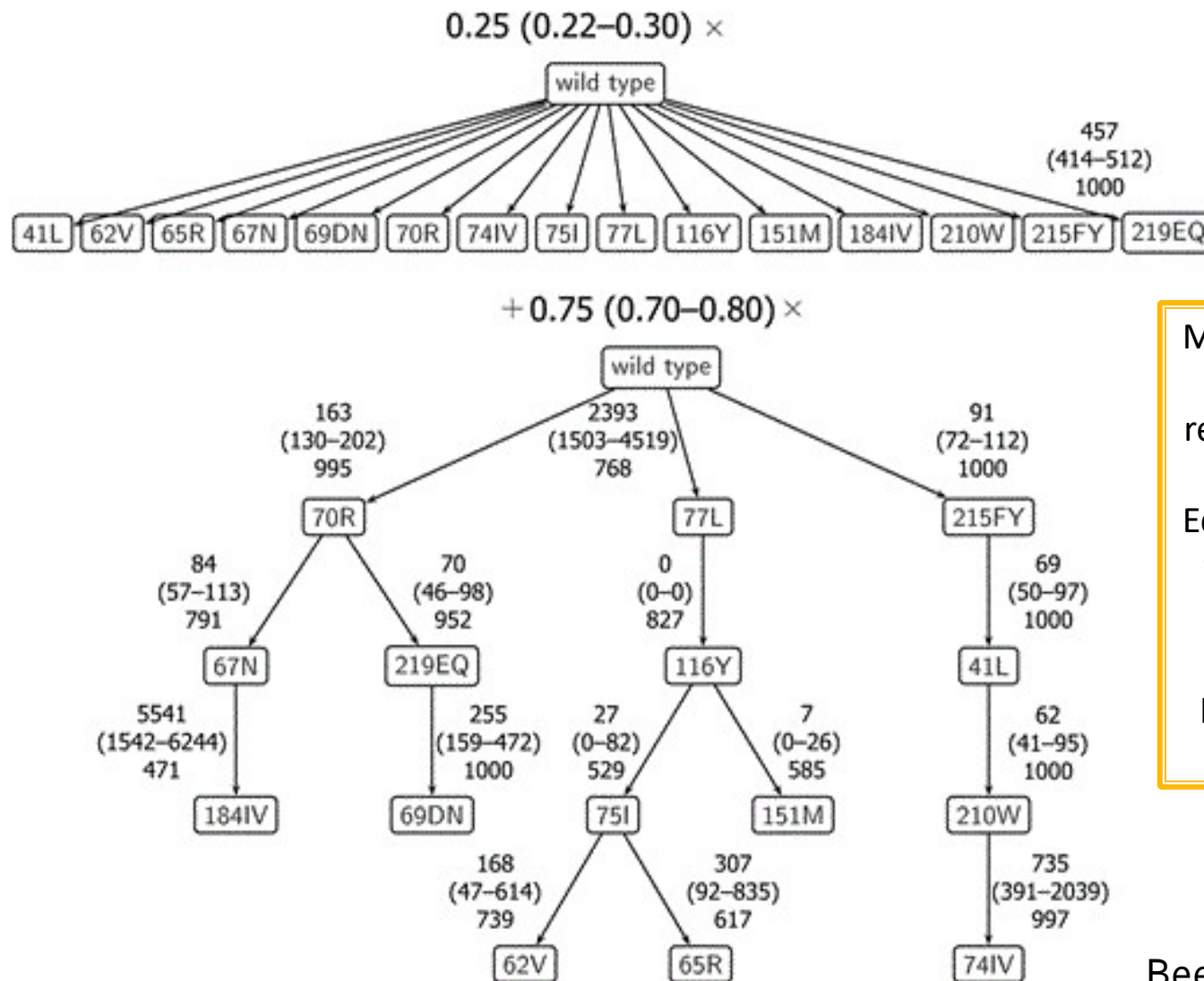
Random Forests

Neural Networks

...but also more popular methods, e. g. logistic regression



# Estimating HIV Evolutionary Pathways and the Genetic Barrier to Drug Resistance



Mixture model of mutagenetic trees for the evolution of resistance during therapy with zidovudine plus didanosine. Edge weights denote expected waiting times in weeks (first row), their 95% confidence intervals (second row, in parentheses), and bootstrap support (third row).



**More and more drug resistance mutations**  
**More and more interactions among mutations**

RESEARCH Open Access

## Open Access

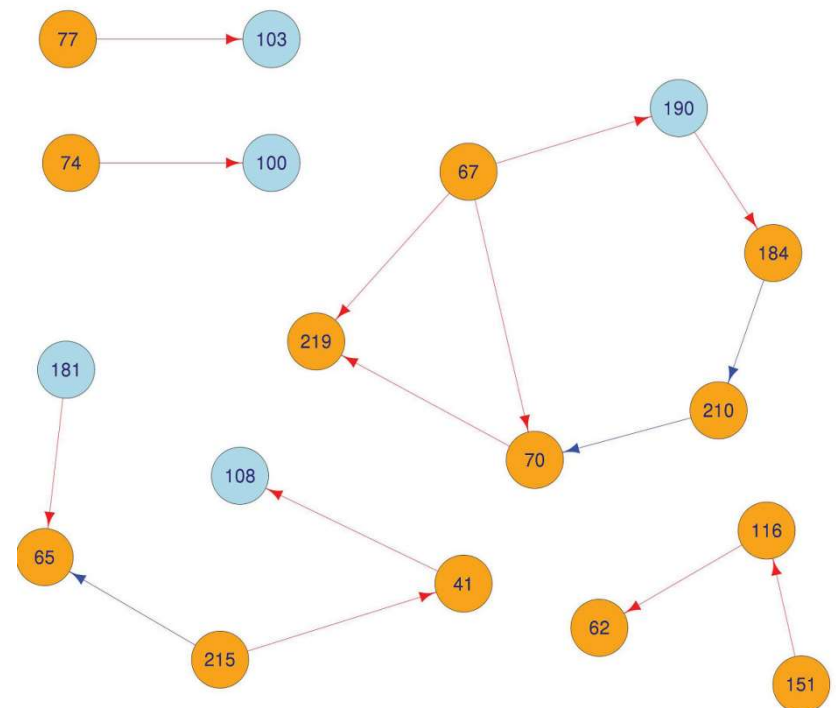
# HIV-1 mutational pathways under multidrug therapy

Glenn Lawyer<sup>1\*</sup>, André Altmann<sup>2</sup>, Alexander Thielen<sup>1</sup>, Maurizio Zazzi<sup>3</sup>, Anders Sönnnerborg<sup>4</sup> and Thomas Lengauer<sup>1</sup>

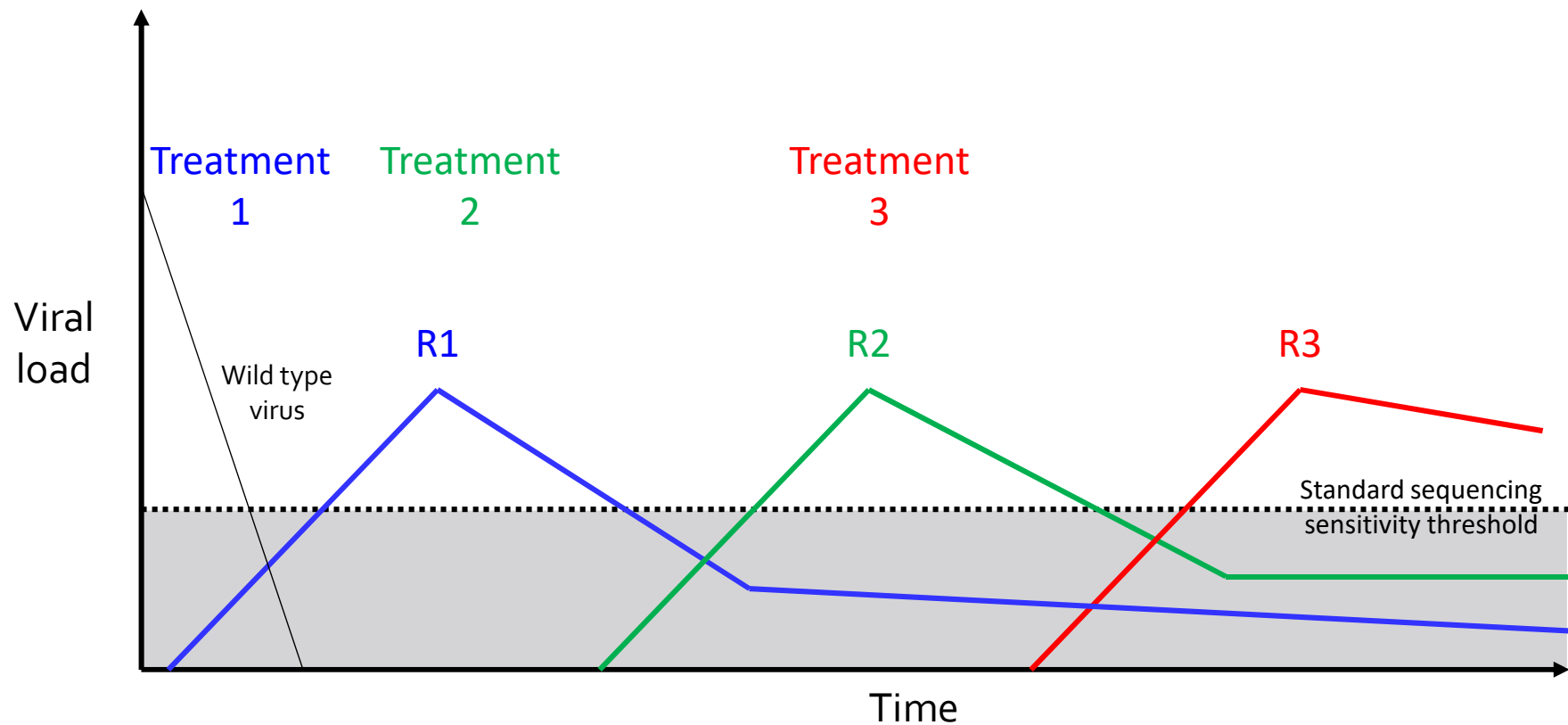
AIDS Research and Therapy 2011, 8:26

**Resistance mutations adjacency graph.** Red pathways indicate an increase in hazard, blue a reduction.

Orange nodes are locations hosting NRTI resistance mutations, blue nodes are NNRTI locations.



# Kinetics of drug resistance species following multiple treatment failures



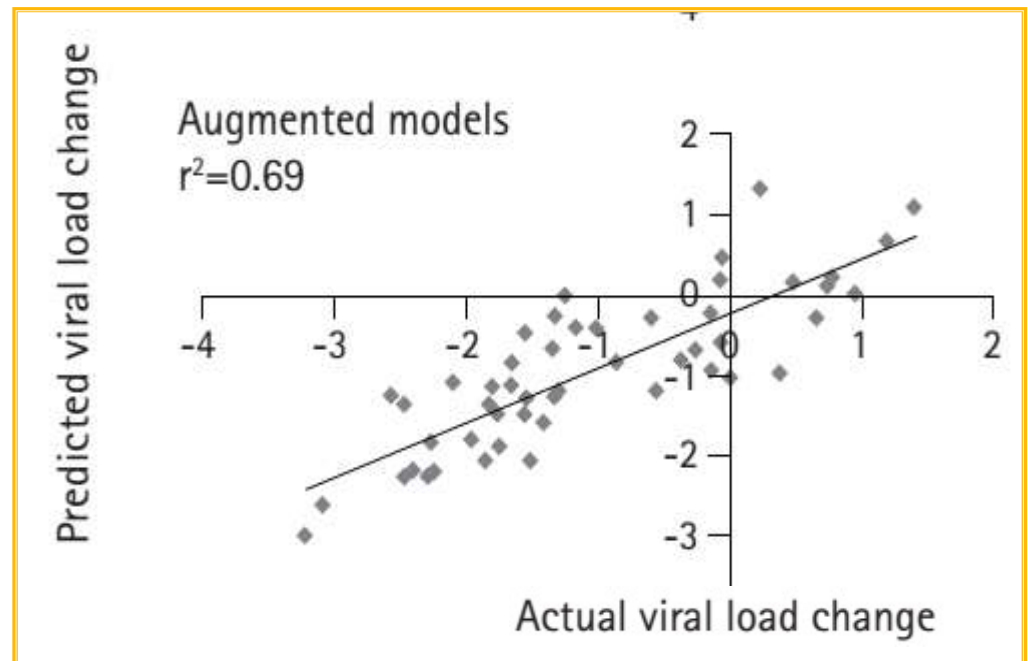
# How to infer response to treatment from HIV sequence analysis (from GIS to TOS)

Antiviral Therapy 12:15–24

- From **Genotype Interpretation Systems to Treatment Optimization Systems**
- Many “complex” learning methods
  - Neural networks
  - Support Vector Machines
  - Random Forests
  - ...
- Partly successful

The development of artificial neural networks to predict virological response to combination HIV therapy

*Brendan Larder<sup>1\*</sup>, Dechao Wang<sup>1</sup>, Andrew Revell<sup>1</sup>, Julio Montaner<sup>2</sup>, Richard Harrigan<sup>2</sup>, Frank De Wolf<sup>3</sup>, Joep Lange<sup>4</sup>, Scott Wegner<sup>5</sup>, Lidia Ruiz<sup>6</sup>, María Jesús Pérez-Eliás<sup>7</sup>, Sean Emery<sup>8</sup>, Jose Gatell<sup>9</sup>, Antonella D'Arminio Monforte<sup>10</sup>, Carlo Torti<sup>11</sup>, Maurizio Zazzi<sup>12</sup> and Clifford Lane<sup>13</sup>*



# How to infer response to treatment from HIV sequence analysis (from GIS to TOS)

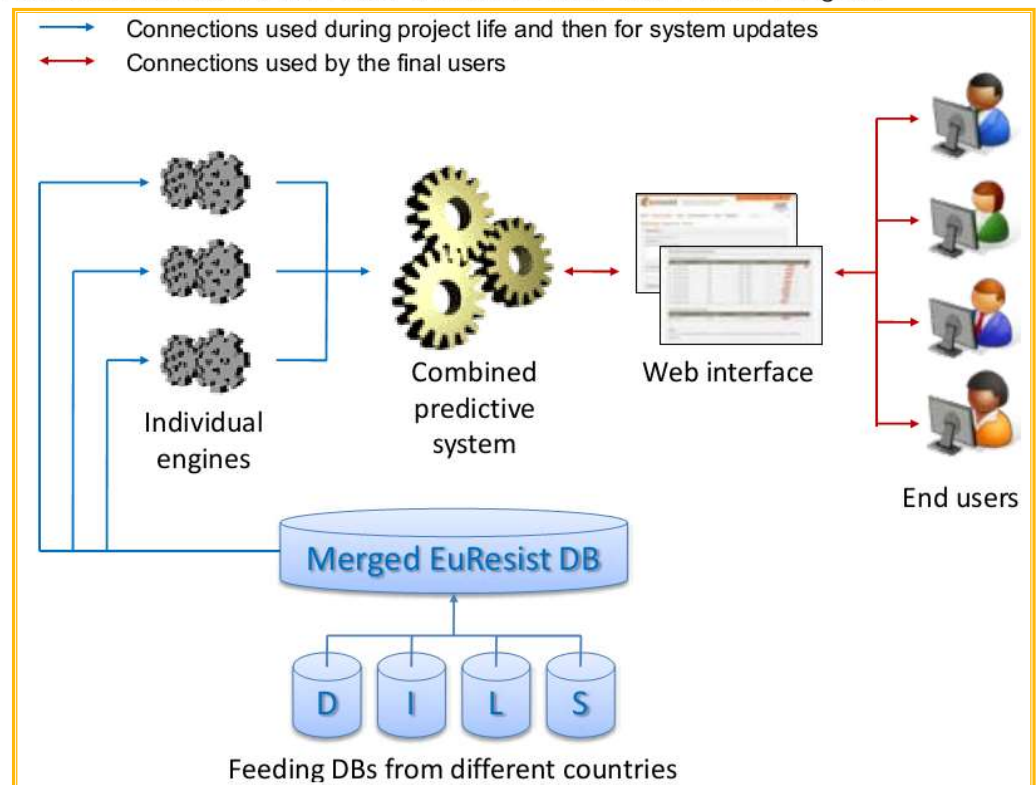
BIOINFORMATICS

Vol. 24 ISMB 2008, pages i399–i406  
doi:10.1093/bioinformatics/btn141

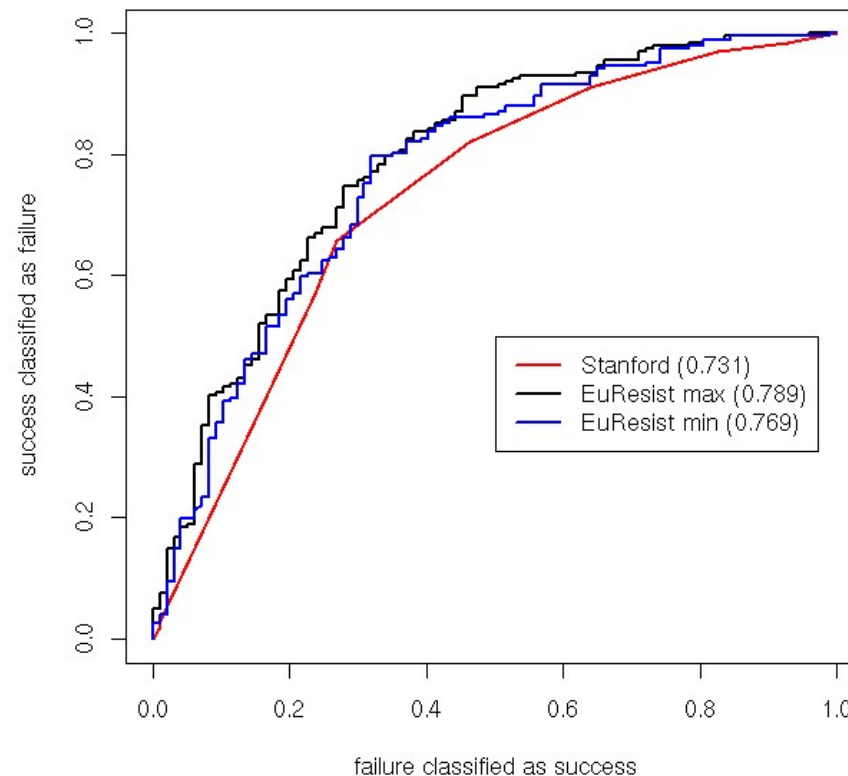
- From **Genotype Interpretation Systems to Treatment Optimization Systems**
- Many “complex” learning methods
  - Neural networks
  - Support Vector Machines
  - Random Forests
  - ...
- Partly successful

## Selecting anti-HIV therapies based on a variety of genomic and clinical factors

Michal Rosen-Zvi<sup>1,\*</sup>, Andre Altmann<sup>2</sup>, Mattia Prosperi<sup>3</sup>, Ehud Aharoni<sup>1</sup>, Hani Neuvirth<sup>1</sup>, Anders Sönnernborg<sup>4</sup>, Eugen Schüller<sup>5</sup>, Daniel Struck<sup>6</sup>, Yardena Peres<sup>7</sup>, Francesca Incardona<sup>8</sup>, Rolf Kaiser<sup>5</sup>, Maurizio Zazzi<sup>9</sup> and Thomas Lengauer<sup>2</sup>

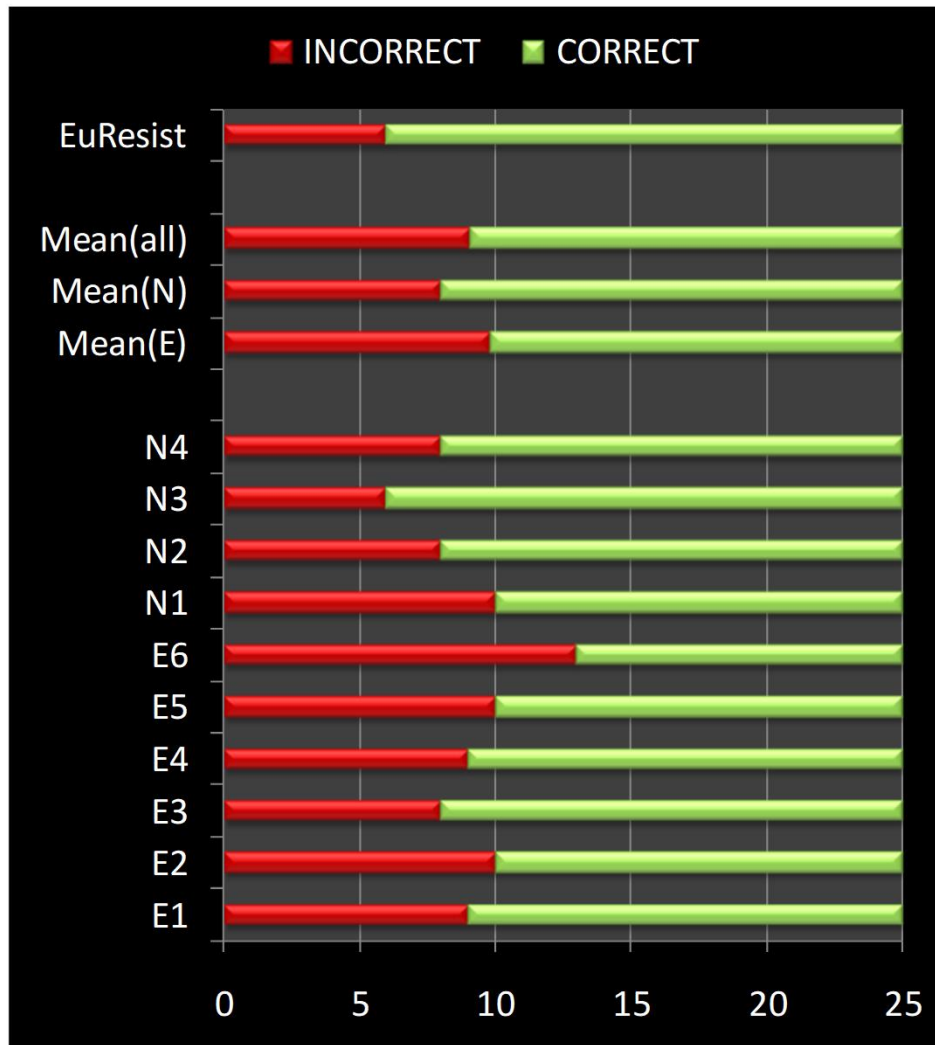


# How to infer response to treatment from HIV sequence analysis



3143 therapies, Short-term outcome (8 weeks)

# EuResist vs. Expert interpretation (EvE study)



- 25 ART cases randomly selected from the EuResist db
  - Obsolete therapies and wild type genotype excluded
  - All clinical and virological information available
- EuResist engine used to predict success/failure
- 10 leading experts examining patient charts to predict success/failure
  - On-line anonymous rating
  - Only European (E) vs. non-European (N) setting traceable
  - Use of any interpretation system allowed
- Top accuracy (76%) reached by EuResist and one expert



