



associazione  
microbiologi  
clinici italiani

**XLVIII  
CONGRESSO  
NAZIONALE  
AMCLI**

**2019**



9-12 NOVEMBRE 2019  
PALACONGRESSI RIMINI

# IL MICROBIOTA

**Microbiota: non solo  
intestino**

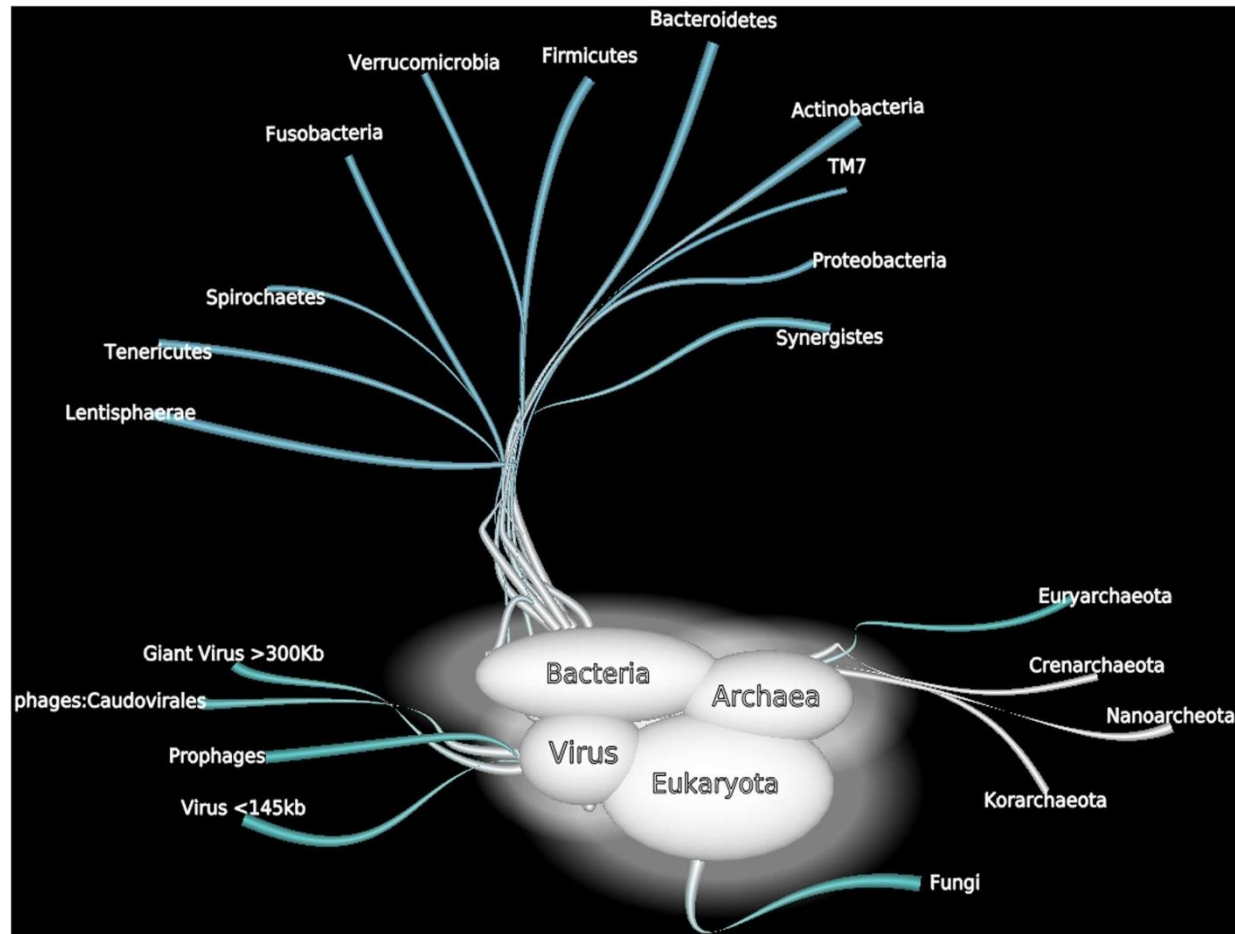
**Maurizio Sanguinetti**

# Glossary of terms

Definition	Explanation
Microbiota	All the microbes that are found in a particular region or habitat; the term “microflora” is no longer used
Microbiome	The totality of the microbes with their genes that are harbored by the microbiota and the milieu in which they interact
Operational taxonomic unit (OTU)	Specific sequences based on sequence similarity (typically threshold is 97%) to reference genes. This is taken as a proxy for species-level
Taxon	A group of phylogenetically related microbes that belong to the same taxonomic group, such as order, family, or genus
Richness	Number of different taxa within a single population
$\alpha$ Diversity	How many types of sequences in a sample
$\beta$ Diversity	How many different types of sequences are shared among samples

- Humans are viewed as composites of human and microbial cells.
- Human microbiota are complex and dynamic microbial communities composed mainly of bacteria, but also includes protozoa, archaea, viruses, and fungi that resides in and on different body niches (oral cavity, throat, esophagus, stomach, colon, urogenital tract, respiratory tract, and skin).