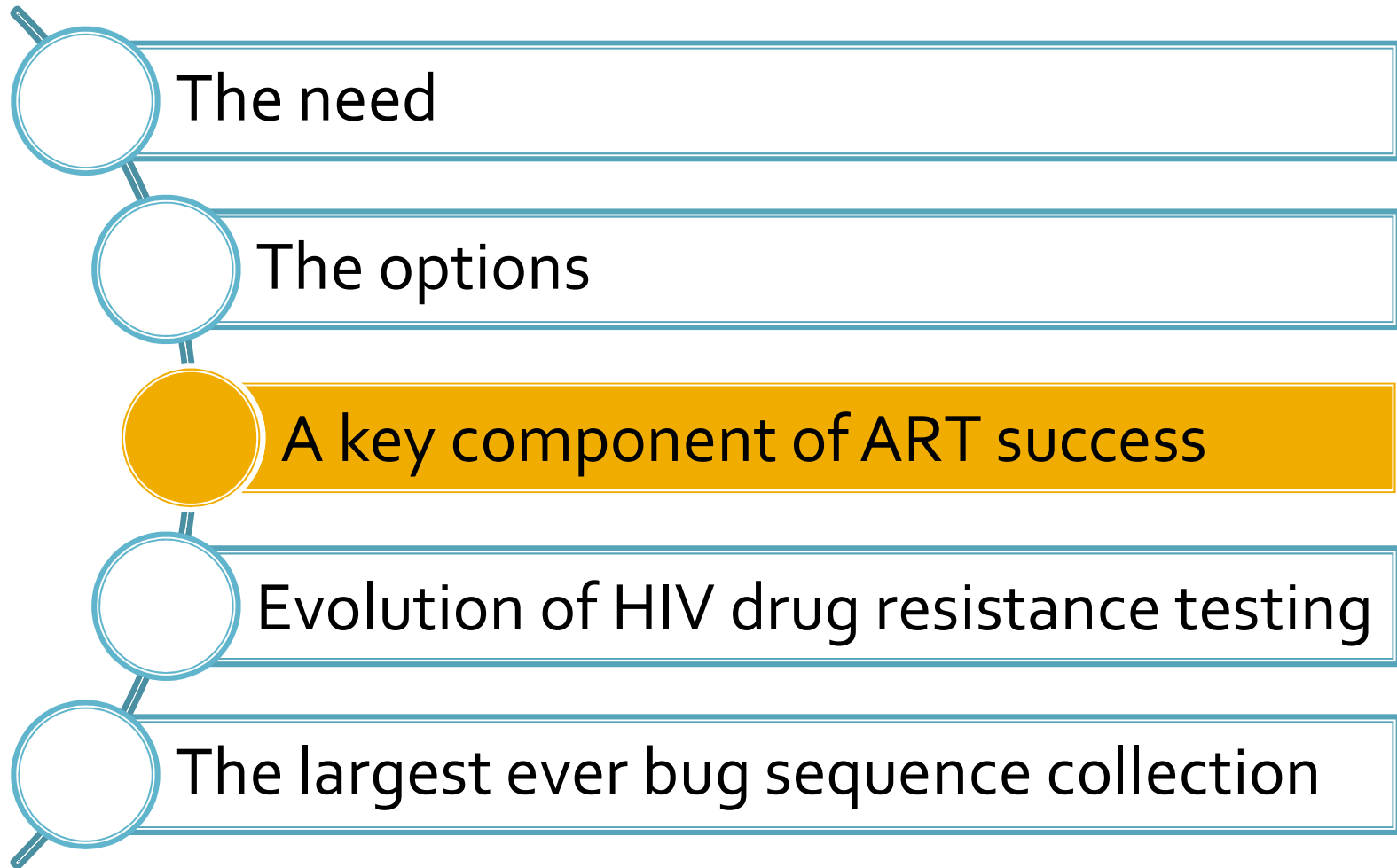
# Why "Treatment Optimization Systems" have not gained wide attention

- **Genotype interpretation systems** are well consolidated and **work satisfactorily** (i.e. HIVdb, REGA, ANRS)
- TOSs are based on complex and non-transparent functions (**black box**), the user is not comfortable with
- The increase in accuracy is not dramatic and is more relevant with **old therapies** (larger training datasets)
- The potential benefits may come at the **expense of providing additional patient data**
- **The incidence and impact of drug resistance have decreased over time**



GRT as a key for  
ART success

# HIV drug resistance testing

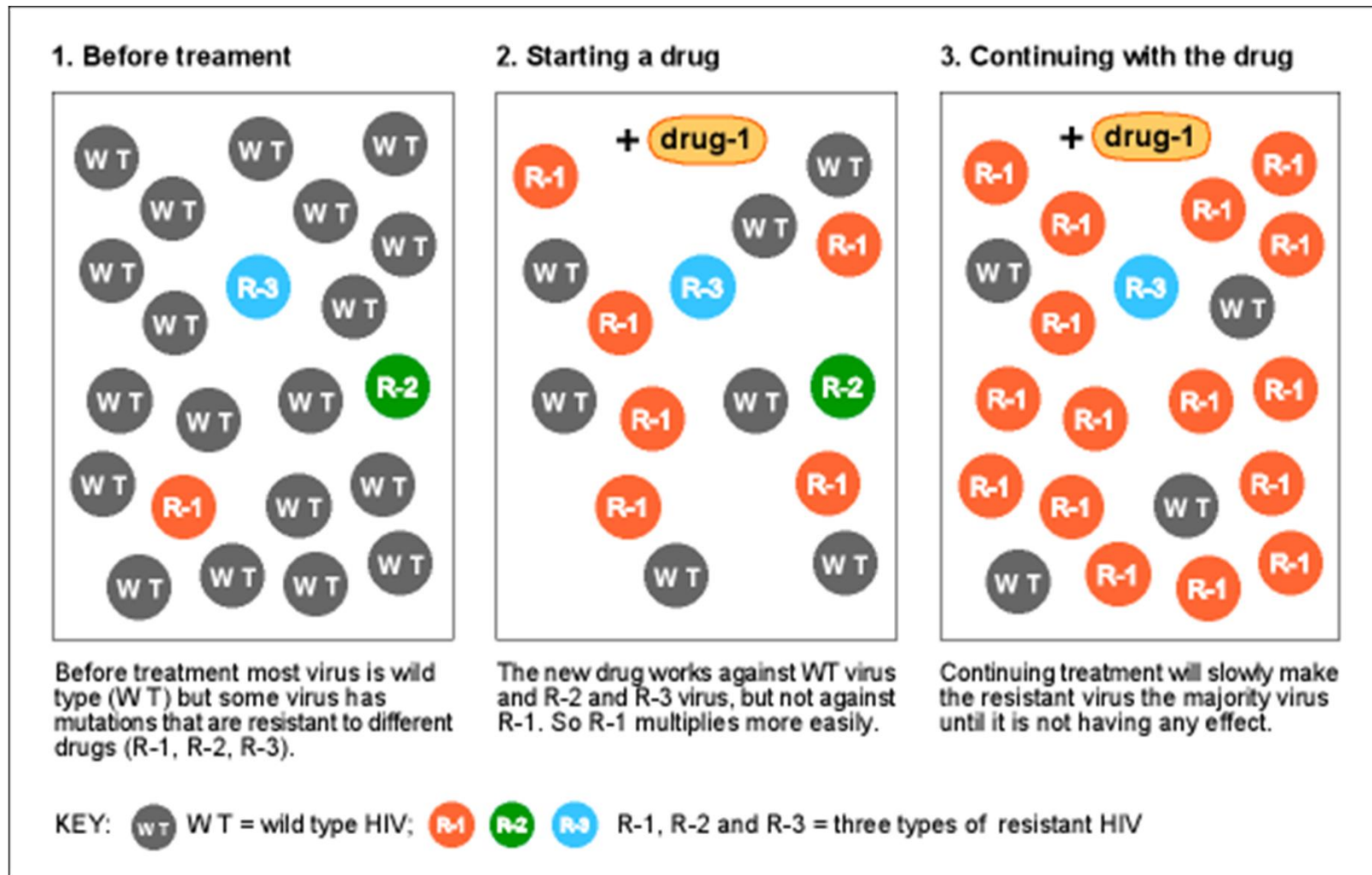


# Drug resistance testing as a key component of the success of antiretroviral therapy

Drug resistance testing provided a basis for combination ART



# Drug resistance emerging as a variability and selection process



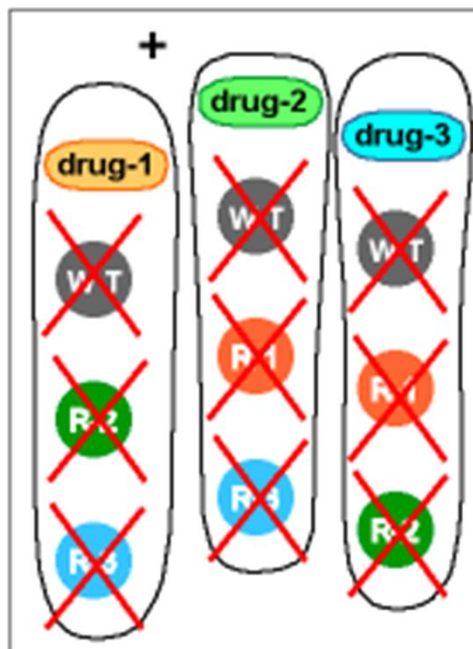
# Drug resistance is the driver for combination therapy

1. Treatment with three active drugs



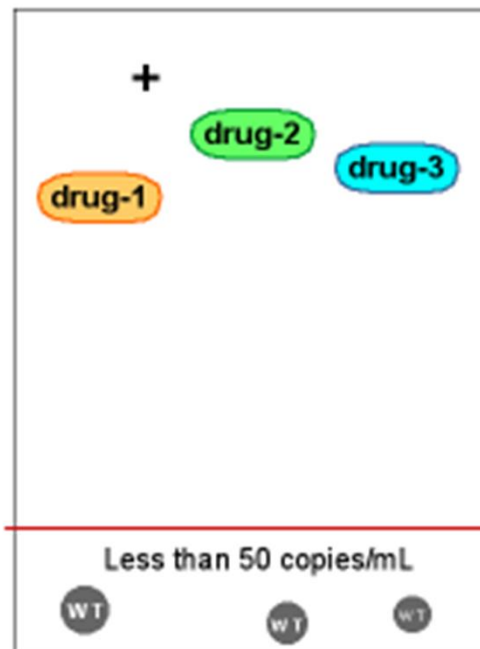
Because single mutations are common before treatment, it is important to use three drugs in combination.

2. Each drug is active on different types of HIV



Each drug is active against the resistant virus that does not affect it. HIV that is resistance to one drug is killed by one of the others.

3. Resistant and wild-type HIV is reduced to 'undetectable'



With three active drugs viral load is reduced, including the resistant virus. The combination needs three active drugs to be strong enough.

KEY: WT = wild type HIV; R-1, R-2, R-3 = three types of resistant HIV

drug-1, drug-2, drug-3 = drug 1, 2 and 3 and three drugs, resistant to R-1, R-2 and R-3 respectively.

# Drug resistance testing as a key component of the success of antiretroviral therapy

Drug resistance testing highlighted the key concept of genetic barrier





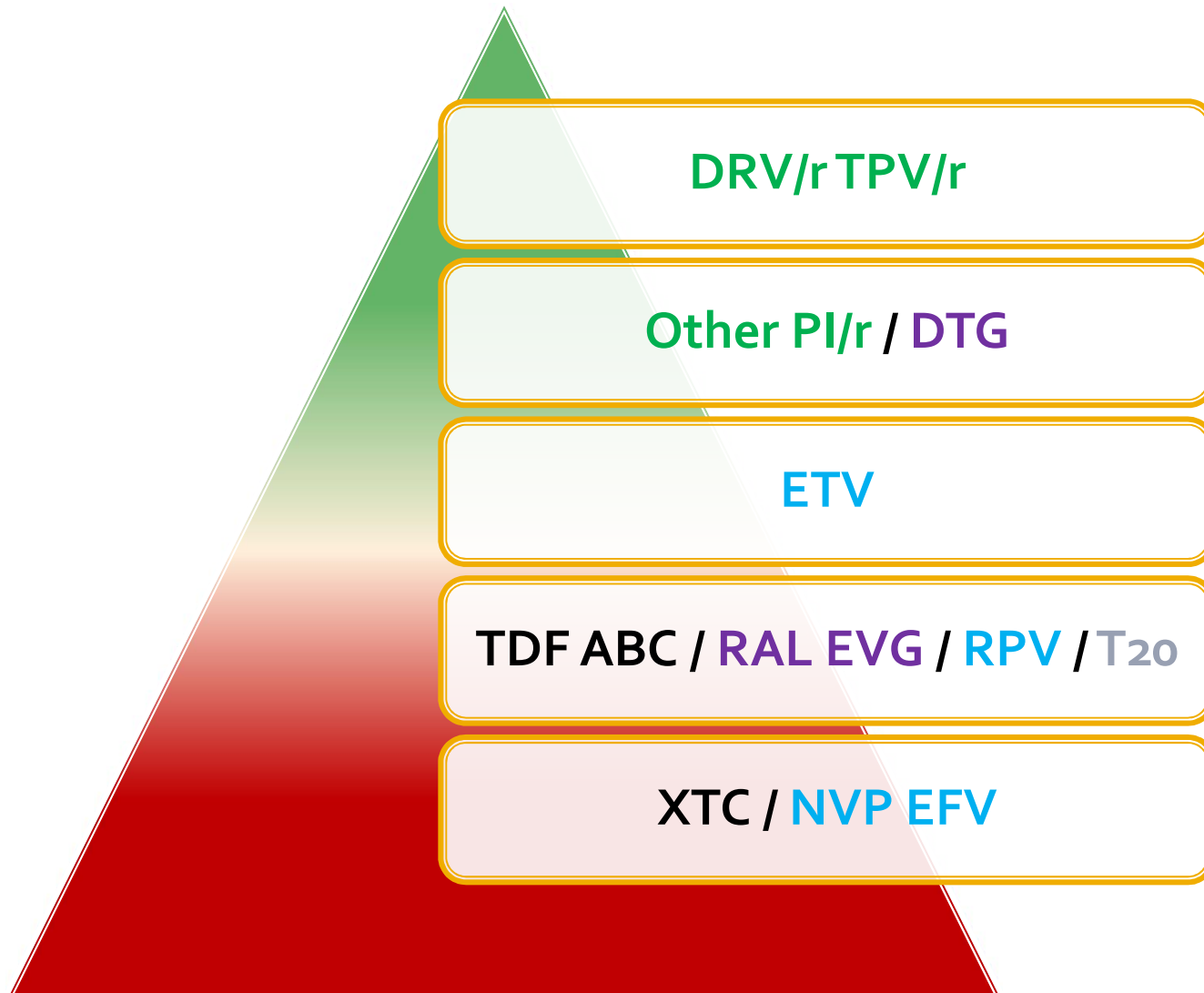
# How can we define the genetic barrier to resistance?



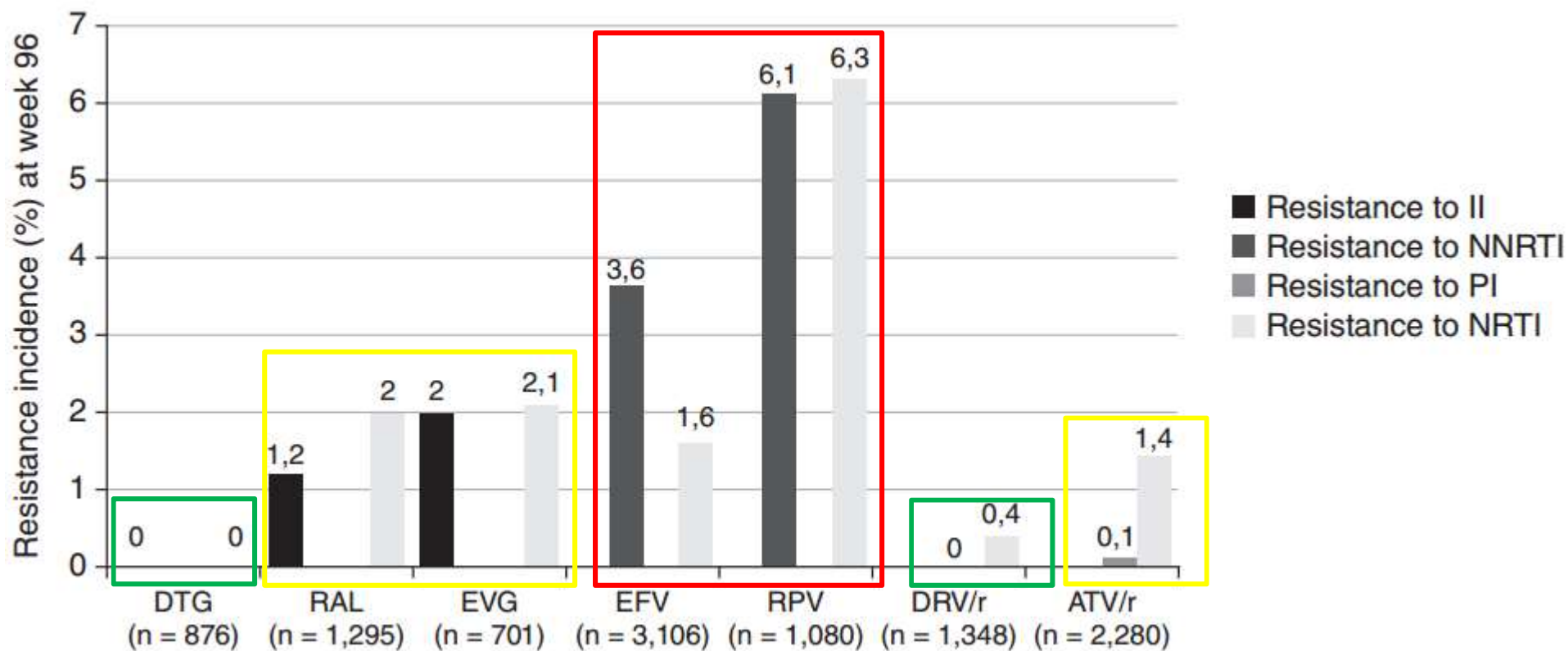
The extent of variation required for a susceptible virus to become resistant to a specific drug or drug combination



# Genetic barrier of individual antiretrovirals

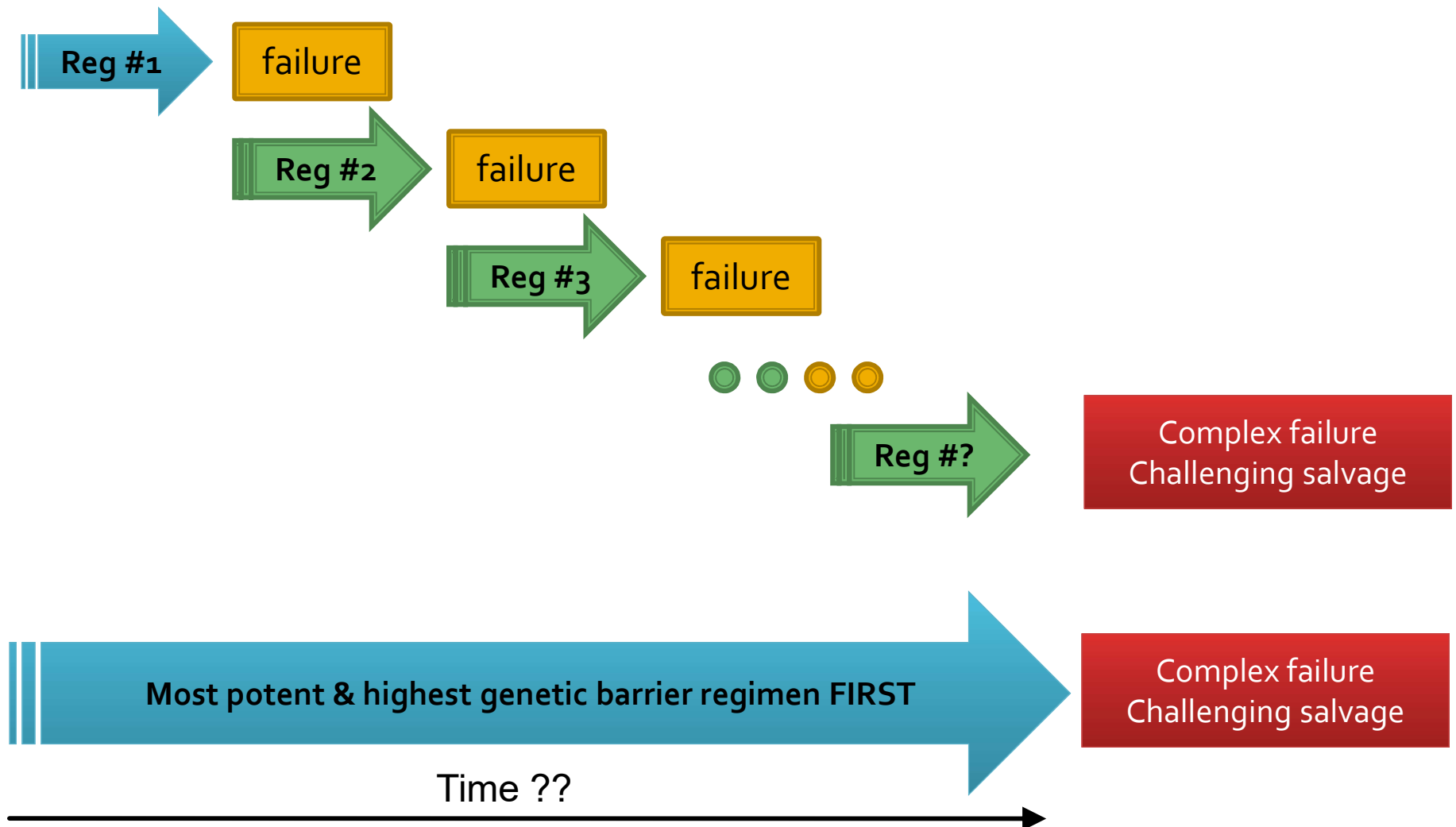


# Incidence of resistance at week 96 in pivotal first-line treatment trials

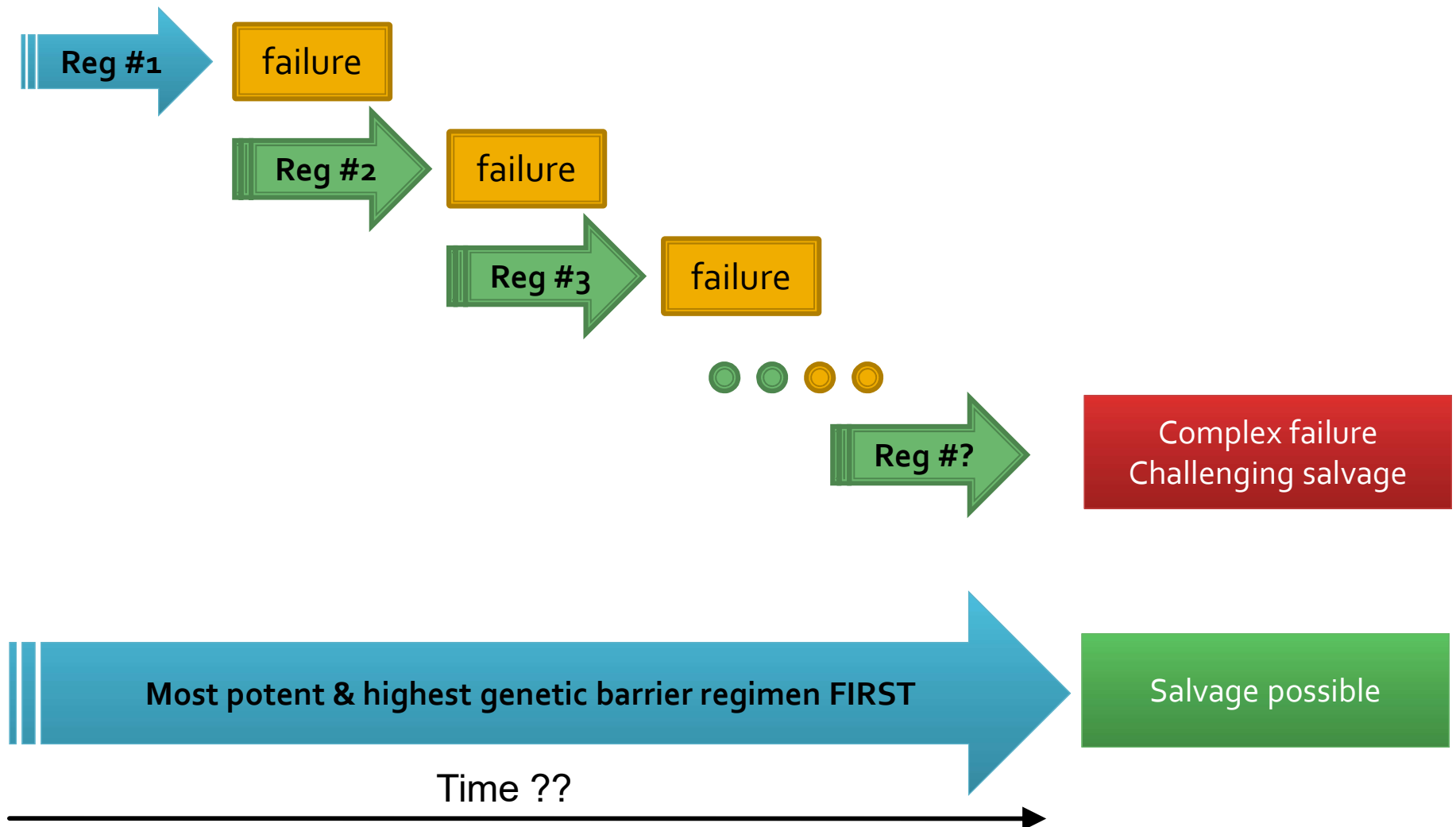


CAUTION: different definitions of virological failure, different proportions of advanced patients, different procedures to collect samples at failure.

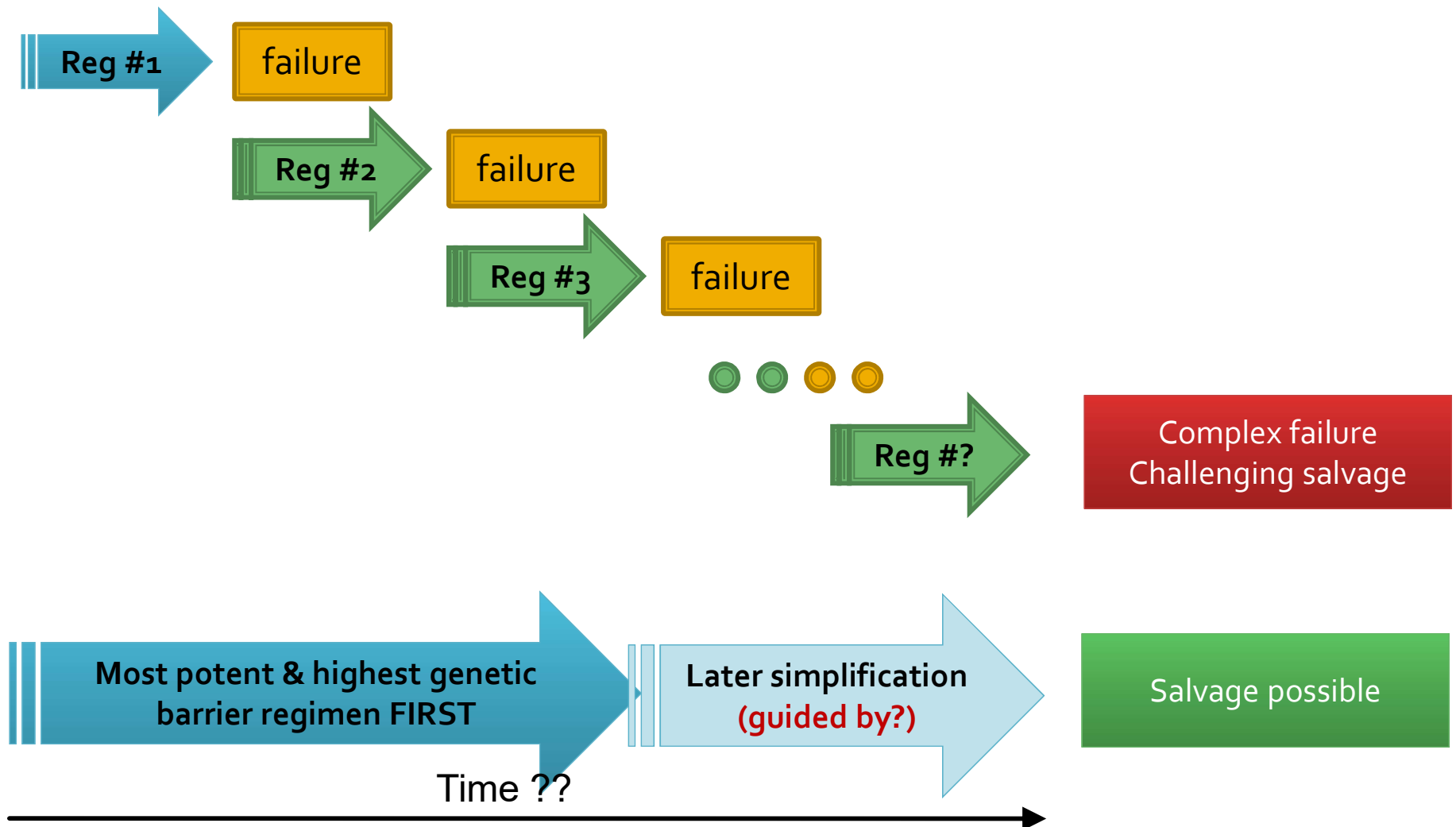
# Durability and sequencing of antiretroviral regimens



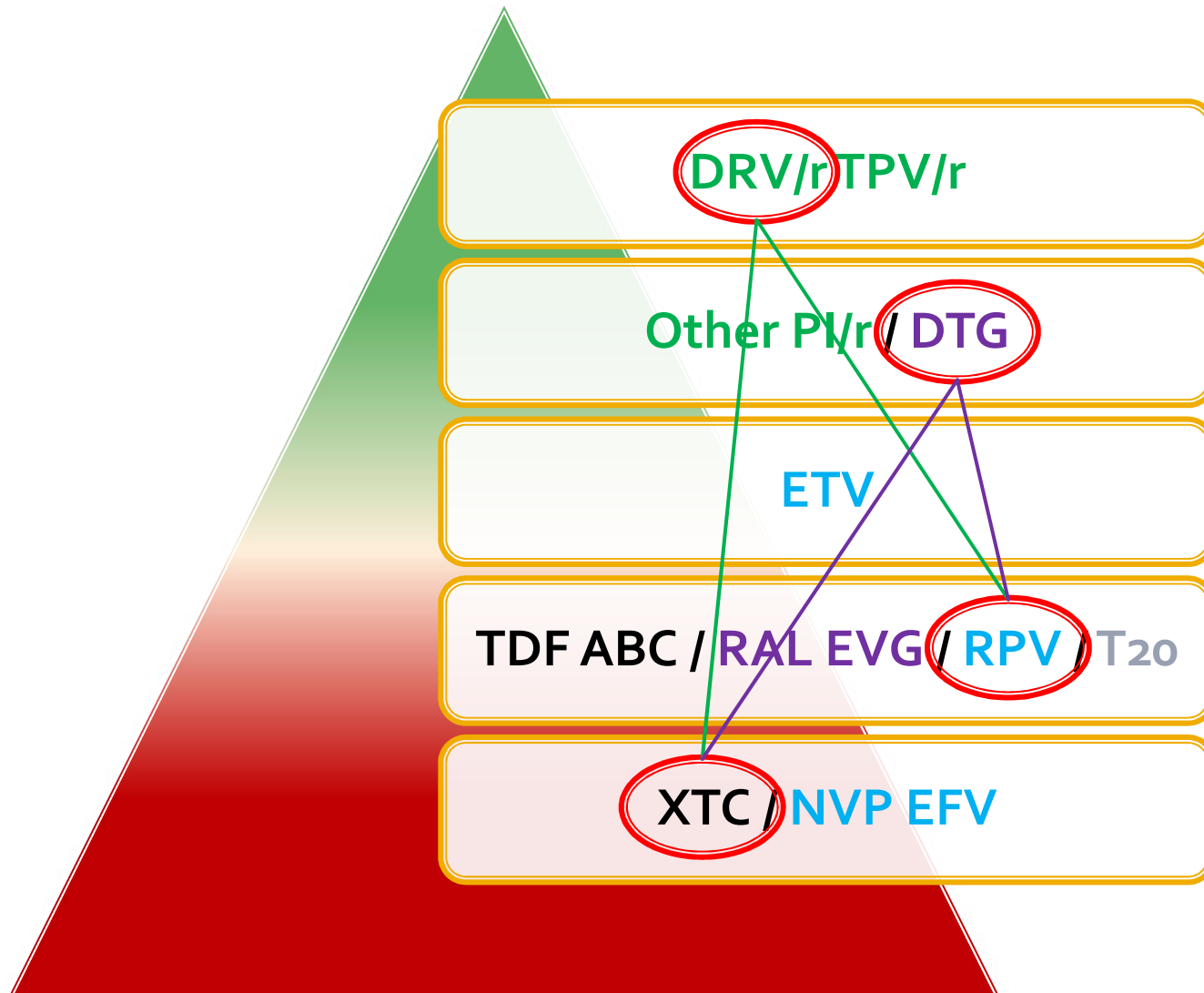
# Durability and sequencing of antiretroviral regimens



# Durability and sequencing of antiretroviral regimens



# Genetic barrier of individual antiretrovirals & how to build a robust 2-drug regimen



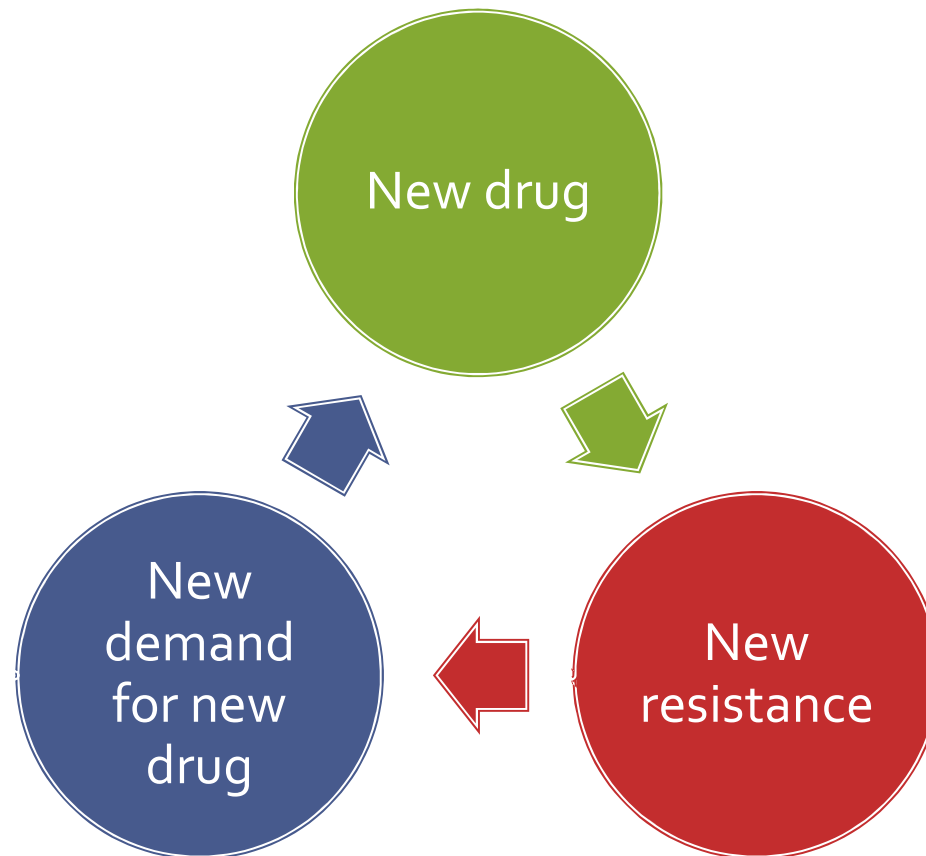
# Drug resistance testing as a key component of the success of antiretroviral therapy

Drug resistance  
testing urged  
development of  
novel drug classes





# Anti-HIV compounds & HIV resistance



# Cross-resistance within antiretroviral classes

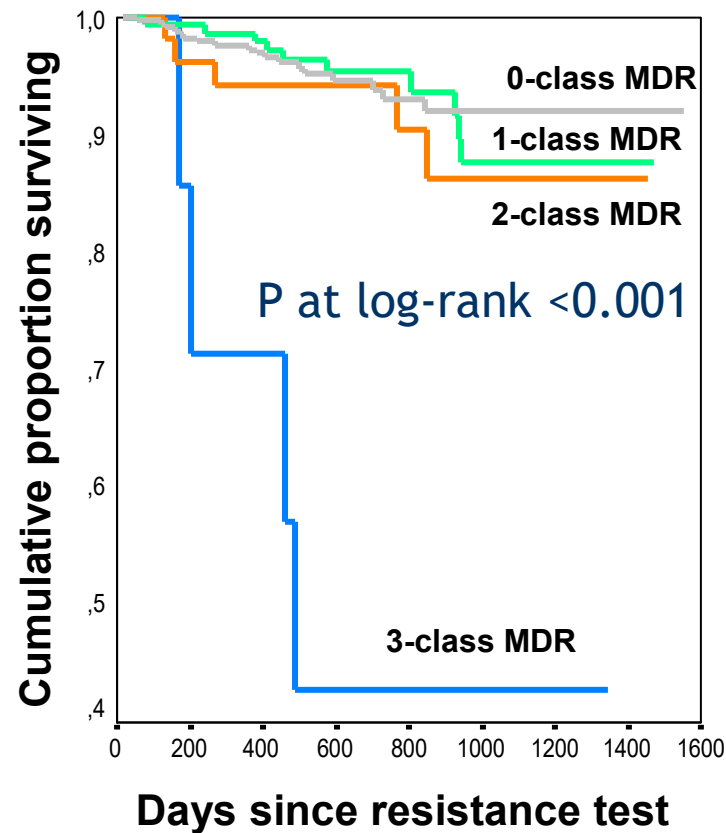
Major Nucleoside RT Inhibitor (NRTI) Resistance Mutations													
	Non-TAMs					TAMs						MDR	
	184	65	70	74	115	41	67	70	210	215	219	69	151
Cons	M	K	K	L	Y	M	D	K	L	T	K	T	Q
3TC	<u>VI</u>	R										Ins	M
FTC	<u>VI</u>	R										Ins	M
ABC	VI	<u>R</u>	E	<u>VI</u>	<u>F</u>	L			W	FY		<u>INS</u>	<u>M</u>
TDF	***	<u>R</u>	E		F	L		R	W	FY		<u>INS</u>	<u>M</u>
ZDV	***	***	*	*		L	N	R	W	FY	QE	<u>INS</u>	<u>M</u>

Major Non-Nucleoside RT Inhibitor (NNRTI) Resistance Mutations								
	100	101	103	106	181	188	190	230
Cons	L	K	K	V	Y	Y	G	M
NVP	I	<u>EP</u>	<u>NS</u>	<u>AM</u>	<u>CIV</u>	<u>LCH</u>	<u>ASEQ</u>	<u>L</u>
EFV	<u>I</u>	<u>EP</u>	<u>NS</u>	<u>AM</u>	CIV	<u>LCH</u>	<u>ASEQ</u>	<u>L</u>
ETR	<u>I</u>	<u>EP</u>			<u>CIV</u>	L	<u>ASEQ</u>	<u>L</u>
RPV	<u>I</u>	<u>EP</u>			<u>CIV</u>	<u>L</u>	<u>ASEQ</u>	<u>L</u>

Major Protease Inhibitor (PI) Resistance Mutations													
	24	32	46	47	48	50	54	76	82	84	88	90	
Cons	D	V	M	I	G	I	I	L	V	I	N	L	
ATV/r		I	IL	V	VM	<u>L</u>	VTALM		ATFS	<u>V</u>	<u>S</u>	<u>M</u>	
DRV/r		I		VA		V	LM	V	F	V			
LPV/r	I	I	IL	<u>VA</u>	VM	V	VTALM	V	<u>AFTS</u>	V			M

Major Integrase Inhibitor (INI) Resistance Mutations									
	66	92	138	140	143	147	148	155	263
Cons	T	E	E	G	Y	S	Q	N	R
RAL	<u>AIK</u>	Q	KAT	<u>SAC</u>	<u>RCH</u>		<u>HRK</u>	<u>H</u>	K
EVG	<u>AIK</u>	Q	KAT	<u>SAC</u>		<u>G</u>	<u>HRK</u>	<u>H</u>	K
DTG	K	Q	KAT	SAC			<u>HRK</u>	H	K

# Drug resistance impacts on HIV related mortality

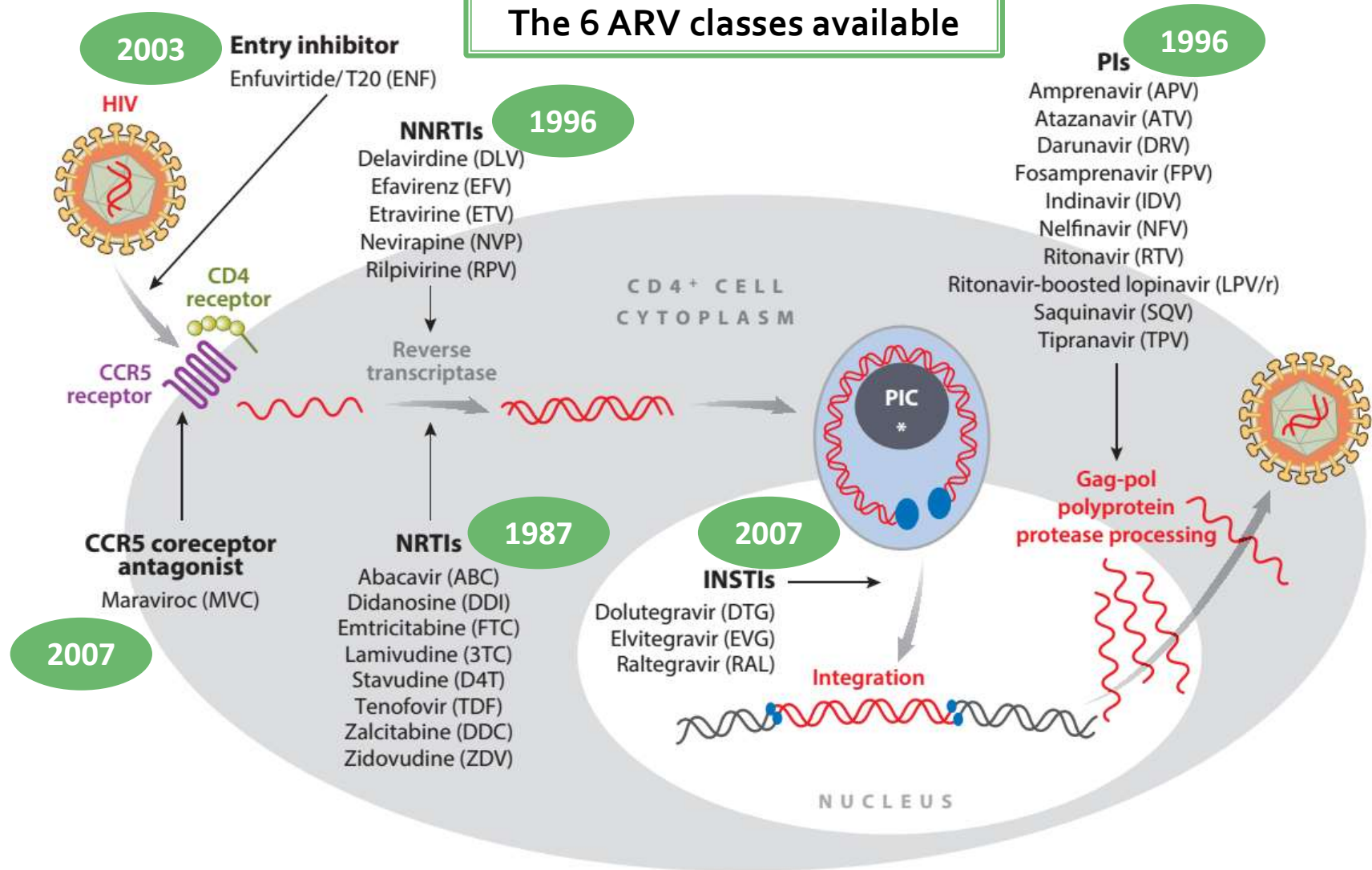


Multidrug resistance predicts mortality

(Zaccarelli, AIDS 2007)

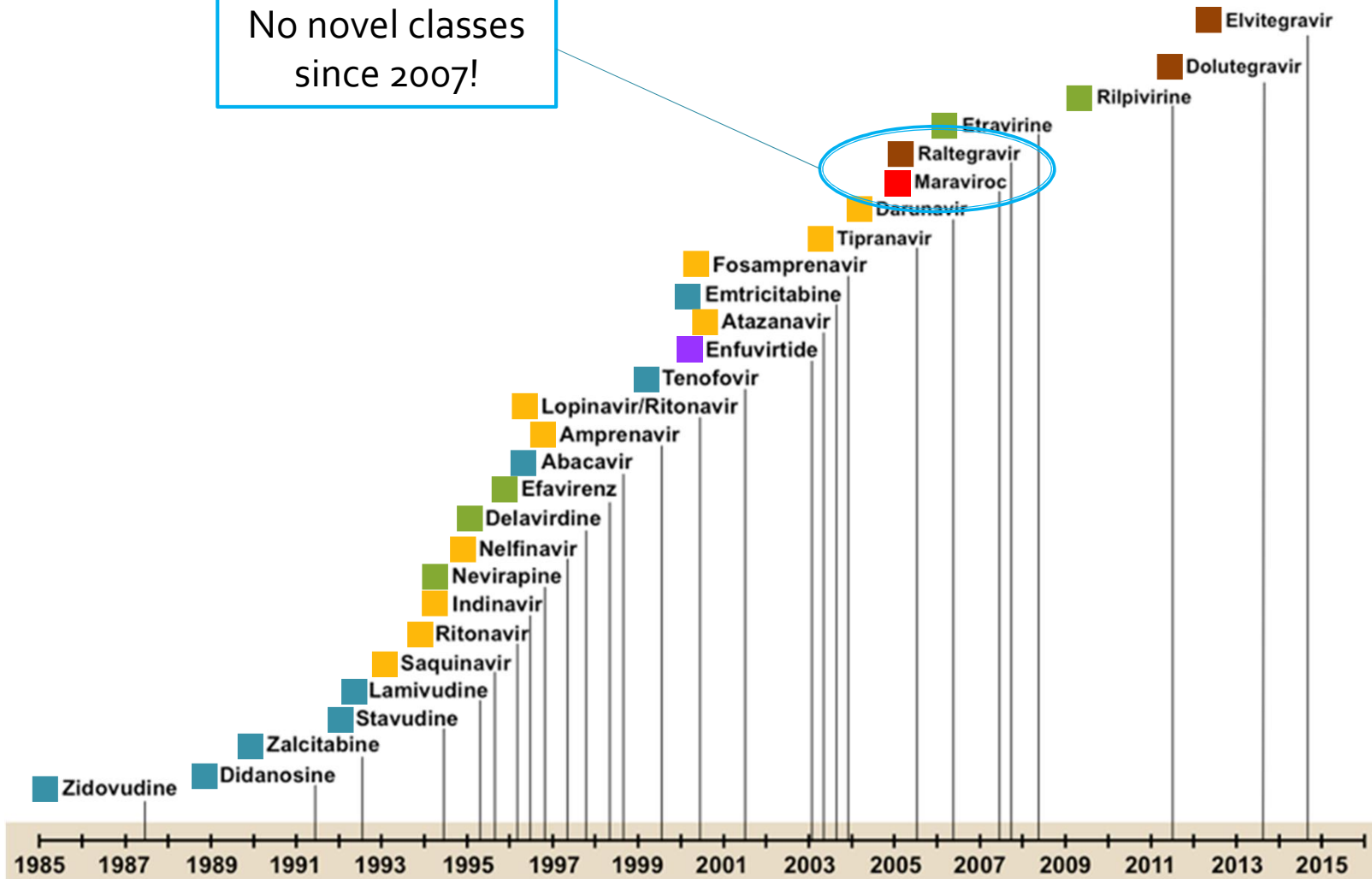
# Antiretroviral drug classes

## The 6 ARV classes available

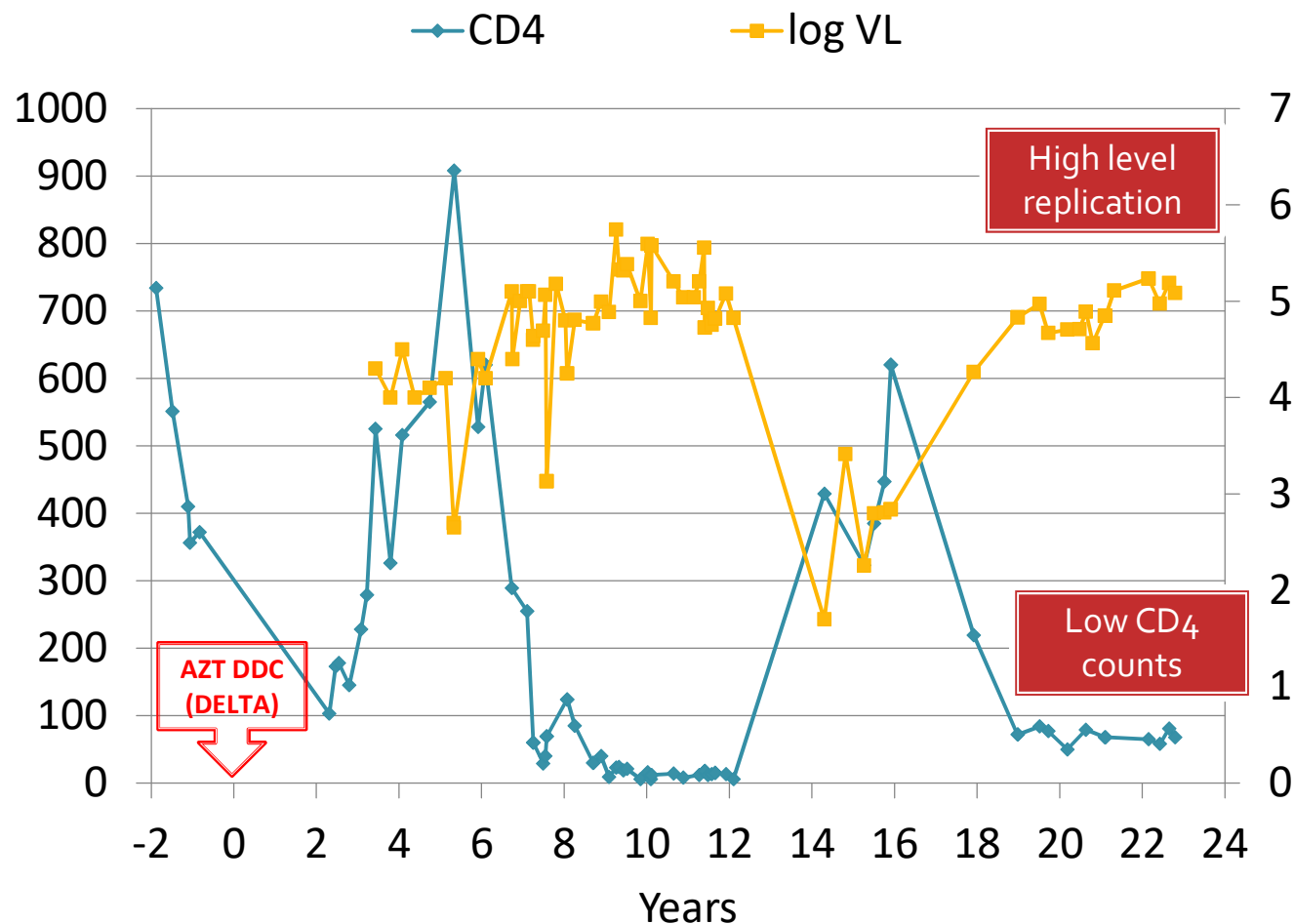


# HIV drugs timeline

No novel classes  
since 2007!