- The colonic microbiota constitutes the most abundant microbial domain within the human body, with the vast majority belonging to the bacterial phyla of *Firmicutes* and *Bacteroidetes*.

# A non-exhaustive overview of human gut microorganisms among bacterial, *Archaea*, viral, and *Eukaryota* domains



# Microbiota influencers

- Sanitation

- Mode of delivery, breast-feeding..

- Diet

- *Composition (calories, fat, fibres, vegetable, meat..)*
- *Cooking*
- *Natural food additives (safrol..)*
- *Artificial chemical food additives:*
  - *Preservatives (benzoic acid, sodium benzoate, nitrite/nitrate, sulfur dioxide/sulfite..)*
  - *Sweeteners, emulsifiers and stabilizers, flavors, thickeners, antifoaming, anticaking, bulking, antioxidants..)*
  - *Others (titanium dioxide..)*

- Exercise

- Sleep

- Stress

- Violence

- Drugs

# How to define an EUBIOTIC enterotype?

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*EU= good*

*BIOS= life*

- **Composition:** *Diversity*  
*Richness*  
*Relative Abundance*

*Our gut is a sophisticated ecosystem that is regulated by the logic of RELATIONAL HARMONY*

*Microbiota and Host live in a COOPERATIVE  
SYSTEMIC AGGREGATION MODEL*

# EUBIOSIS



***Failure of HOST-MICROBIOTA equilibrium***



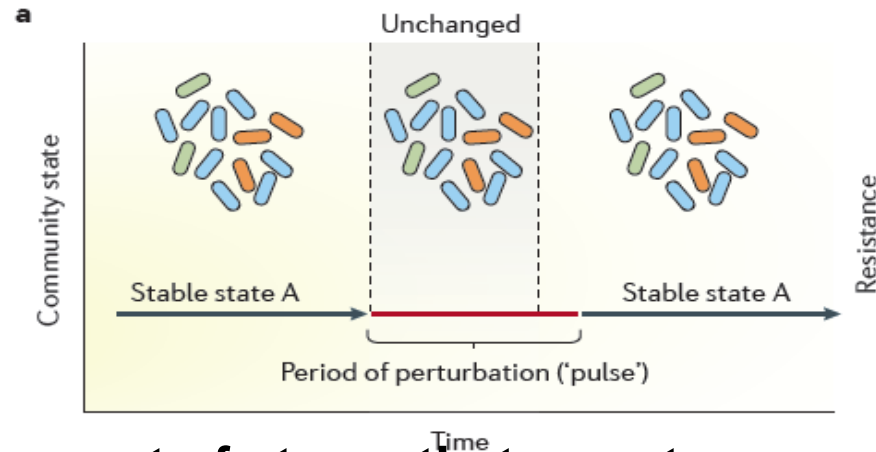
***Quali-quantitative alterations of oral,  
esophageal, gastric, small bowel and/or  
colonic microbiota***