# Difficult-to-treat 6-class resistant viruses



- Born in 1961, HIV-1 diagnosis in 1990, IDU
- Therapy started in 1993, >20 treatment changes (including DELTA trial)
- Multiple treatment failures due to poor adherence
- 6-class experienced
- Extrapulmonary TB in 2008, anal cancer in 2013
- From 2012, VL~ $10^5$  cp/ml and CD4+ <100 cells/μl



# Difficult-to-treat 6-class resistant viruses

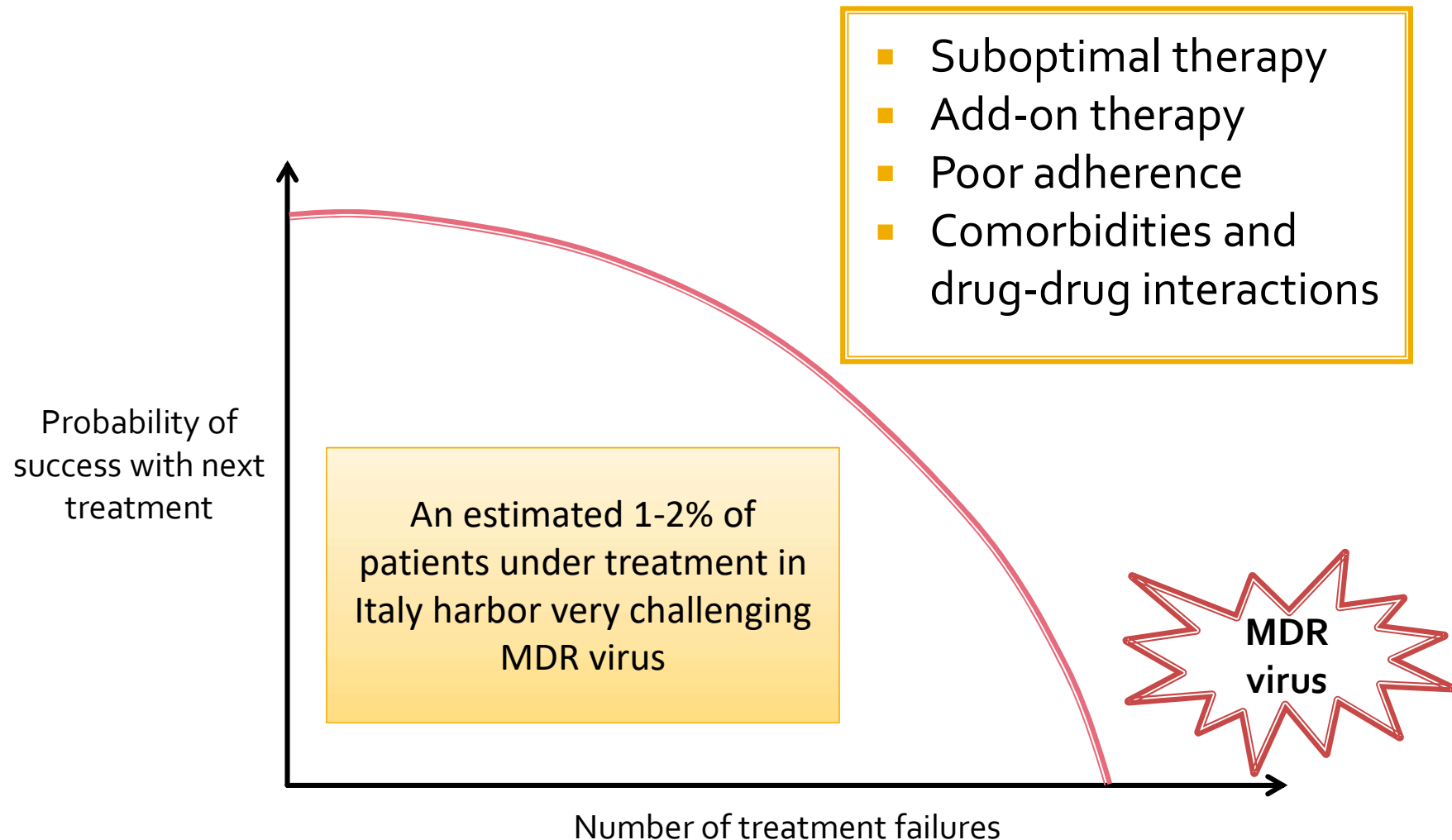
NRTI							NNRTI				PI								INI			FI	CA	
3TC	ABC	AZT	D4T	DDI	FTC	TDF	EFV	ETR	NVP	RPV	ATV	ATV /rtv	DRV /rtv	FPV /rtv	IDV /rtv	LPV /rtv	NFV	SQV /rtv	TPV /rtv	DTG	EVG	RAL	T20	MVC
0	0	0,25	0	0	0	0,5	0	0,6	0	0,3	0	0	0	0	0	0	0	0	0	0,5	0,25	0	0,25	0

<b>Mutazioni PR:</b>	10I 10V 11I 13V 20I 20T 32I 33F 36I 43T 46I 54L 54V 60E 63P 71I 77I 82A 82I 82T 84V 89V 90M 93L
<b>Altre mutazioni PR:</b>	12P 12S 15V 16R 19P 37D 37E 41K 55N 62V 72R 73D 79A 79S 91S 92K 94D
<b>Mutazioni RT:</b>	66d 67E 69G 70R 74I 103N 108I 118I 179I 181C 184V 215F 219E 228H
<b>Altre mutazioni RT:</b>	37F 39K 40A 40T 43Q 47F 67K 69R 98S 100F 111L 122E 135T 165A 177N 201I 223Q 245E
<b>Mutazioni IN:</b>	140S 148H
<b>Altre mutazioni IN:</b>	49S 49T 61L 101I 116G
<b>Mutazioni GP41:</b>	43D
<b>Altre mutazioni GP41:</b>	7L 23A 24L 32L

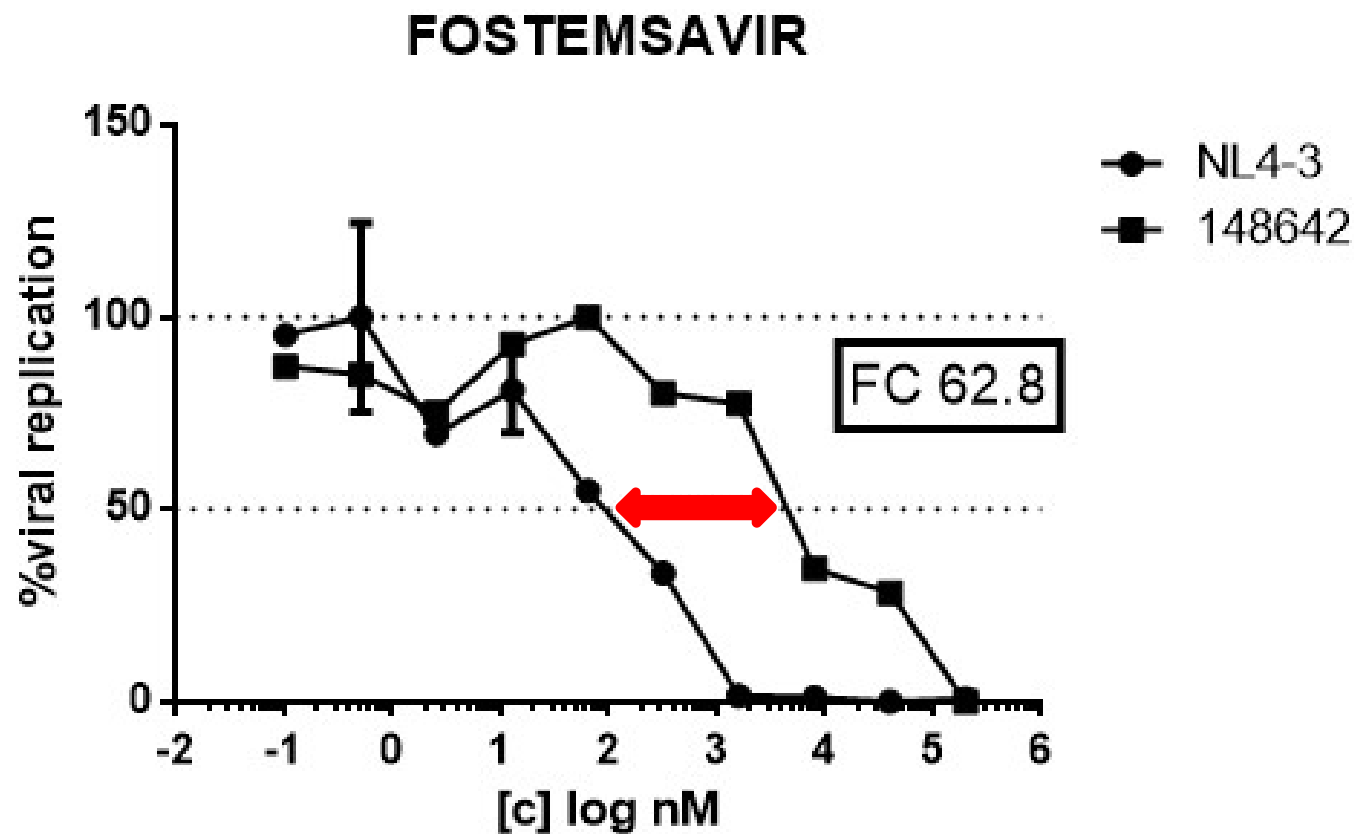
- Cumulative (1999-2017) genotype based on
  - 18 PR/RT sequences (note that 33 of 99 PR aminoacids have changed!)
  - 6 IN sequences
  - 2 gp41 sequences
  - 4 V3 sequences



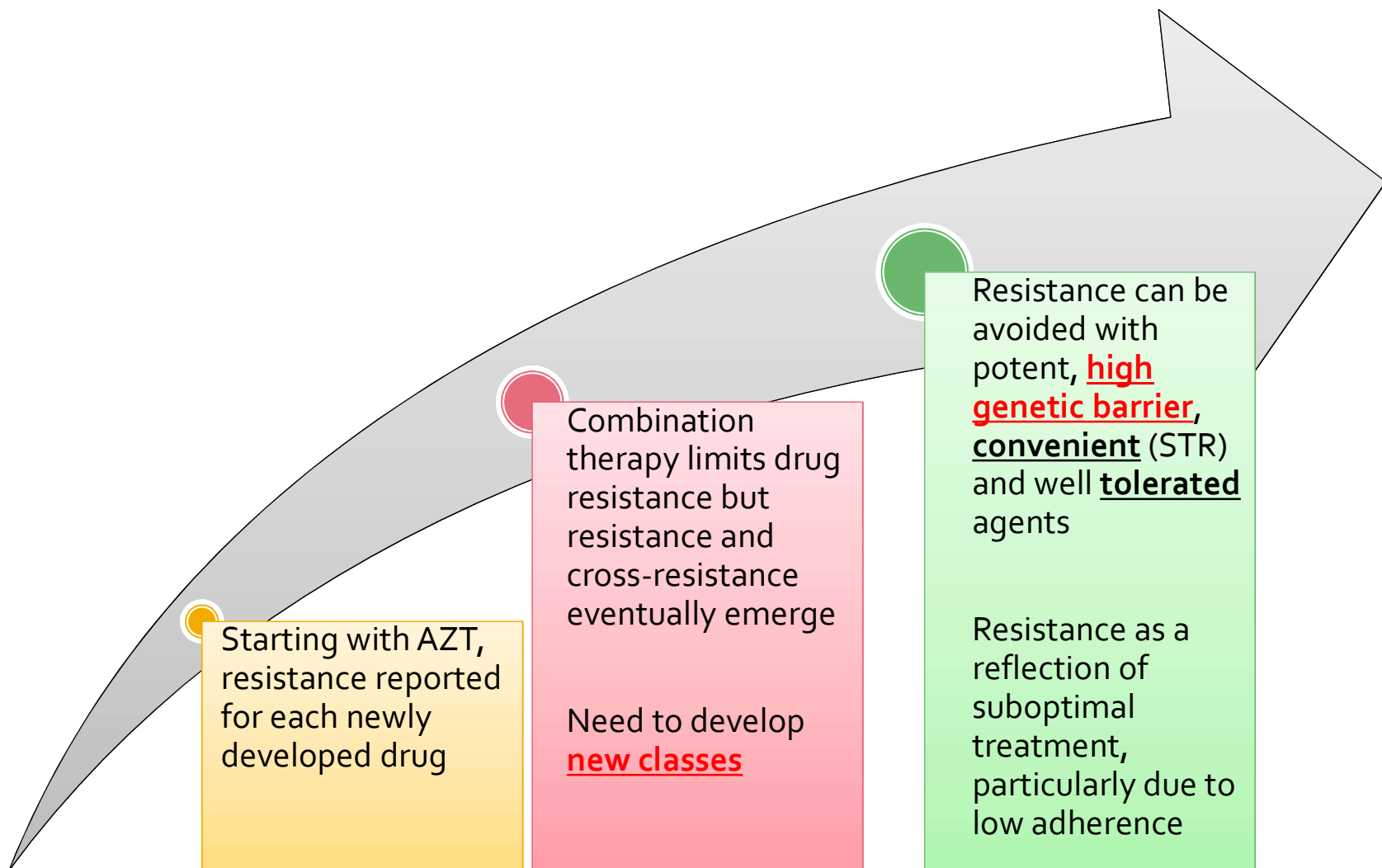
# How MDR builds up



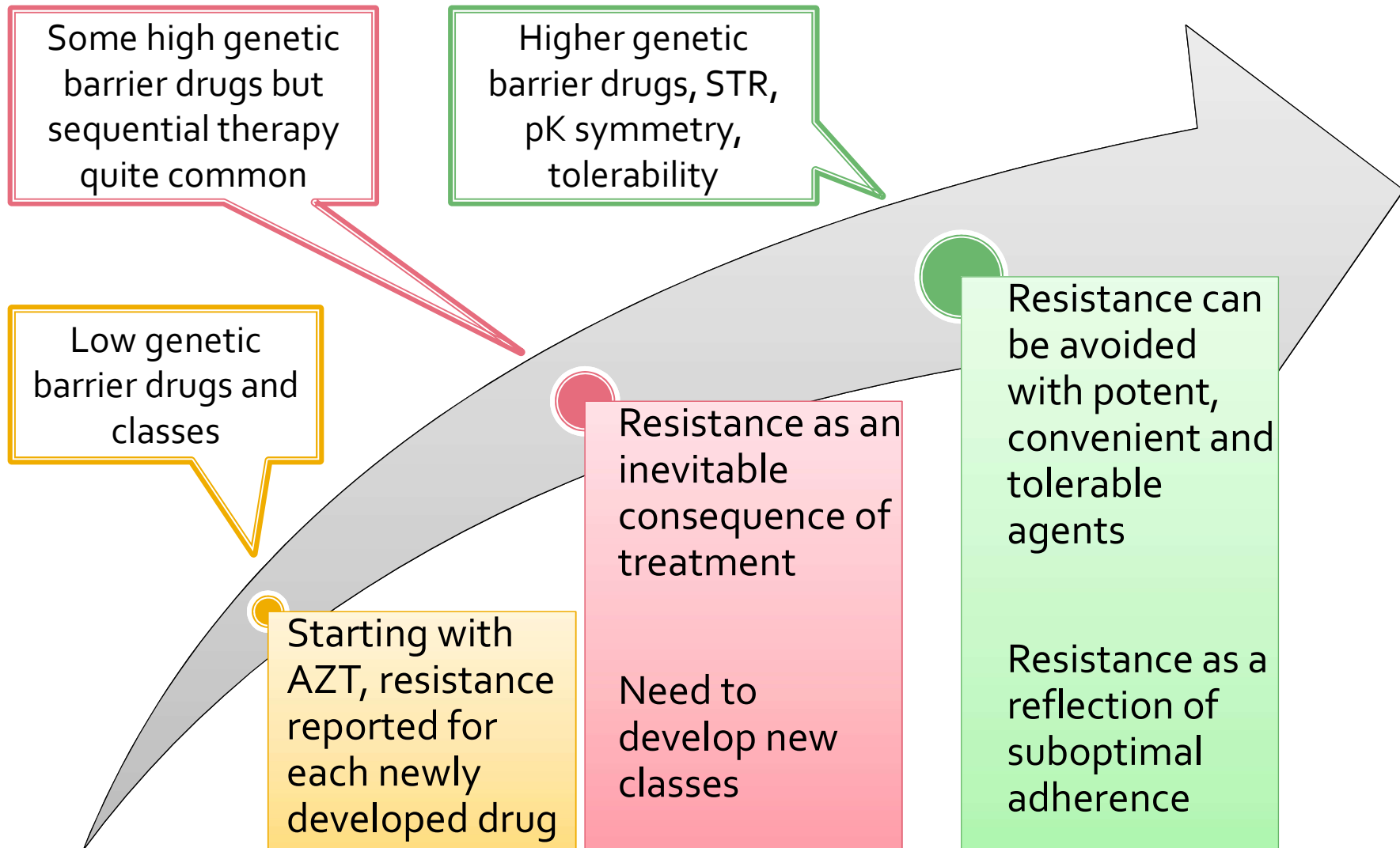
# Difficult-to-treat 7-class resistant virus



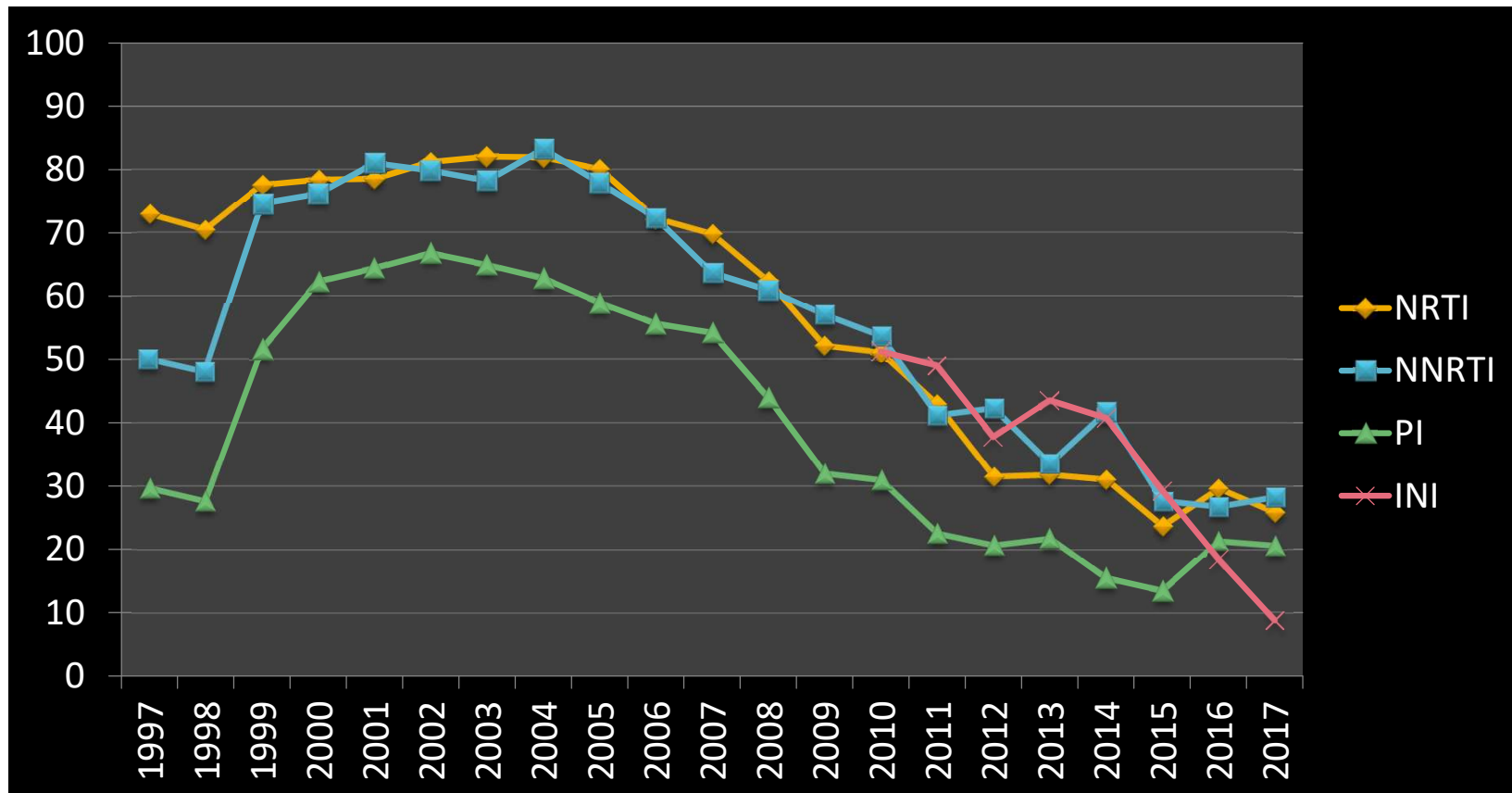
# Evolution of HIV treatment and drug resistance over antiretroviral therapy eras



# Evolution of HIV treatment and drug resistance over time

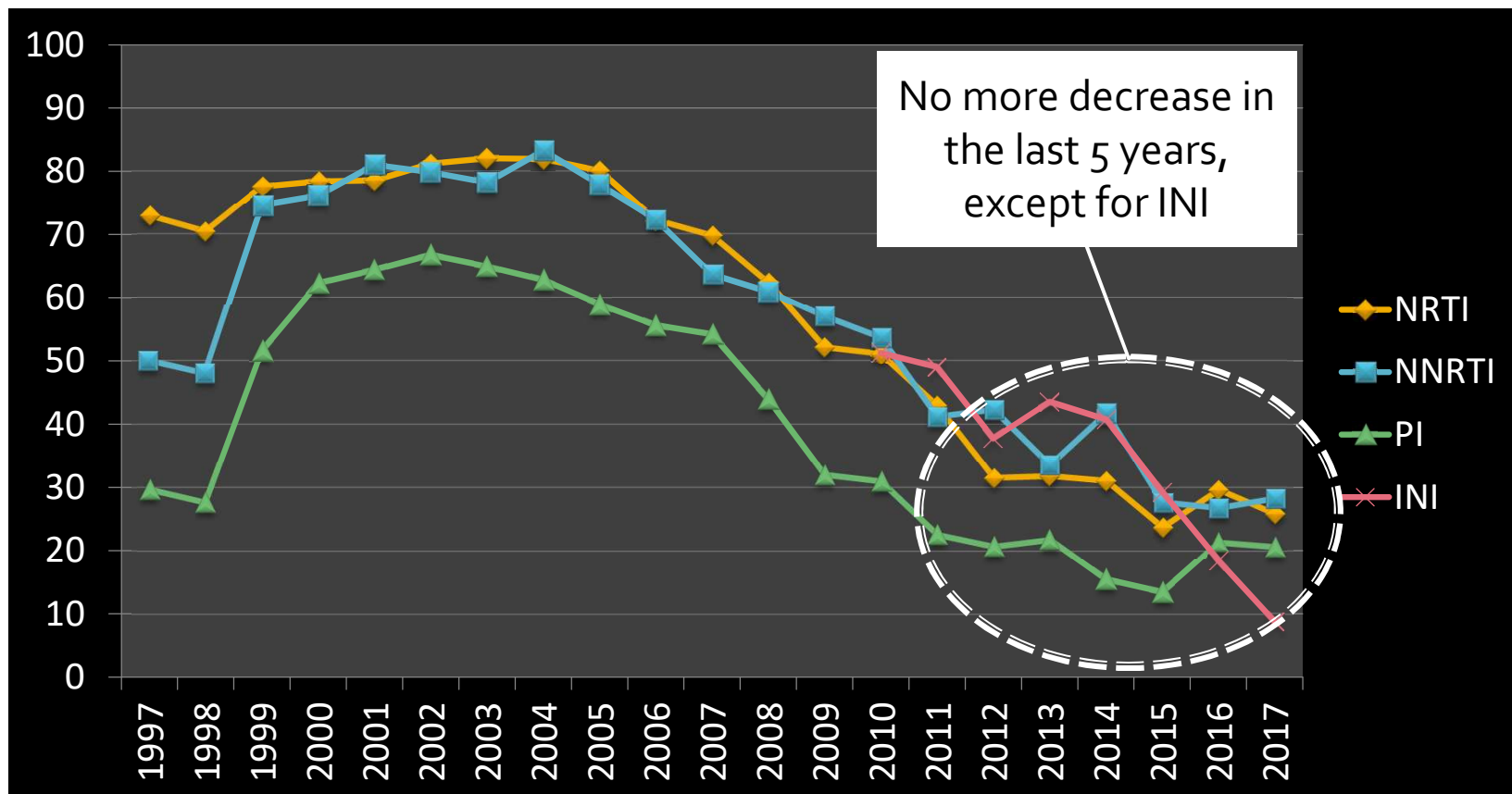


# Time trends of emergent resistance at failure with the corresponding drug class in Italy



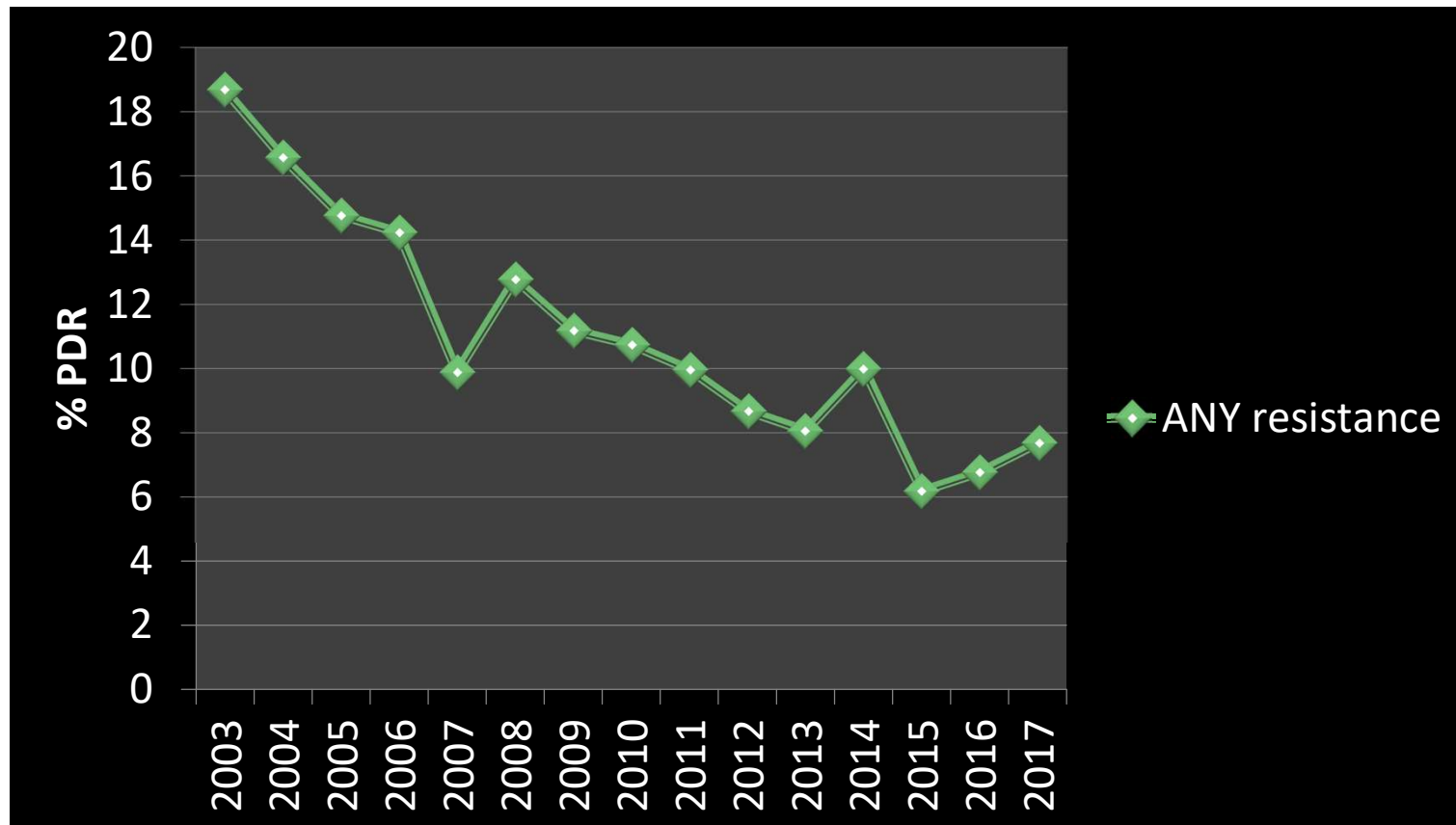
ARCA, accessed October 10, 2017; based on 14,947 sequences from patients on NRTI, 4,357 on NNRTI, 9,619 on PI, 565 on INI

# Time trends of emergent resistance at failure with the corresponding drug class in Italy



ARCA, accessed October 10, 2017; based on 14,947 sequences from patients on NRTI, 4,357 on NNRTI, 9,619 on PI, 565 on INI

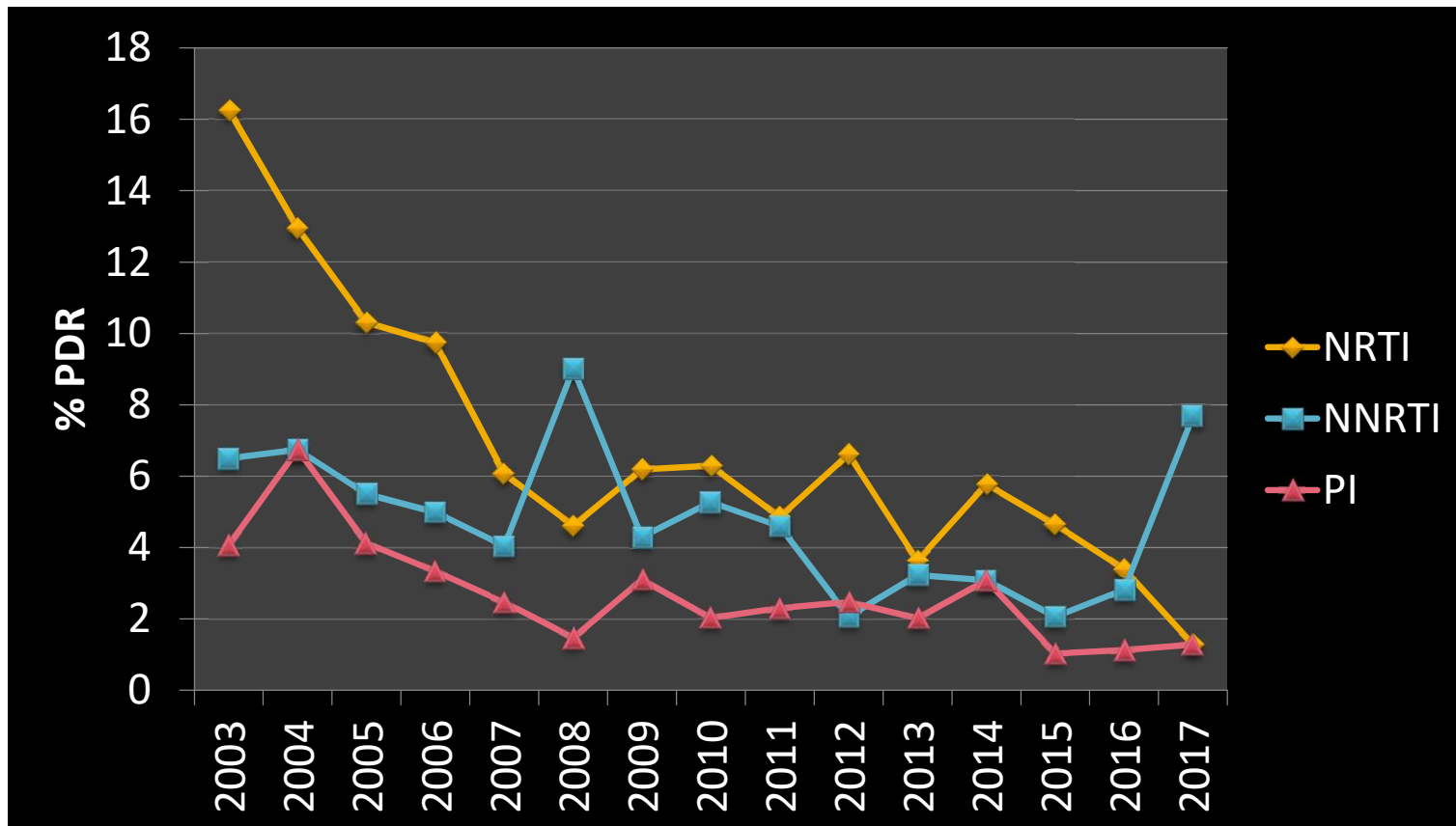
# HIV pre-treatment drug resistance in Italy



Based on 4453 PR/RT sequences from drug naïve individuals

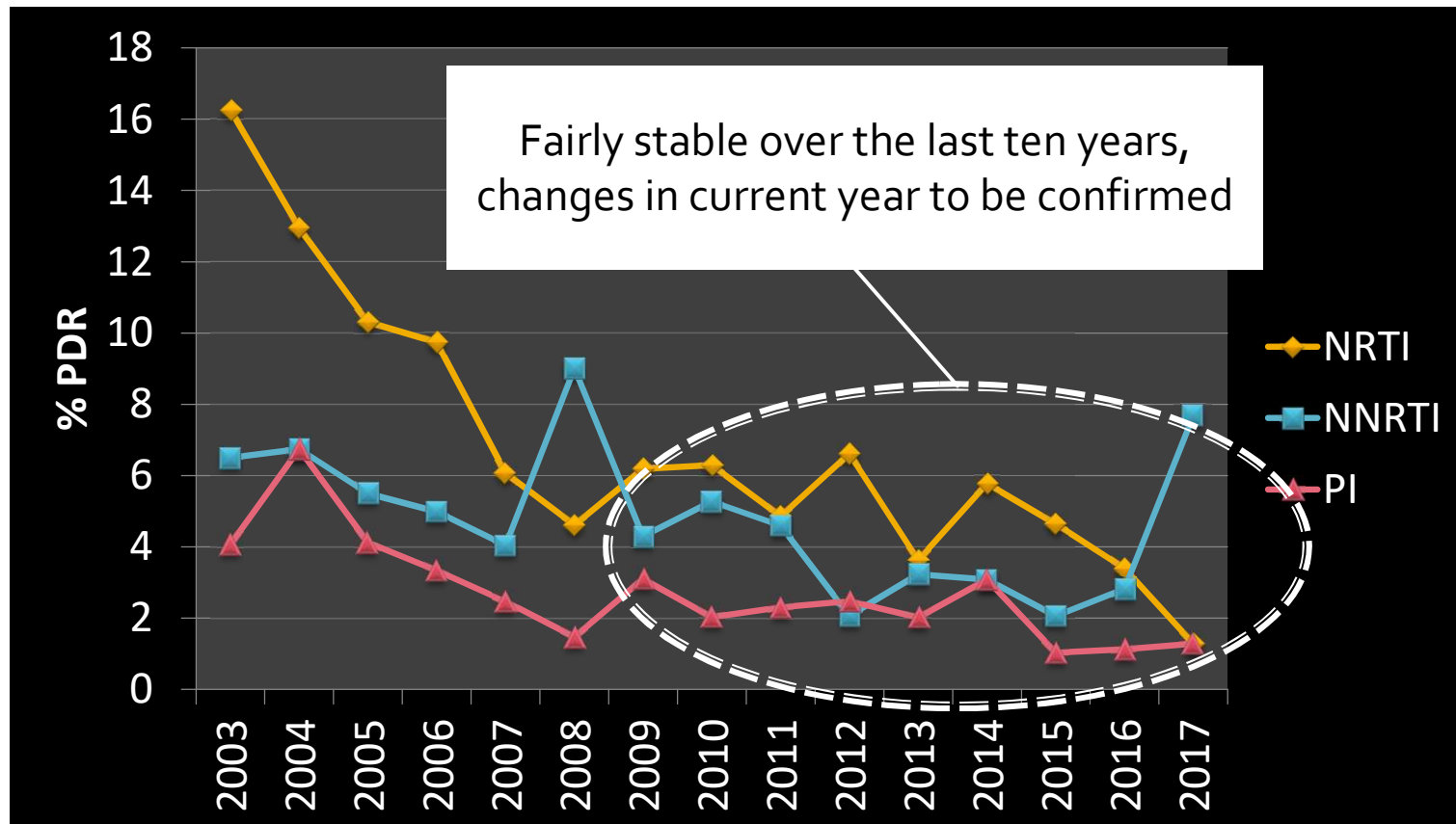


# HIV pre-treatment drug resistance in Italy



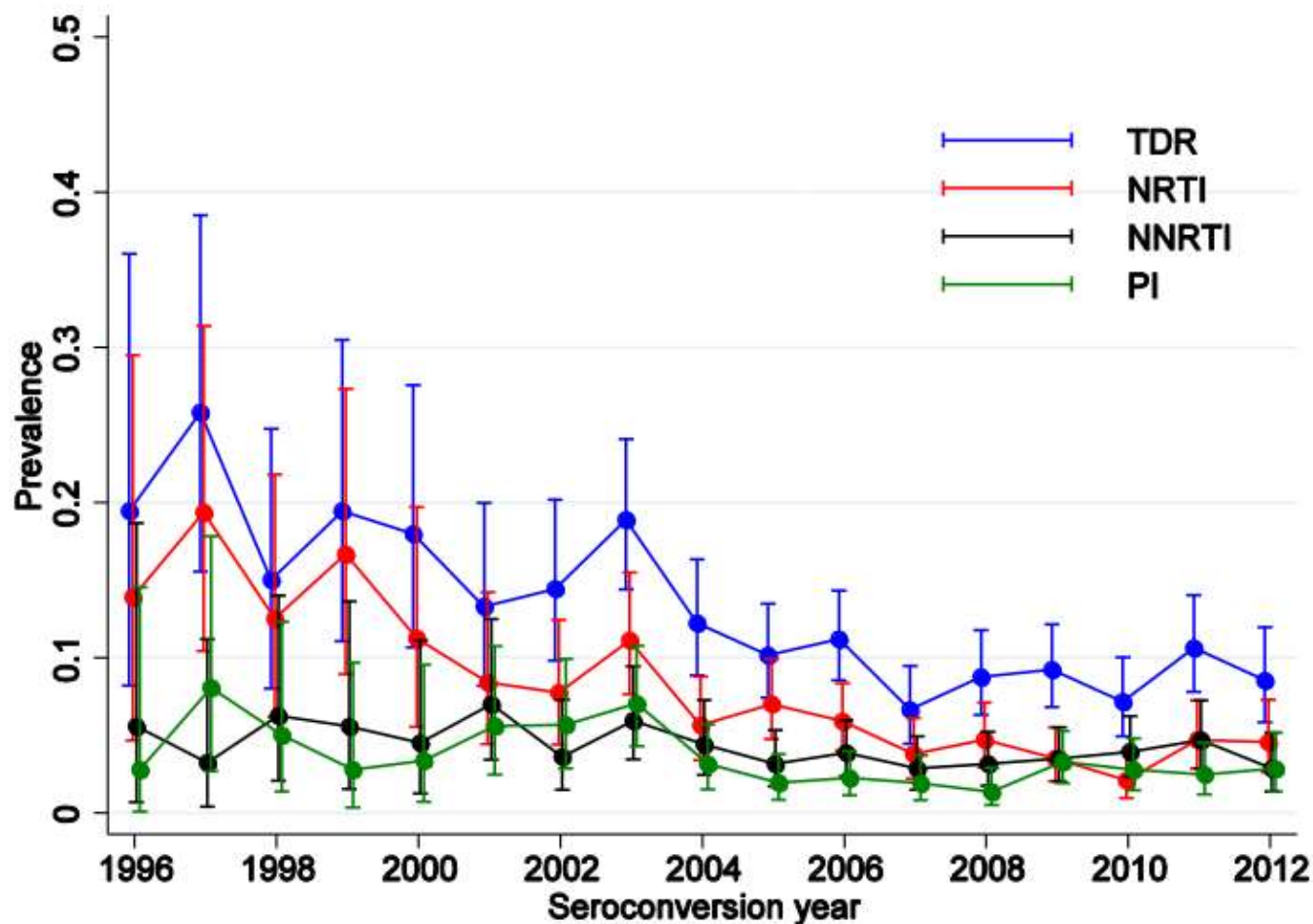
Based on 4453 PR/RT sequences from drug naïve individuals

# HIV pre-treatment drug resistance in Italy



Based on 4453 PR/RT sequences from drug naïve individuals

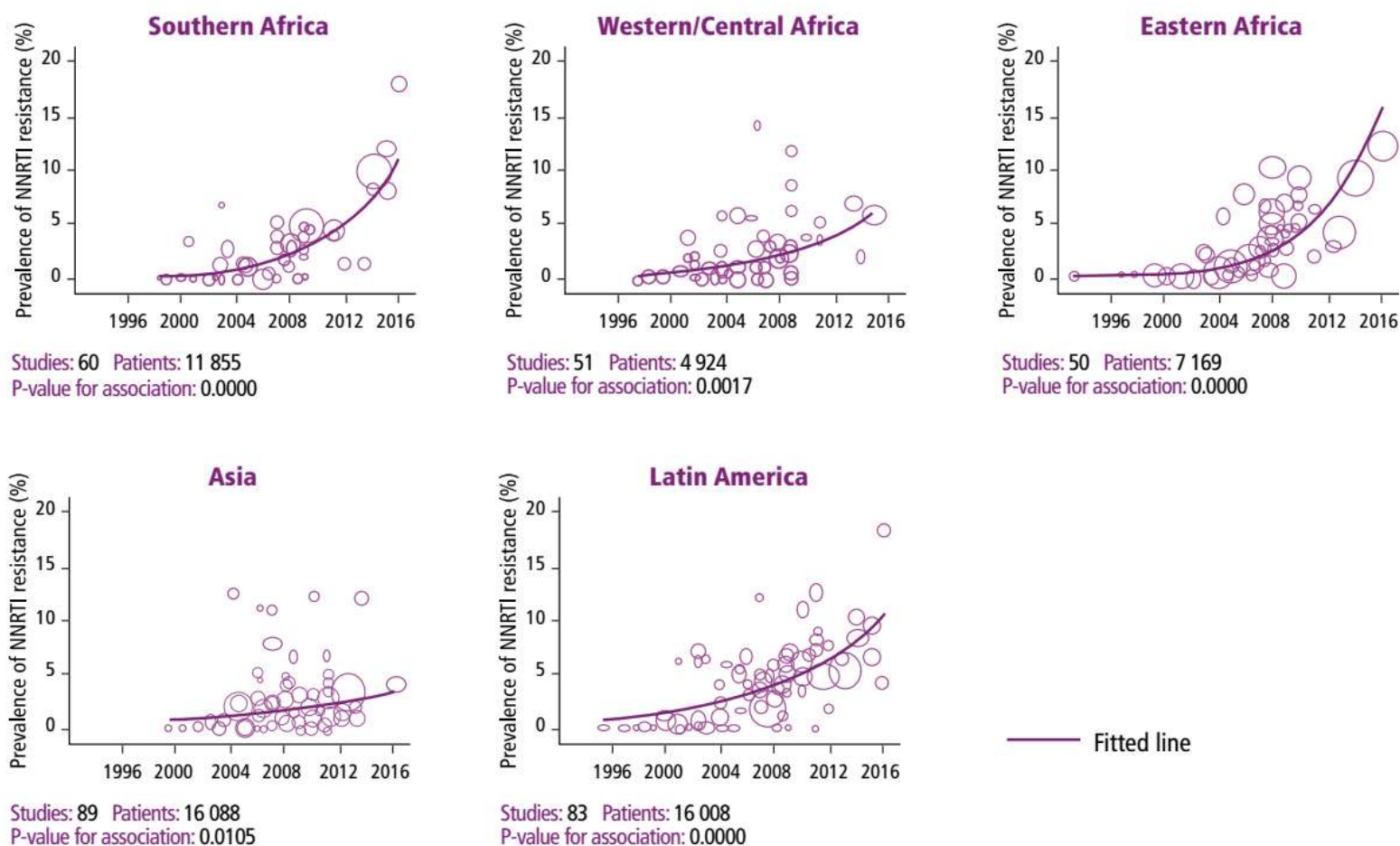
# HIV pre-treatment drug resistance in European seroconverters (CASCADE cohort)



- 4717 individuals seroconverting between 1996-2012
- MSM 80%
- Subtype B 73%
- TDR prevalence significantly decreasing from 19.4% in 1996 to 8.5% in 2012

# HIV PDR in low/middle income countries

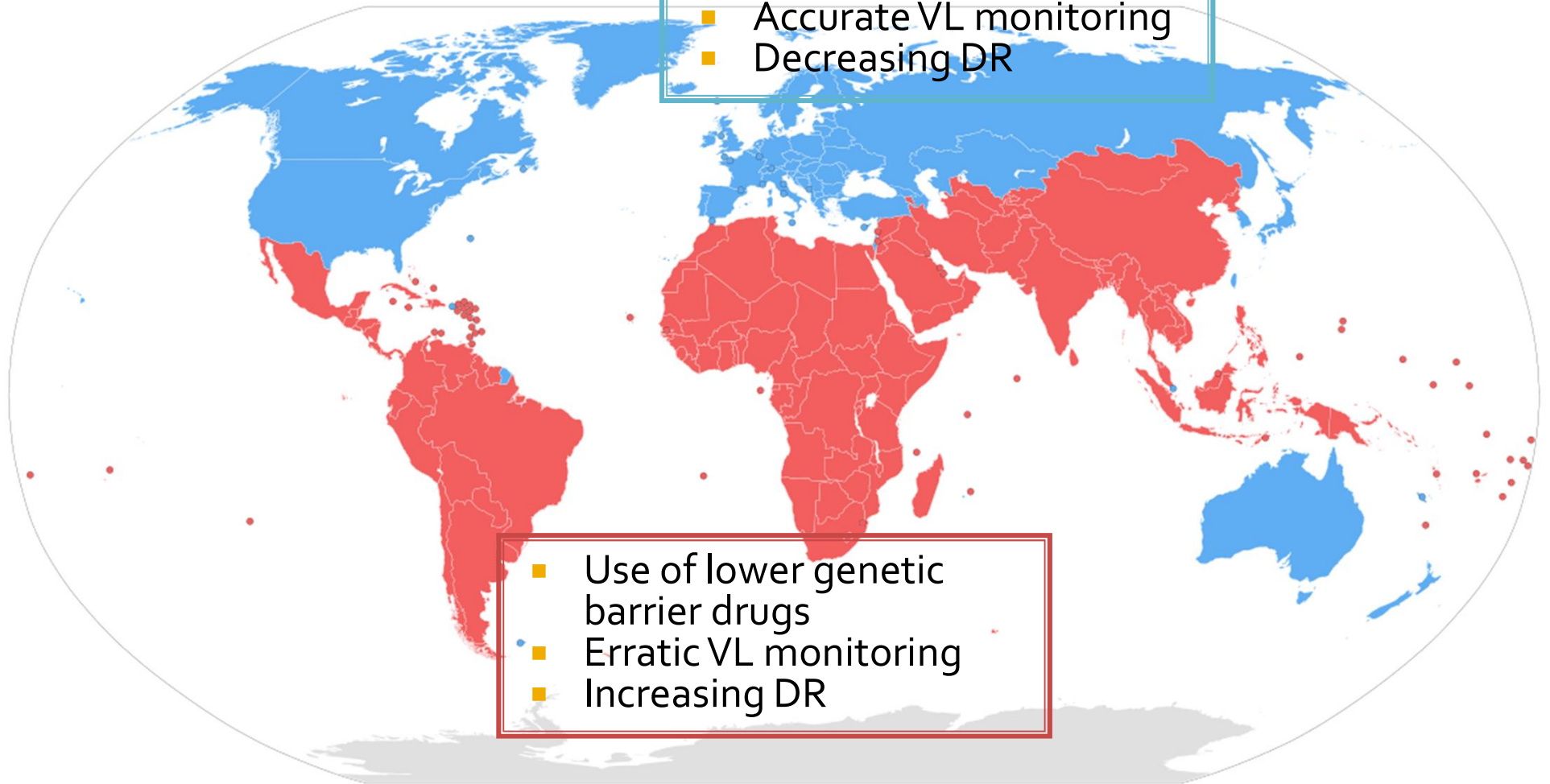
Fig. 7: Prevalence of NNRTI pretreatment resistance by calendar year across studies included in the systematic review