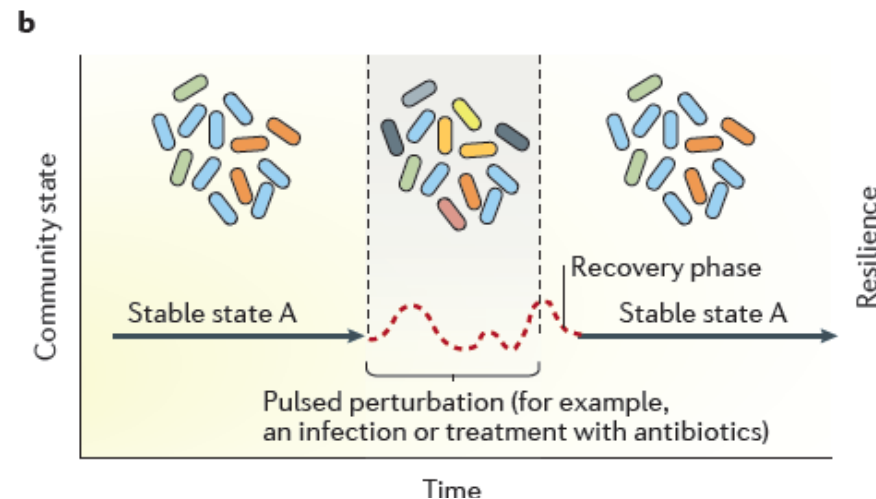
# DYSBIOSIS

# From *EUBIOSIS* to *DYSBIOSIS*

**RESISTANCE:** a given ecosystem to stand unchanged in the face of a disturbance



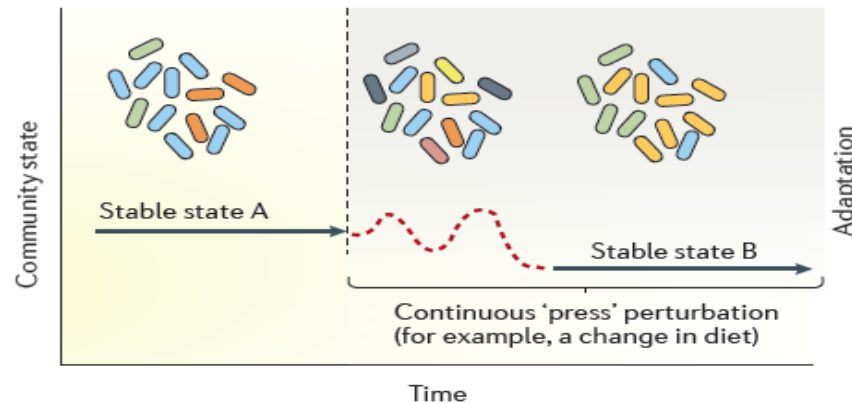
**RESILIENCE:** amount of stress that a system can tolerate before its homeostatic state shifts towards a new equilibrium with different functions and services



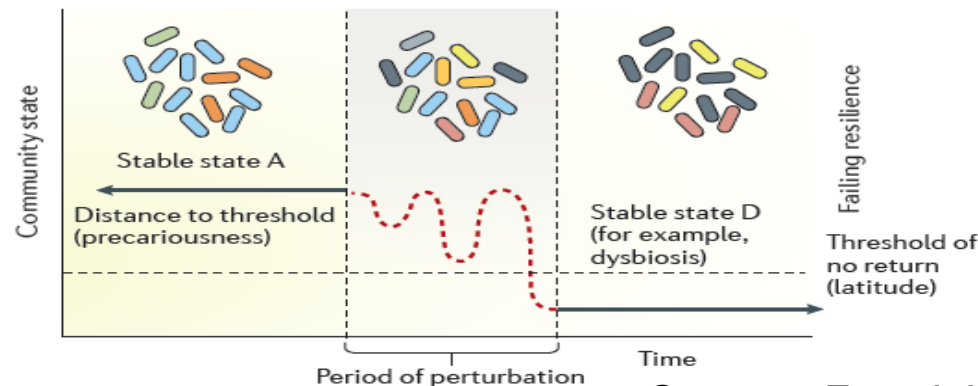


# From *EUBIOSIS* to *DYSBIOSIS*

Facing a continuous perturbation, the composition of the microbiota may adopt a new beneficial or detrimental state: **ADAPTATION**