# HIV import from migrants and implications on PDR

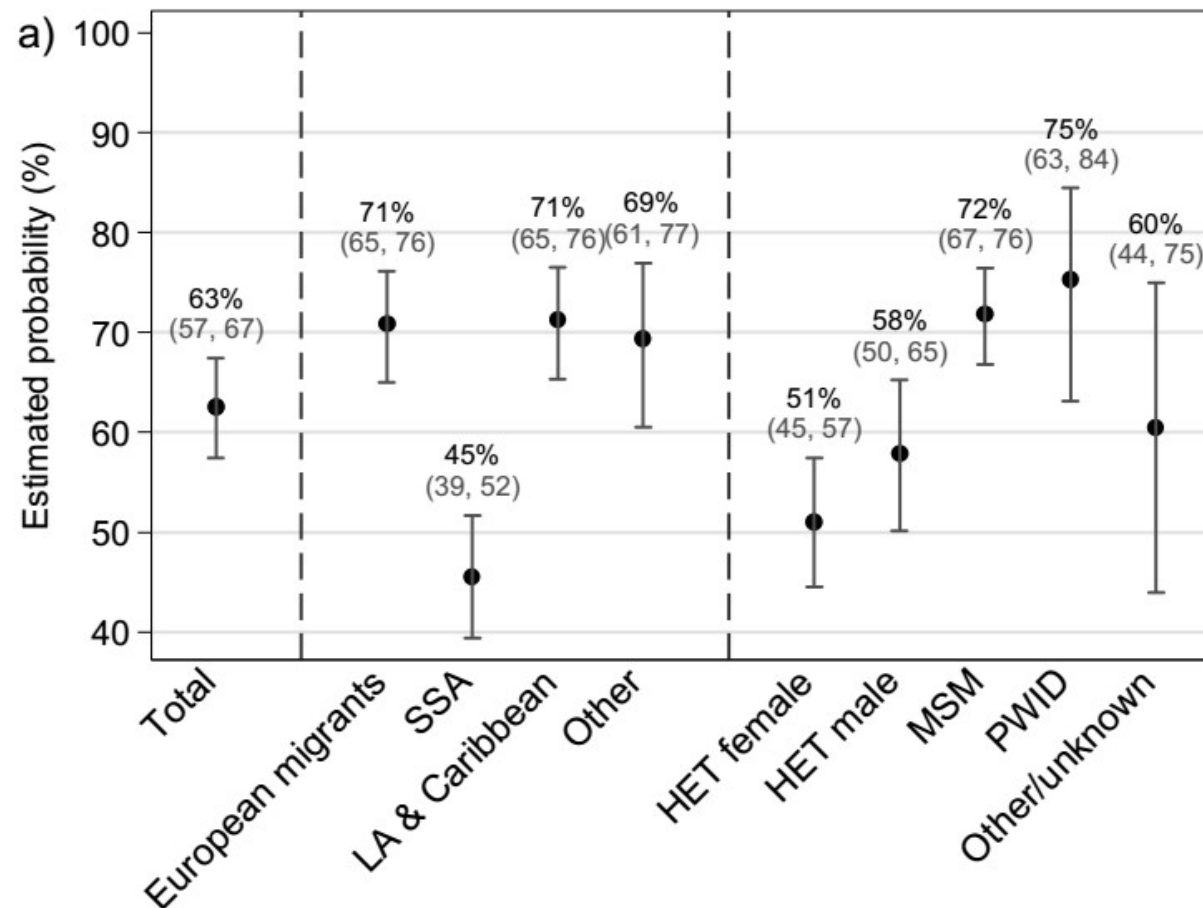
- Use of higher genetic barrier drugs
- Accurate VL monitoring
- Decreasing DR

- Use of lower genetic barrier drugs
- Erratic VL monitoring
- Increasing DR



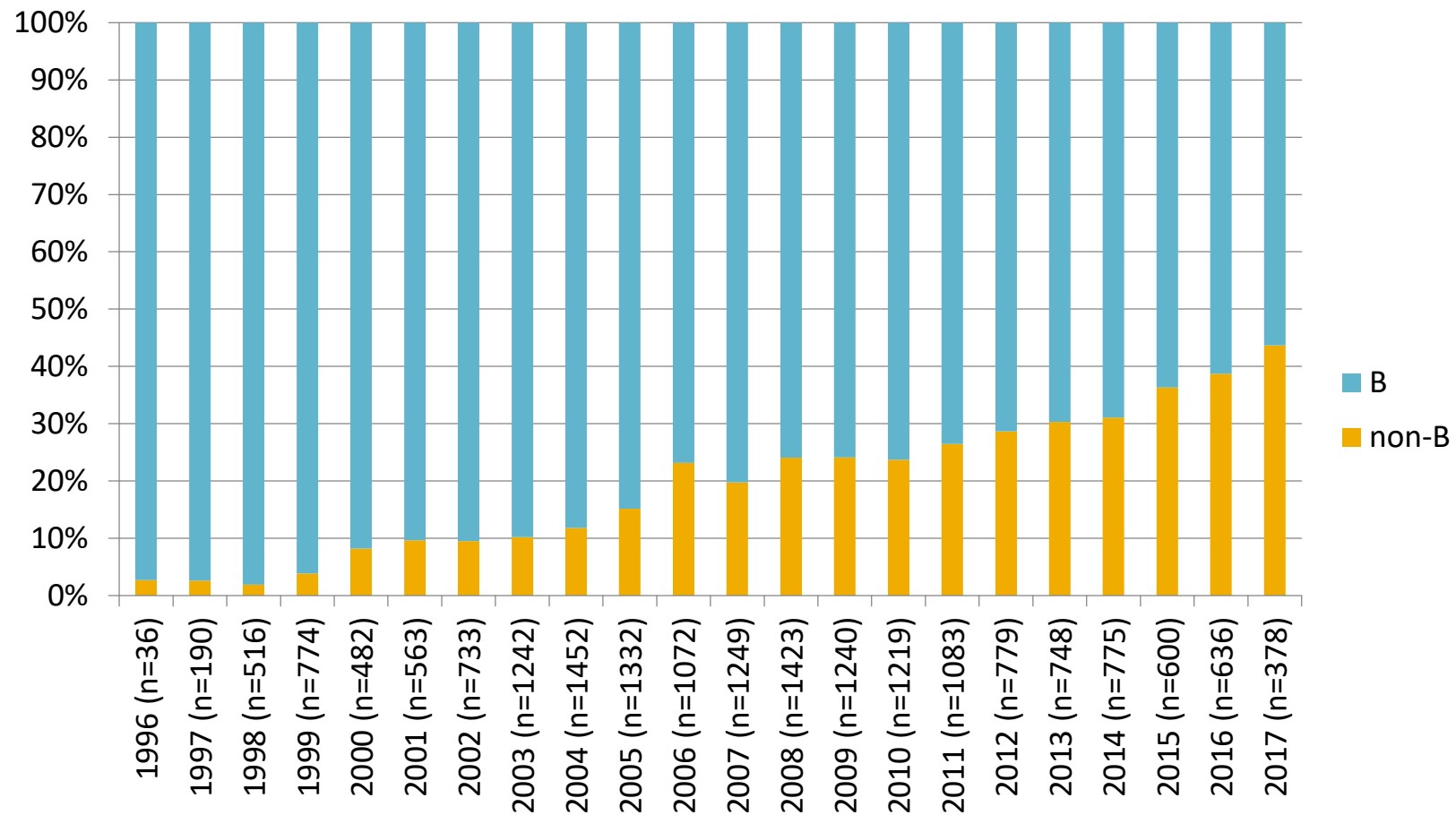


# HIV acquisition in migrants



Estimated post-migration HIV acquisition probability (95% CI) by a) mode of transmission and geographical origin

# Time trends of B and non-B HIV subtypes in Italy

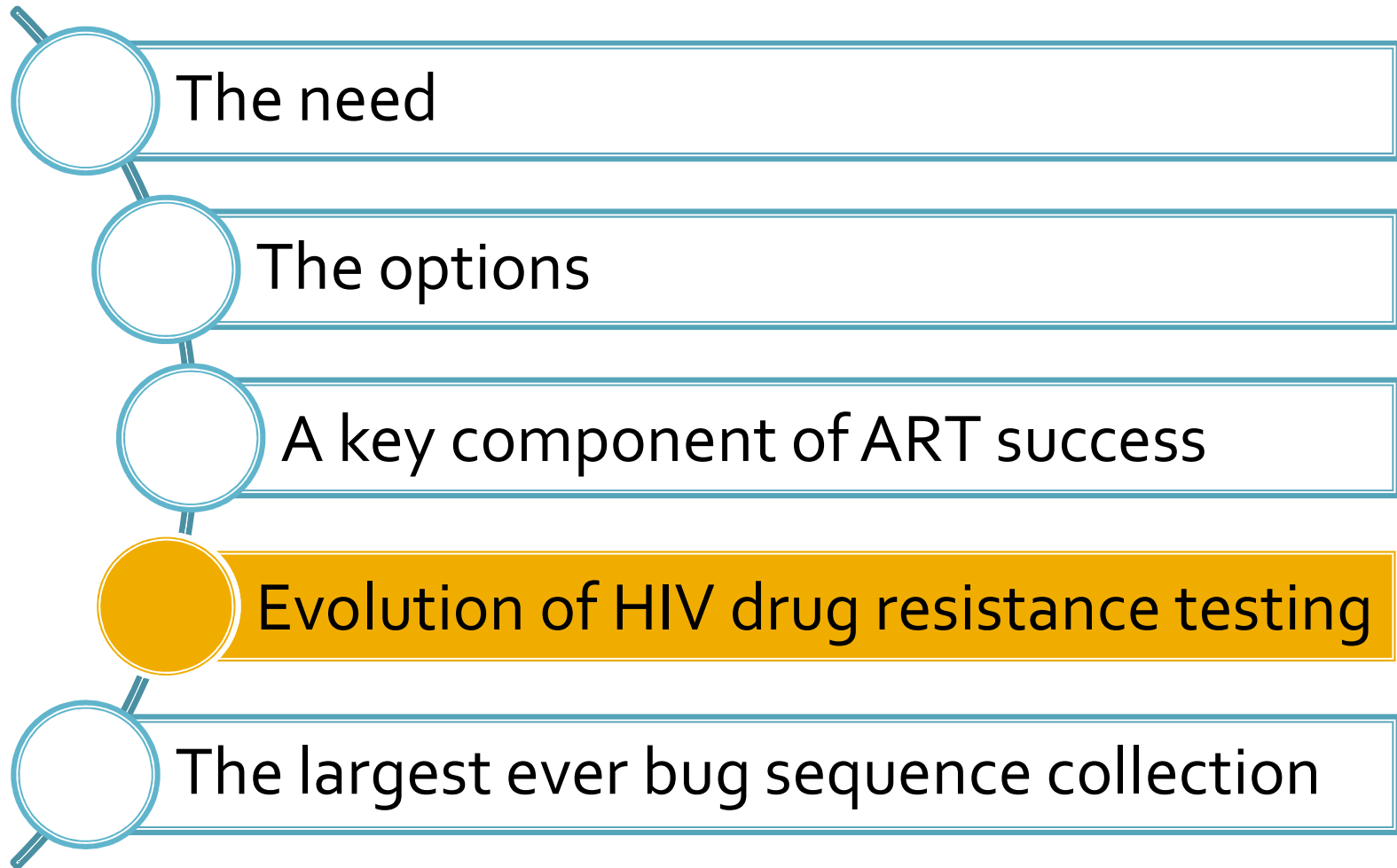


ARCA, accessed October 10, 2017; based on first sequence available per individual patient (n=23,588)

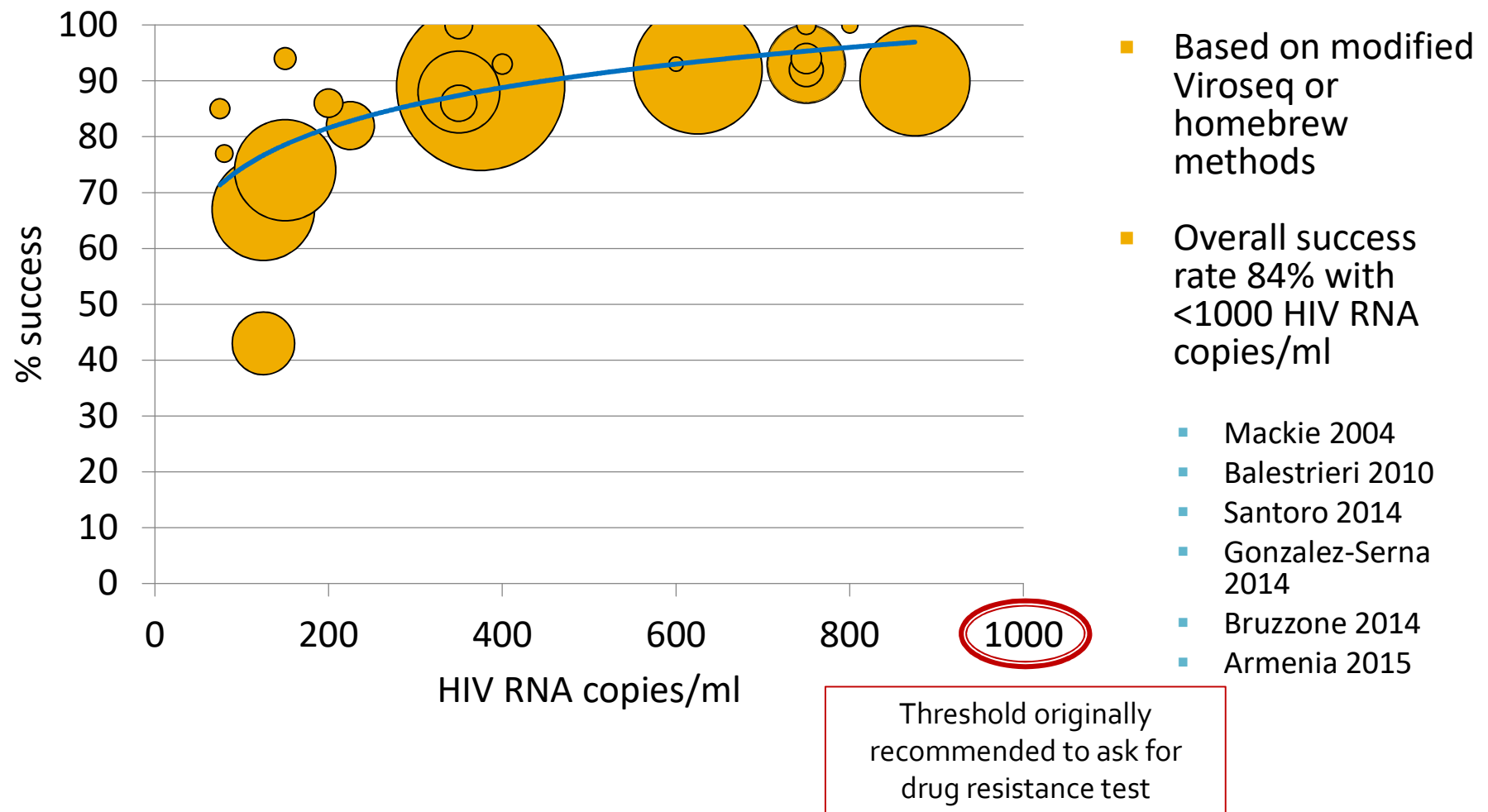


# The evolution of GRT

# HIV drug resistance testing



# Drug resistance genotyping at low-level viremia



# Drug resistance can be detected at low-level viremia



Volume 39, Issue 7

1 October 2004

## Genotypic Resistance in HIV-1–Infected Patients with Persistently Detectable Low-Level Viremia while Receiving Highly Active Antiretroviral Therapy

Richard E. Nettles,<sup>1</sup> Tara L. Kieffer,<sup>1</sup> Rachel P. Simmons,<sup>1</sup> Joseph Cofrancesco, Jr.,<sup>1</sup> Richard D. Moore,<sup>1</sup> Joel E. Gallant,<sup>1</sup> Deborah Persaud,<sup>2</sup> and Robert F. Siliciano<sup>1,3</sup>

Departments of <sup>1</sup>Medicine and <sup>2</sup>Pediatrics, Johns Hopkins University School of Medicine, and <sup>3</sup>Howard Hughes Medical Institute, Baltimore, Maryland

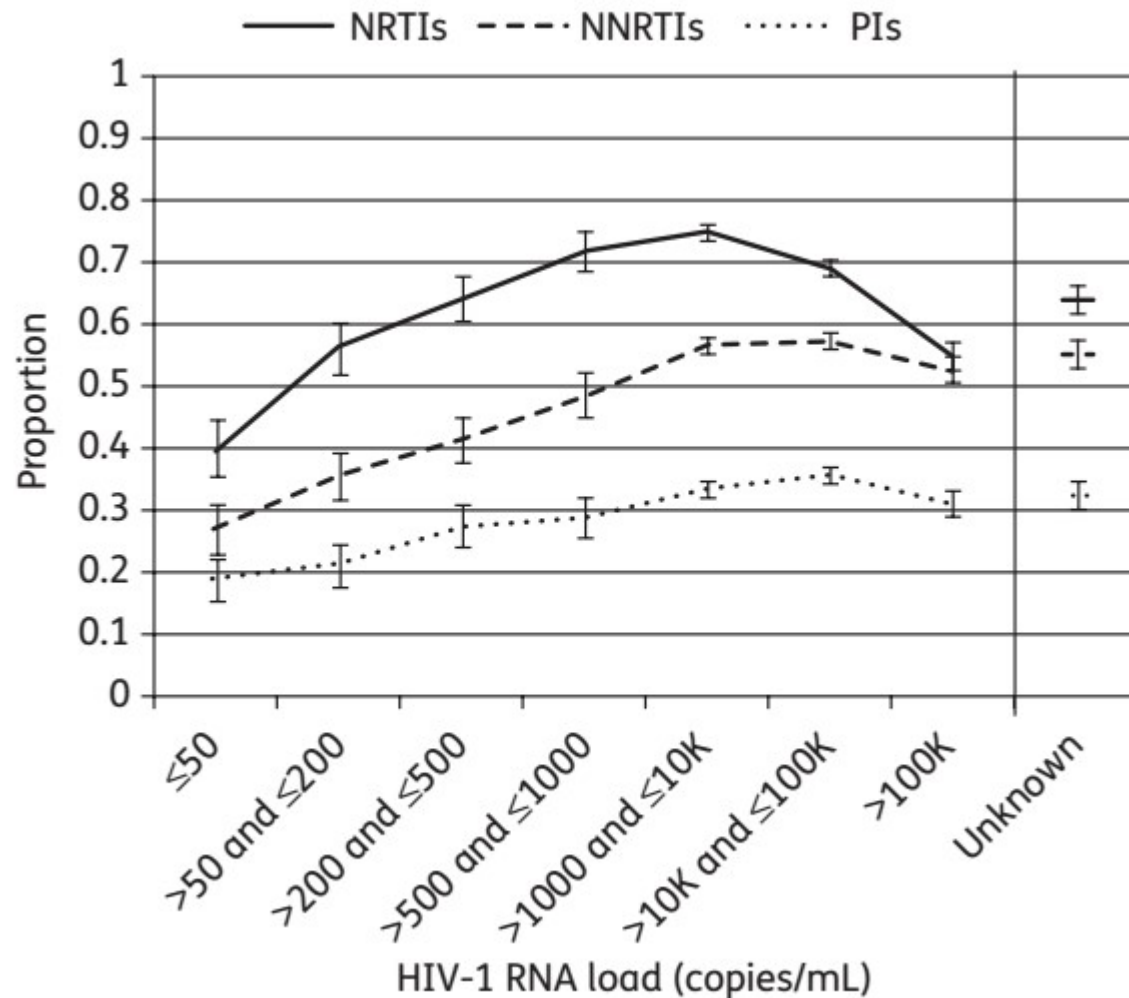
**Background.** Technical limitations in the sensitivity of commercial genotyping methods may prevent clinicians from determining whether drug-resistant human immunodeficiency virus type 1 (HIV-1) is present in patients with low-level viremia. We performed ultrasensitive HIV-1 genotyping for patients with persistent plasma virus loads of 50–400 copies/mL to better define the prevalence of drug resistance and the most common resistance mutations during persistently detectable low-level viremia.

**Methods.** Genotyping of HIV-1 was performed with an ultrasensitive clonal genotyping method.

**Results.** We studied 21 patients who had persistent, detectable, low-level viremia for a median of 11 months. Nine (43%) of 21 patients had HIV-1 isolates with significant resistance mutations. The most common mutations were M184V, K65R, and M41L/T215Y.

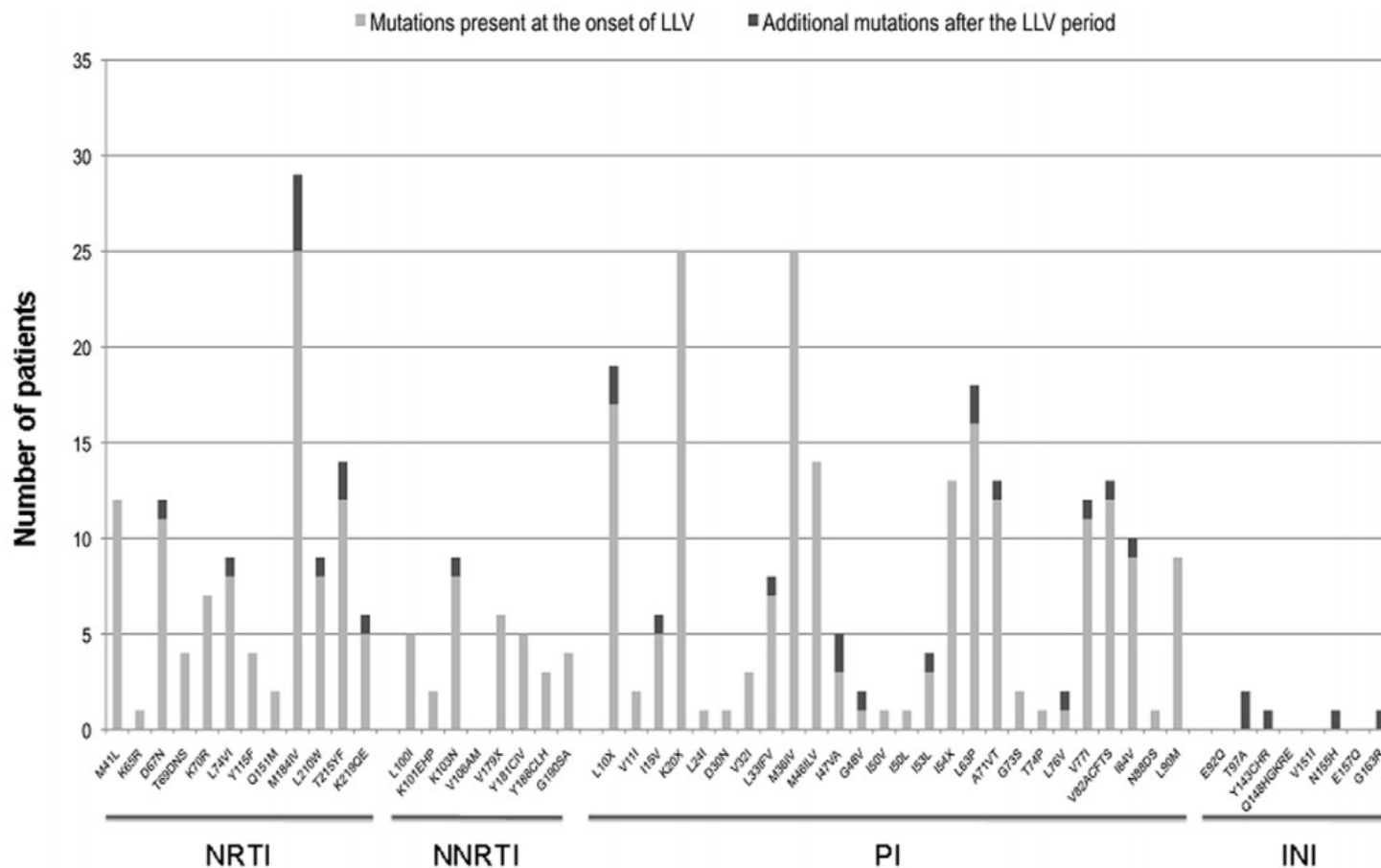
**Conclusions.** The finding that clinically significant resistance mutations were present in some but not all patients with persistent viremia (range, 50–400 copies/mL) highlights the need to improve the sensitivity of current clinical assays for detection of drug resistance.

# Drug resistance can be detected at low-level viremia



- SEHERE consortium (I, UK, P, D, B, E, S)
- 16,511 PR/RT sequences from 11,492 treatment-experienced patients
- 2,500/16,511 (15.14%) test results were obtained at a viral load  $<1,000$  copies/mL

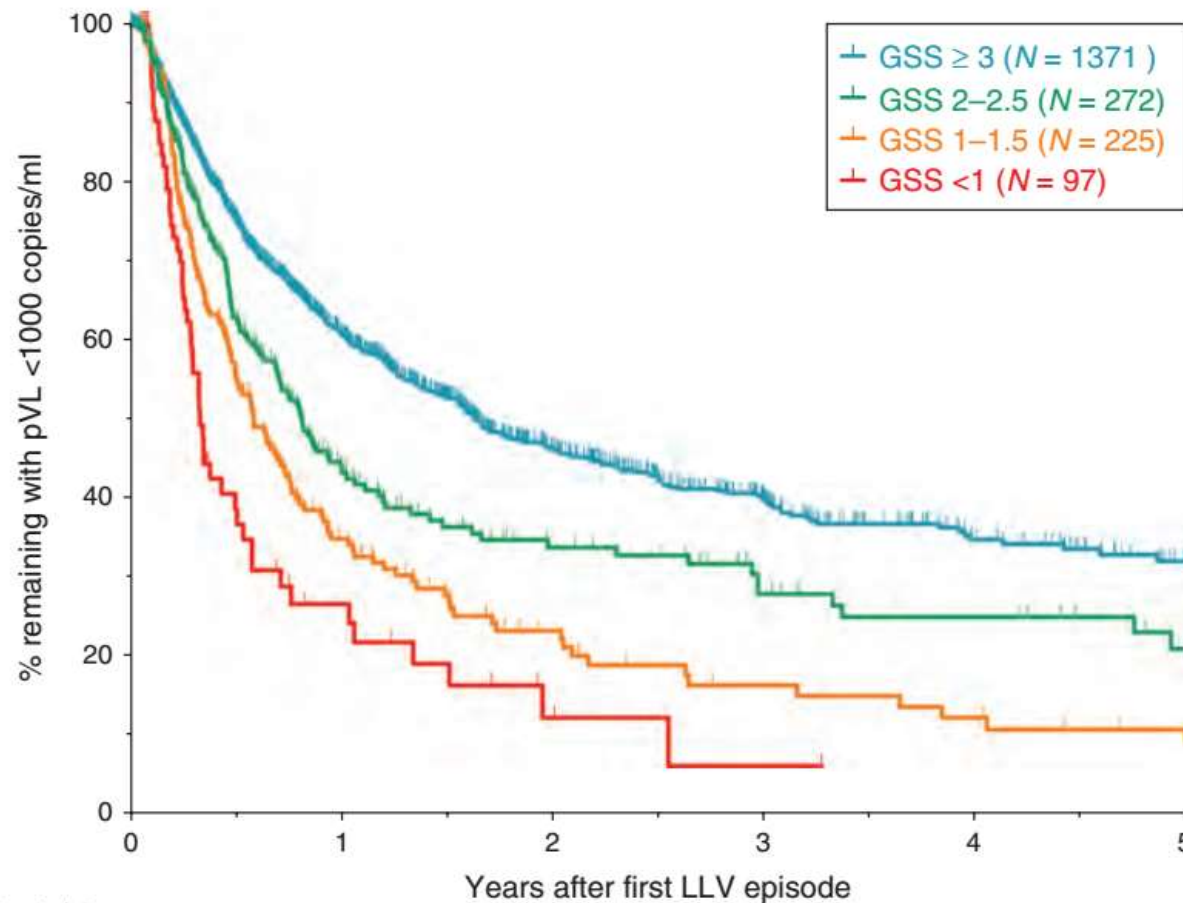
# Drug resistance can emerge during persistent low-level viremia



- 48 patients (4 naive and 44 pretreated) with LLV episode with a median duration of 11 months
- Successful resistance testing at both onset and end of the LLV episode obtained for 37 patients (77%)
- 11 (30%) acquired at least 1 DRAM during the LLV period: for NRTI in 6, for NNRTI in 1, for PI in 4, and for raltegravir in 2



# Drug resistance at low-level viremia predicts virological failure



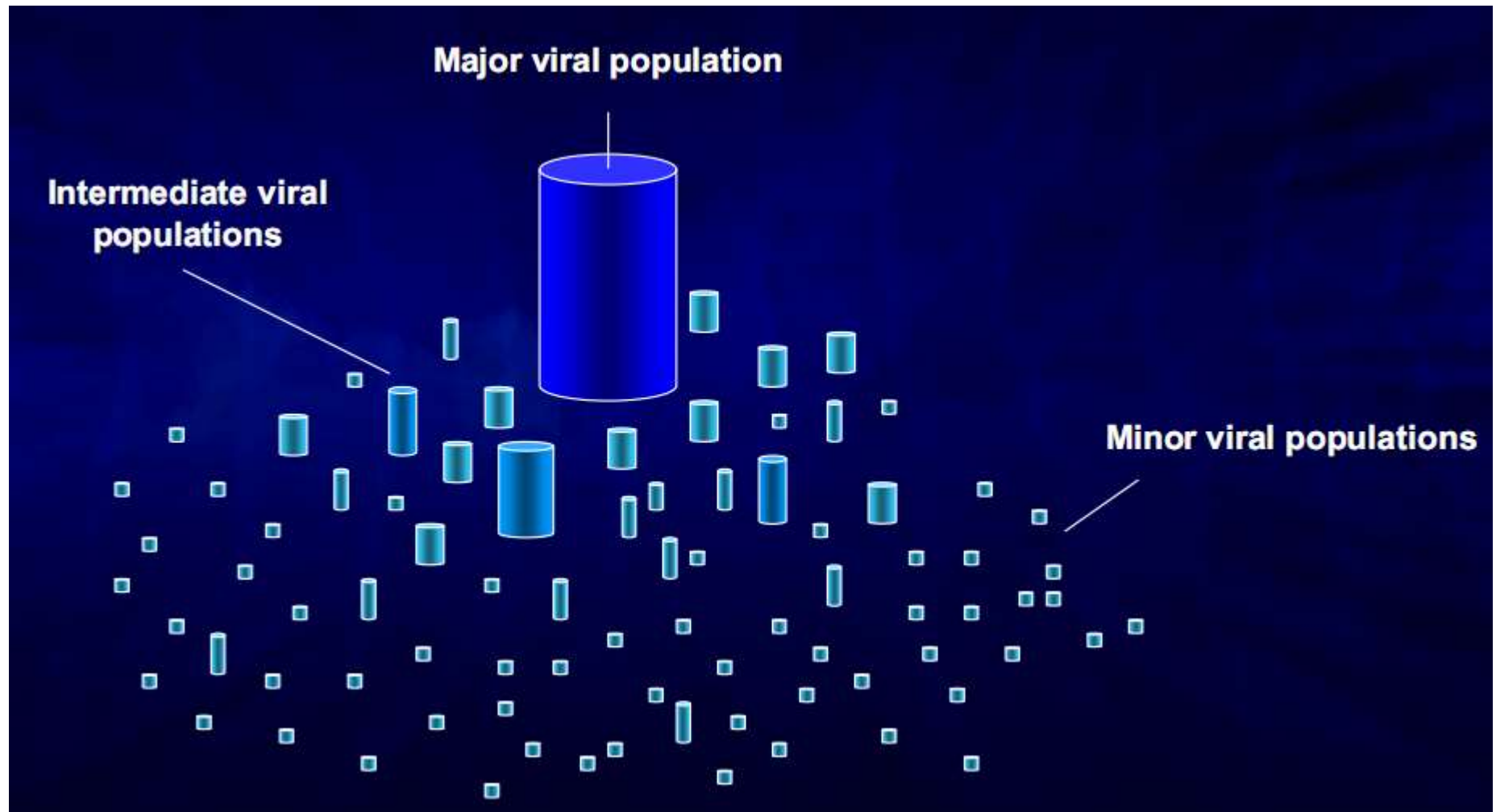
- LLV defined as <1000 copies/mL
- 1702 patients with follow-up on constant therapy eligible for analysis
- 'dose-dependent' increase in the hazard ratio for virologic failure with susceptibility categories at LLV (<1000 copies/mL)

Subjects at risk

$\geq 3$	1228	749	481	349	252	184	133	89	67	51	34
2-2.5	215	108	64	46	37	34	24	18	16	14	11
1-1.5	181	85	48	33	23	16	13	12	10	7	6
<1	78	21	12	8	4	3	1	0	0	0	0

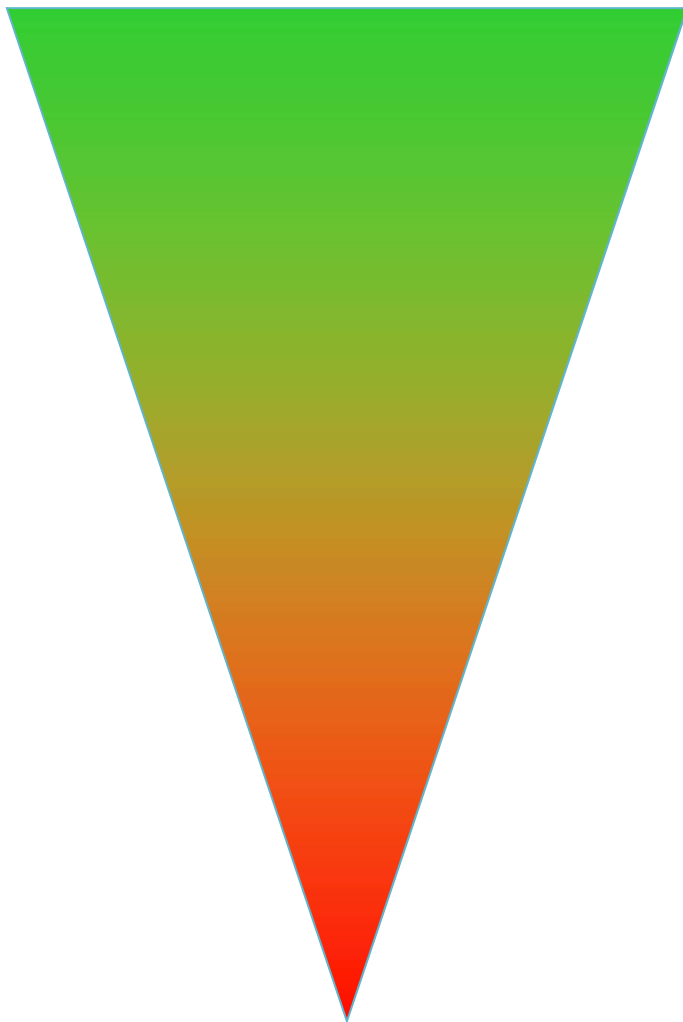


# HIV replicates as quasispecies



# Detectability of drug-resistant variants

## Available methods



>25% of virus population

Standard population sequencing

>5% of virus population

Multiple clones or "single genome sequencing"

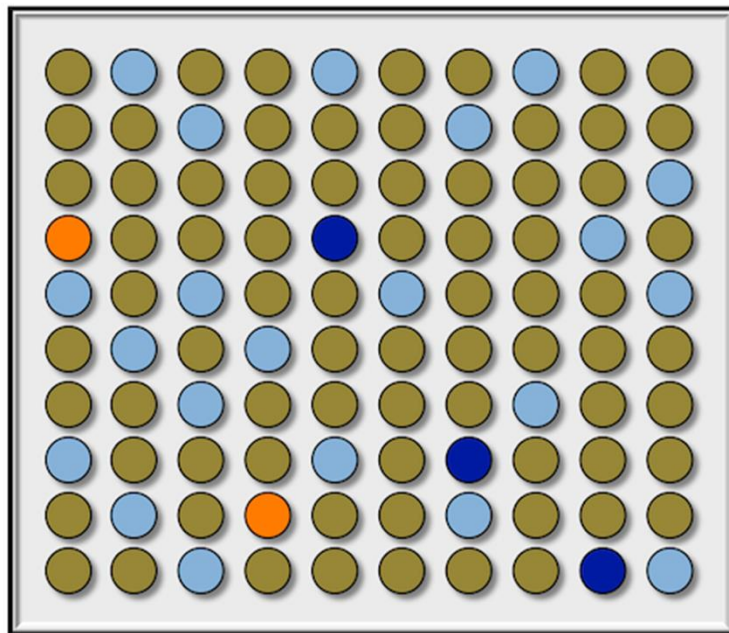
>0.1% of virus population

Point mutation assays (apply to defined mutations)

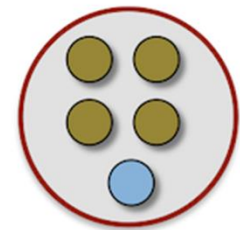
Next generation sequencing

# Sensitivity of genotypic HIV drug resistance testing

## Conventional Bulk Sequencing

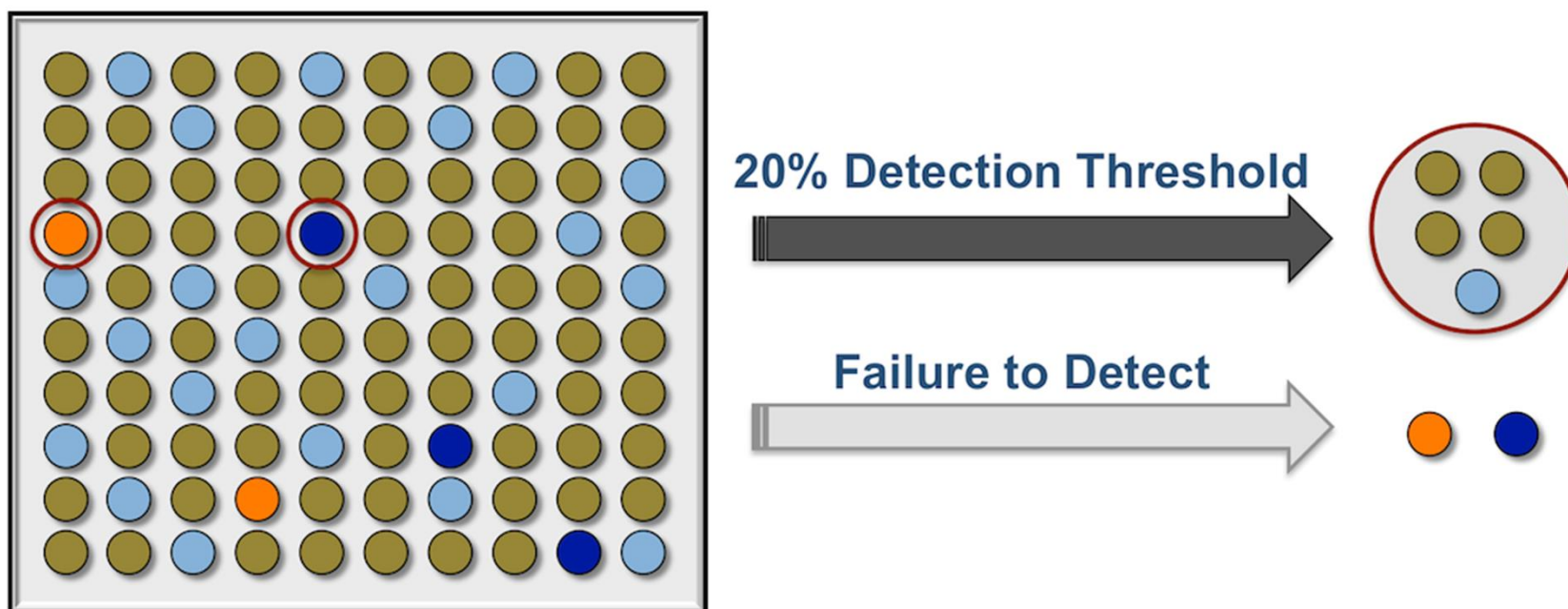


20% Detection Threshold



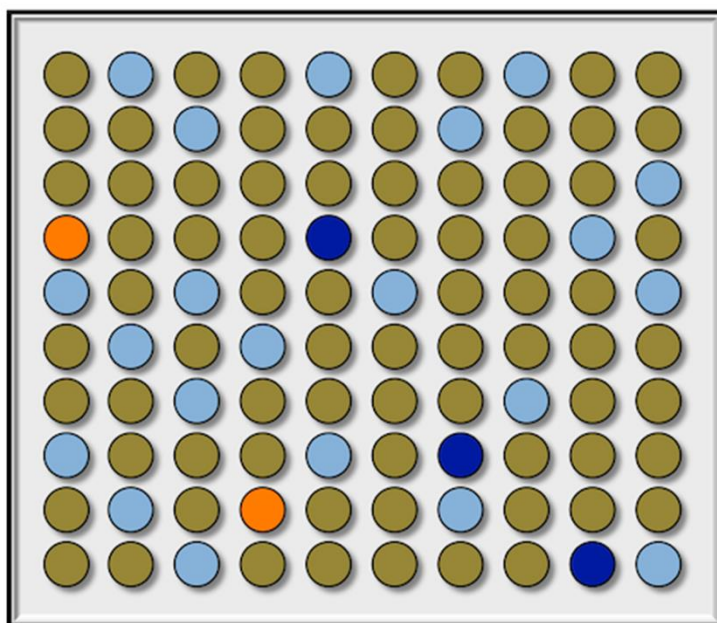
# Sensitivity of genotypic HIV drug resistance testing

## Conventional Bulk Sequencing



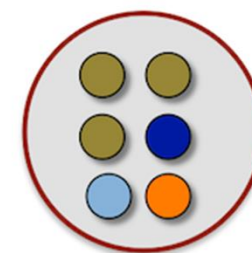
# Sensitivity of genotypic HIV drug resistance testing

## Minority-Variant Sequencing



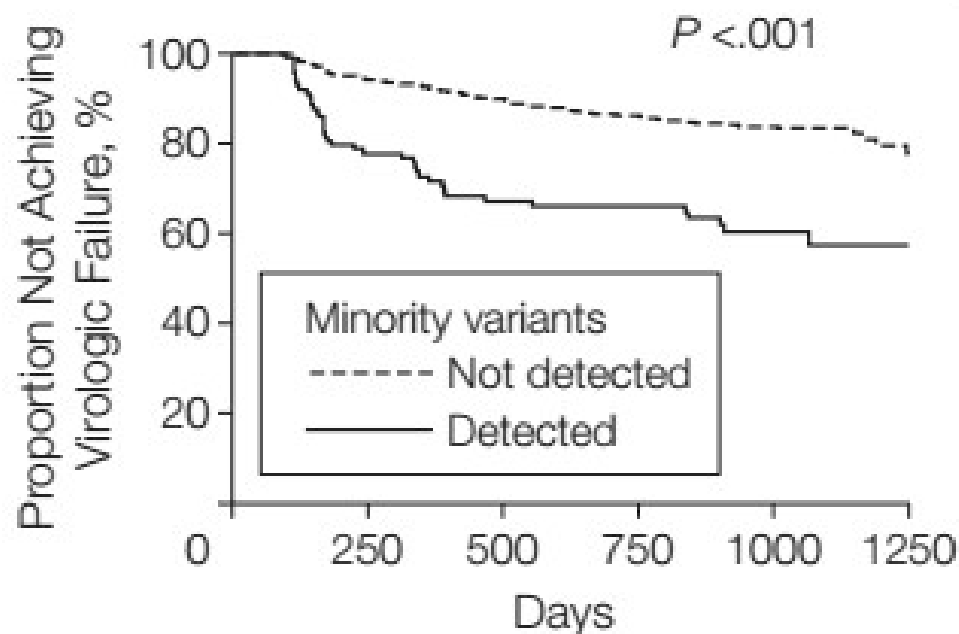
Point Mutation Assays  
Clonal Sequencing  
Ultra-deep Sequencing

< 1% Detection Threshold





# Low-Frequency HIV-1 Drug Resistance Mutations and Risk of NNRTI-Based Antiretroviral Treatment Failure – *A systematic review and pooled analysis*

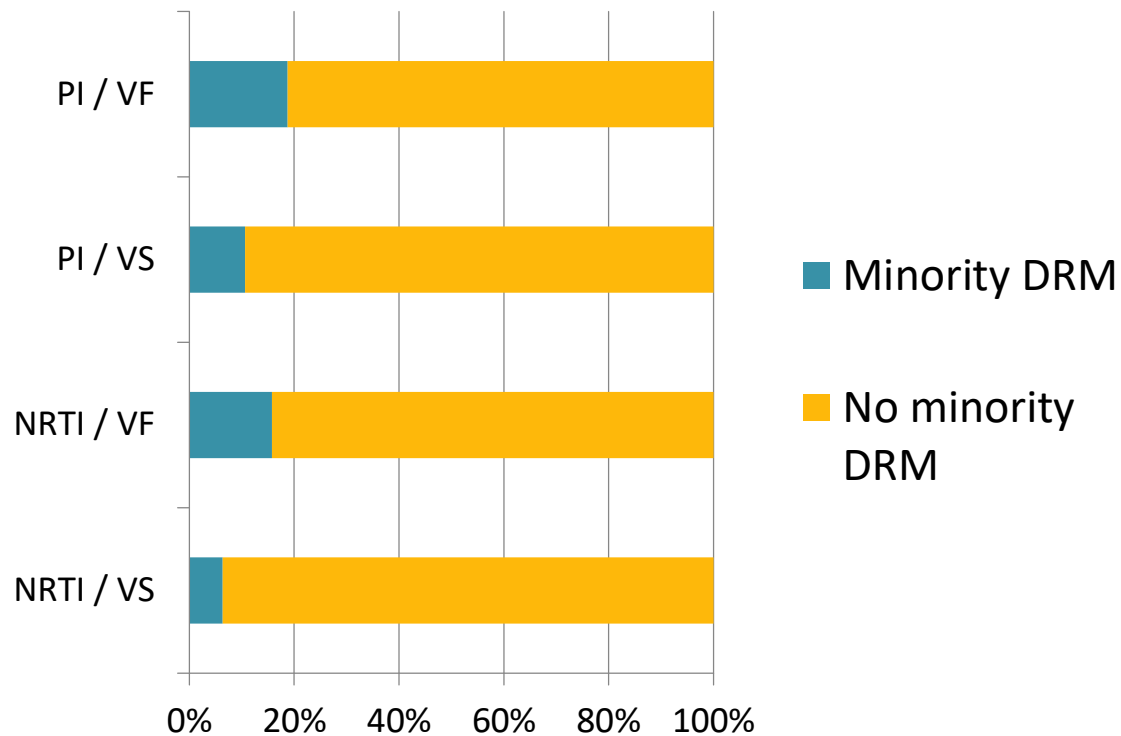


No. at risk						
Minority variants						
Not detected	691	620	455	398	344	46
Detected	117	86	60	53	37	7

- 10 studies, 985 patients, 187 with minority drug resistance mutations (mDRMs)
- mDRMs associated with an increased risk of virologic failure (HR = 2.3; 95% CI 1.7-3.3;  $P < .001$ ) after controlling for medication adherence, race/ethnicity, baseline CD4 cell count, and plasma HIV-1 RNA levels
- Risk most strongly associated with NNRTI mDRMs**
- Dose-dependent increased risk of virologic failure found in participants with a higher proportion or quantity of mDRMs

Kaplan-Meier Curves for Proportion of Patients Without Virologic Failure by Presence of Drug-Resistant HIV-1 Minority Variants

# No impact of HIV-1 protease minority resistant variants on the virological response to a first-line PI-based regimen containing darunavir or atazanavir

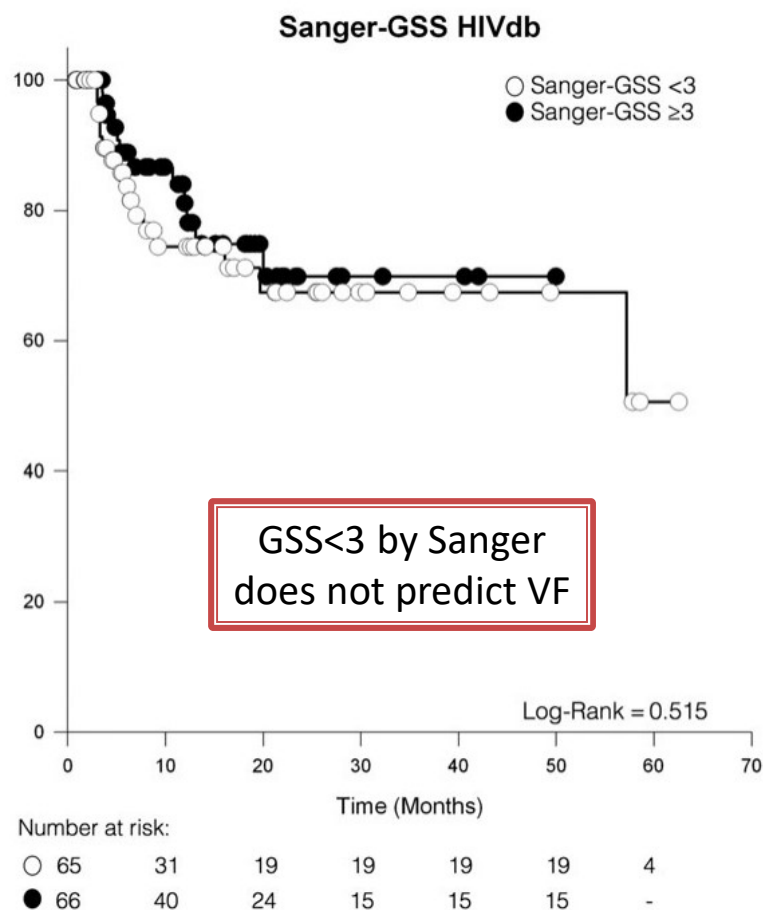
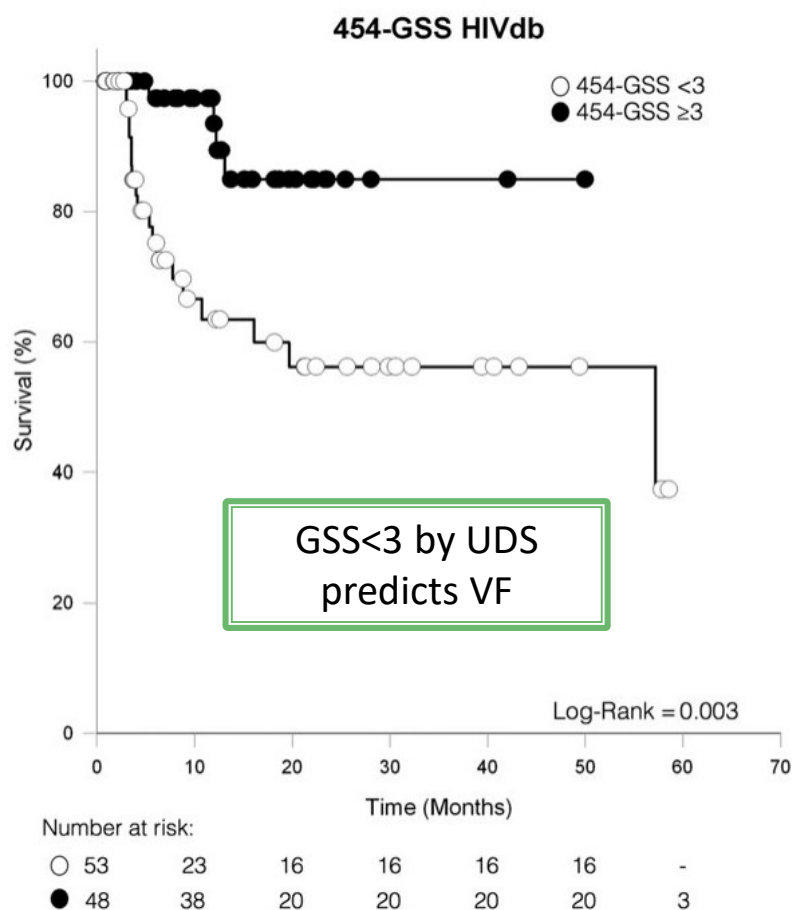


- ART-naïve patients initiating a first-line regimen including two NRTI and DRV/r (n = 94) or ATV/r (n = 16), Jan 2012 - Mar 2015
- Ultra-deep sequencing (Illumina)
- Virological failure occurring in 13 patients (13.8%) with DRV and 3 (18.8%) with ATV
- 13 (11.8%) subjects had PI MRV at baseline in the median proportion of 1.3% (IQR = 1.1–1.7)
- The most prevalent PI MRV were G73C (n = 5) and M46I (n = 3)



# Improved Prediction of Salvage Antiretroviral Therapy Outcomes Using Ultrasensitive HIV-1 Drug Resistance Testing

132 subjects starting salvage therapy with at least one among PI/r, RAL, ETR; 28 (21%) developed VF.



# Next-Generation Sequencing to Help Monitor Patients Infected with HIV: Ready for Clinical Use?

- **Evidence supporting clinical utility (slowly) increasing**
  - Drug resistance in untreated patients
  - Choice of salvage regimen in highly treatment experienced patients
  - Coreceptor tropism assay for treatment with CCR5 antagonists
- **Barriers to widespread use remaining**
  - Cost of NGS instrumentation plus data analysis and storage solutions
  - Technical expertise required (platforms evolving or superseded rapidly)
  - No clinical trial testing NGS available
  - No IVD approval yet

# Next-Generation Sequencing to Help Monitor Patients Infected with HIV: Ready for Clinical Use?

## Vela Next Generation Sequencing

NGS automation for the IVD routine laboratory



CE-IVD marked on  
Aug 21, 2017

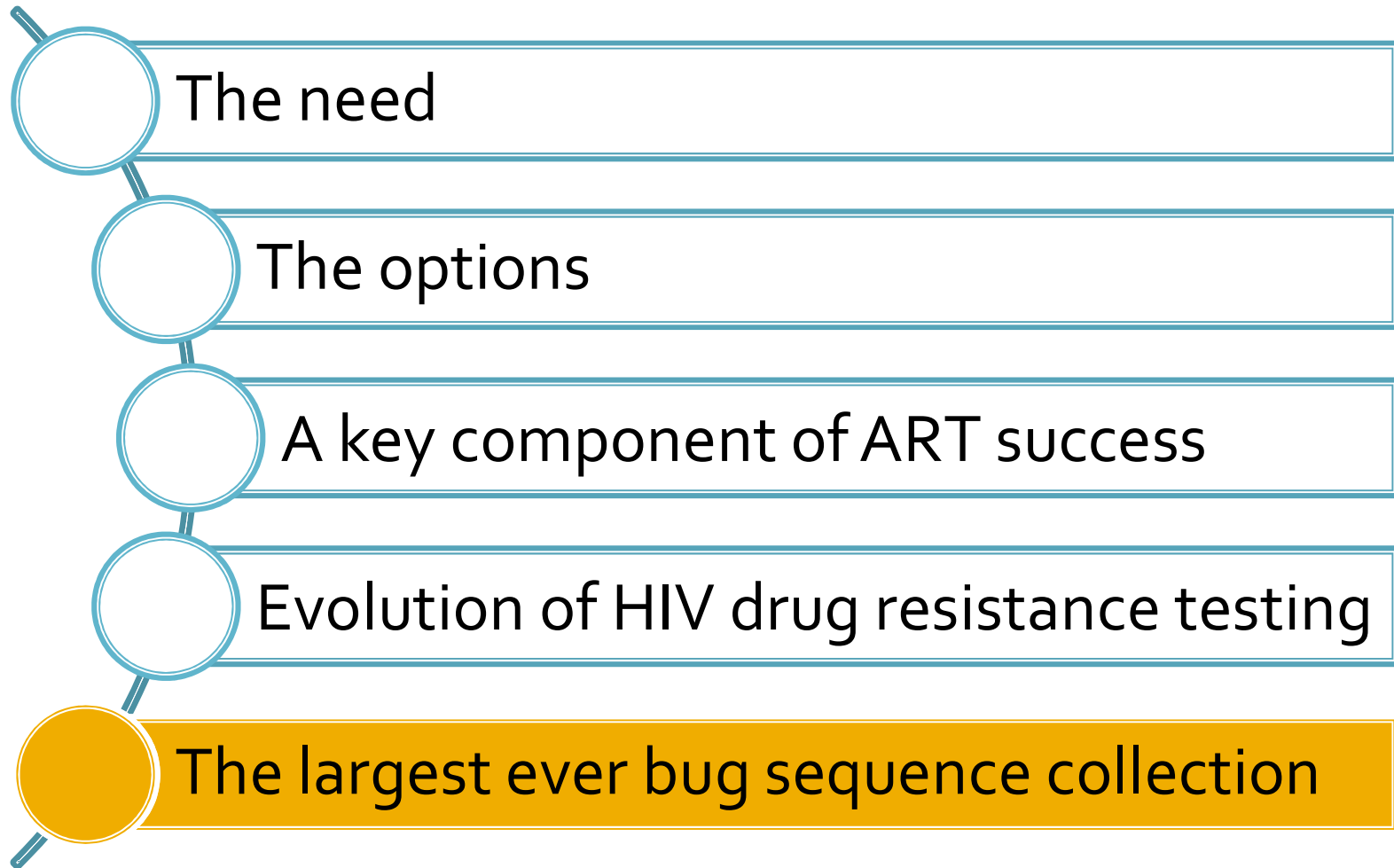


### Key Features

- **CE-IVD**
- Ready to use in 2 weeks from installation
- IT Connectivity
- Sample Tracking
- Control System
- Automation
- Ready to use Reagents
- Actionable Design
- Data Analysis
- Data Reporting

Opportunities  
from the largest  
ever bug sequence  
collection

# HIV drug resistance testing



# Massive use of HIV drug resistance genotyping in the clinic

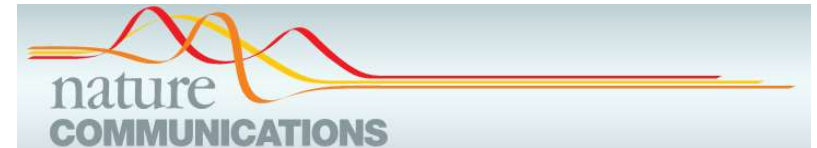
- Thousands HIV sequences generated in each country
- Huge international databases
  - E.g. Stanford, RDI, EuResist
- Opportunity for novel analysis
  - Phylodynamics, phylogeography, cluster analysis



# Challenges from huge HIV (drug resistance) sequence databases

## ARTICLE

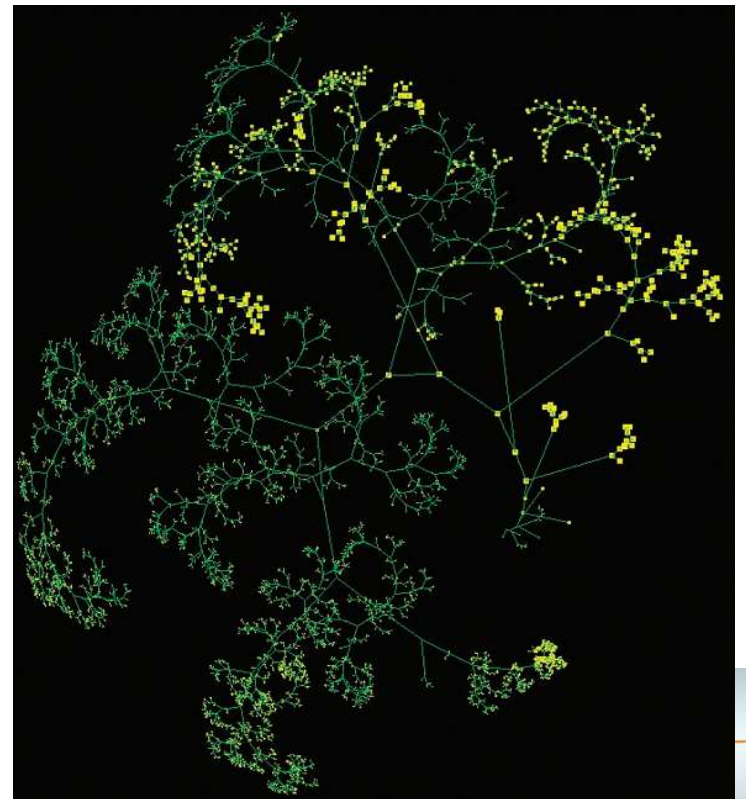
Received 17 Jan 2011 | Accepted 21 Apr 2011 | Published 24 May 2011



# A novel methodology for large-scale phylogeny partition

Mattia C.F. Prosperi<sup>1,2,3</sup>, Massimo Ciccozzi<sup>4</sup>, Iuri Fanti<sup>1</sup>, Francesco Saladini<sup>5</sup>, Monica Pecorari<sup>6</sup>, Vanni Borghi<sup>7</sup>, Simona Di Giambenedetto<sup>1</sup>, Bianca Bruzzone<sup>8</sup>, Amedeo Capetti<sup>9</sup>, Angela Vivarelli<sup>10</sup>, Stefano Rusconi<sup>11</sup>, Maria Carla Re<sup>12</sup>, Maria Rita Gismondo<sup>13</sup>, Laura Sighinolfi<sup>14</sup>, Rebecca R. Gray<sup>2,3</sup>, Marco Salemi<sup>2,3</sup>, Maurizio Zazzi<sup>5</sup> & Andrea De Luca<sup>1,15</sup> on behalf of the ARCA collaborative group

**Figure 2 | Phylogeny of Italian HIV-1 subtype B *pol* isolates.** Maximum-likelihood phylogenetic tree of 11,541 HIV-1 subtype B *pol* gene sequences from the Italian ARCA cohort. Tree is rooted on subtype J and depicted using three-dimensional hyperbolic geometry. Nodes and leaves are highlighted by yellow points.





# Opportunities from huge HIV (drug resistance) sequence databases

## Retrovirology

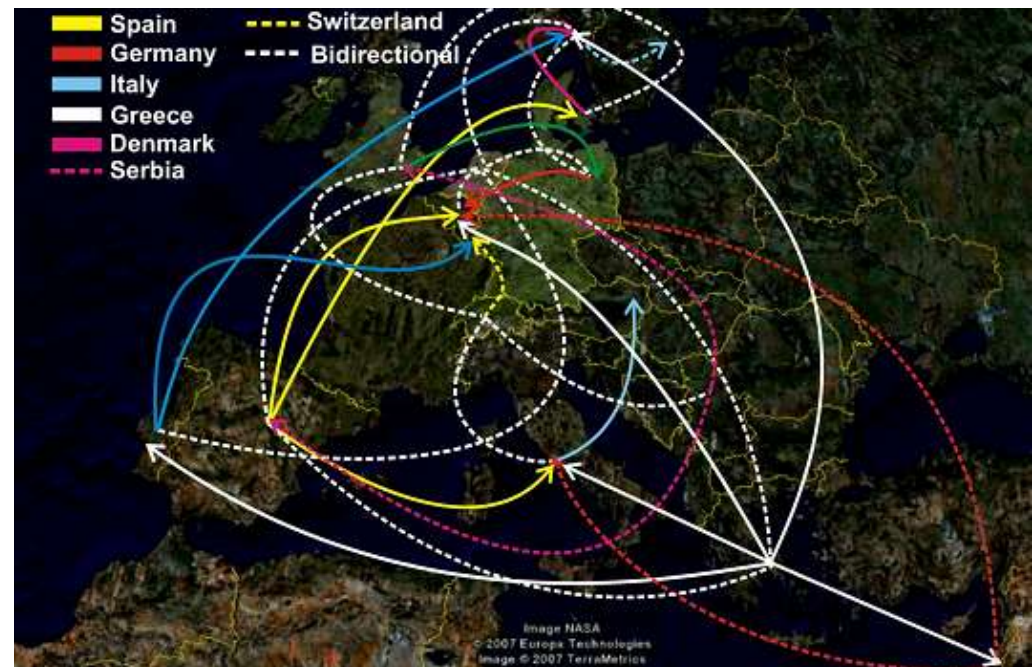


Research

Open Access

### Tracing the HIV-1 subtype B mobility in Europe: a phylogeographic approach

Dimitrios Paraskevis<sup>\*1,2</sup>, Oliver Pybus<sup>3</sup>, Gkikas Magiorkinis<sup>2</sup>, Angelos Hatzakis<sup>2</sup>, Annemarie MJ Wensing<sup>4</sup>, David A van de Vijver<sup>5</sup>, Jan Albert<sup>6,7</sup>, Guiseppe Angarano<sup>8</sup>, Birgitta Åsjö<sup>9</sup>, Claudia Balotta<sup>10</sup>, Enzo Boeri<sup>11</sup>, Ricardo Camacho<sup>12</sup>, Marie-Laure Chaix<sup>13</sup>, Suzie Coughlan<sup>14</sup>, Dominique Costagliola<sup>15</sup>, Andrea De Luca<sup>16</sup>, Carmen de Mendoza<sup>17</sup>, Inge Derdelinckx<sup>18</sup>, Zehava Grossman<sup>19</sup>, Osama Hamouda<sup>20</sup>, IM Hoepelman<sup>21</sup>, Andrzej Horban<sup>22</sup>, Klaus Korn<sup>23</sup>, Claudia Kücherer<sup>20</sup>, Thomas Leitner<sup>6,7</sup>, Clive Loveday<sup>24</sup>, Eilidh MacRae<sup>25</sup>, I Maljkovic-Berry<sup>6,7</sup>, Laurence Meyer<sup>25</sup>, Claus Nielsen<sup>26</sup>, Eline LM Op de Coul<sup>27</sup>, Vidar Ormaasen<sup>28</sup>, Luc Perrin<sup>29</sup>, Elisabeth Puchhammer-Stöckl<sup>30</sup>, Lidia Ruiz<sup>31</sup>, Mika O Salminen<sup>32</sup>, Jean-Claude Schmit<sup>33</sup>, Rob Schuurman<sup>4</sup>, Vincent Soriano<sup>17</sup>, J Stanczak<sup>22</sup>, Maja Stanojevic<sup>34</sup>, Daniel Struck<sup>33</sup>, Kristel Van Laethem<sup>1</sup>, M Violin<sup>10</sup>, Sabine Yerly<sup>29</sup>, Maurizio Zazzi<sup>35</sup>, Charles A Boucher<sup>4,5</sup>, Anne-Mieke Vandamme<sup>1</sup> for the SPREAD Programme



*Retrovirology* 2009, 6:49

# Mapping HIV TDR in Europe



## ESAR - SPREAD

### Prevalence of TDR

#### Menu

Public access:

[Subtype distribution](#)

[Prevalence of TDR](#)

#### Database Login:

username:

password:

[\(support\)](#)

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RESEARCH DEDICATED TO LIFE  
(developed and hosted by  
Luxembourg Institute of Health)

#### Route of transmission

- ☒ All
- ☐ HSX
- ☐ MSM
- ☐ IDU
- ☐ OTHER
- ☐ UNK

#### Gender

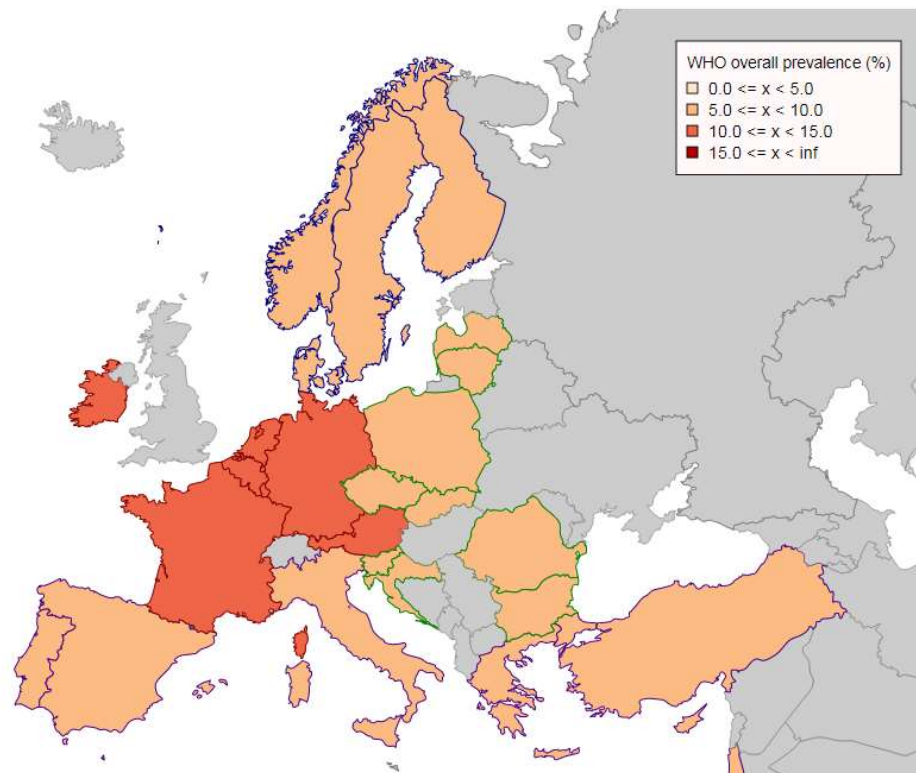
- ☒ All
- ☐ Male
- ☐ Female

#### Year of HIV diagnosis

- ☒ All
- ☐ 2000-2005
- ☐ 2006-2007
- ☐ 2008-2010

#### TDR

mutations associated with resistance to NRTIs, NNRTIs or PIs  
mutations associated with resistance to PIs  
mutations associated with resistance to NRTIs  
mutations associated with resistance to NNRTIs



# Mapping HIV subtype distribution in Europe



## ESAR - SPREAD

### Subtype distribution

#### Menu

Public access:

[Subtype distribution](#)

[Prevalence of TDR](#)

#### Database Login:

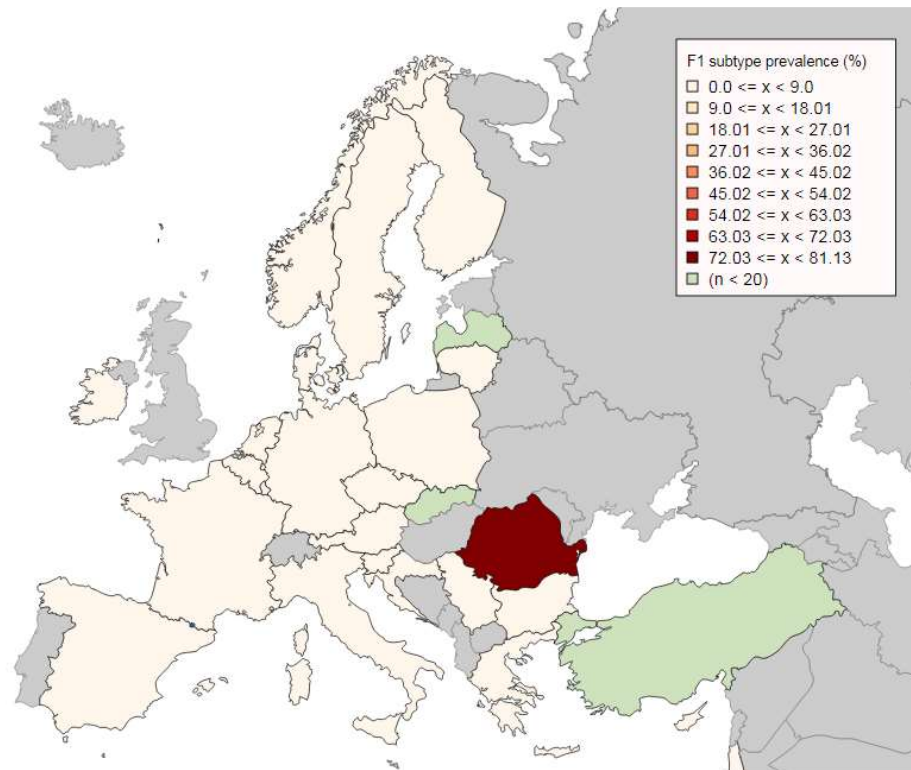
username:

password:

[\(support\)](#)

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(developed and hosted by  
[Luxembourg Institute of Health](#))

<b>Route of transmission</b>	<b>Gender</b>	<b>Year of HIV diagnosis</b>	<b>Subtype</b>
<input checked="" type="checkbox"/> All <input type="checkbox"/> HSX <input type="checkbox"/> MSM <input type="checkbox"/> IDU <input type="checkbox"/> OTHER <input type="checkbox"/> UNK	<input checked="" type="checkbox"/> All <input type="checkbox"/> Male <input type="checkbox"/> Female	<input type="checkbox"/> All <input type="checkbox"/> 2000-2005 <input type="checkbox"/> 2006-2007 <input checked="" type="checkbox"/> 2008-2010	<input type="button" value="submit"/> B non-B A1 C 02_AG 01_AE F1



# Conclusions



# Lessons learned: antiretroviral drug resistance testing...

- ...has guided **drug development** and shaped the success of antiretroviral therapy
- ...has been the first example of integration of a sophisticated molecular assay into **clinical practice** in the infectious diseases domain
- ...has generated a huge HIV sequence repositories allowing to track HIV evolution and inform **surveillance and education programs**