**Acquisition of an unhealthy and dysbiotic microbiota with high resilience potential may contribute to chronicity of human microbiota-associated diseases**



# Stress and stability: applying the Anna Karenina principle to animal microbiomes

Jesse R. Zaneveld<sup>1\*</sup>, Ryan McMinds<sup>2</sup> and Rebecca Vega Thurber<sup>2\*</sup>

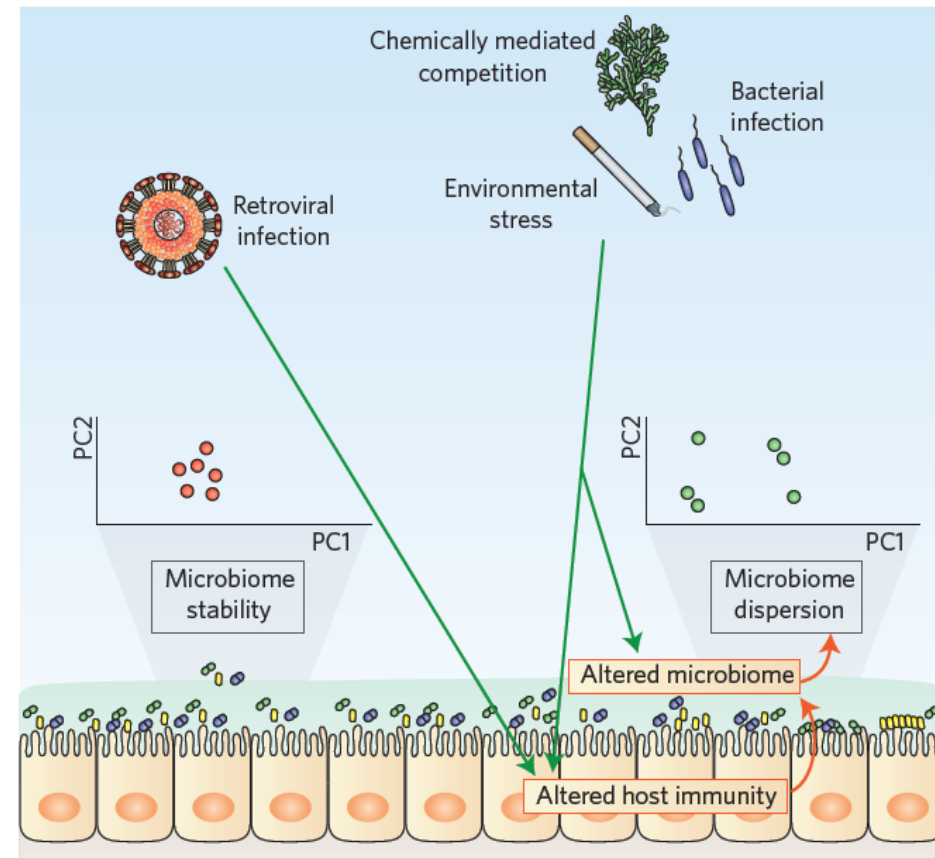
- “all happy families look alike; each unhappy family is unhappy in its own way”
- “Tutte le famiglie felici sono simili le une alle altre; ogni famiglia infelice è infelice a modo suo”



# “all healthy microbiomes are similar; each dysbiotic microbiome is dysbiotic in its own way”

- Typically, healthy hosts possess relatively stable microbiomes that form tight clusters in ordination space.
- In contrast to movement of these clusters to a new place in ordination space a variety of external stressors have been shown to disrupt this stability, resulting in more dispersed microbiomes.
- More dispersed microbiomes have been associated with a variety of negative outcomes for the host, including increased invasibility and altered clinical parameters (for example, endotoxaemia in alcoholics).
- In principle, these disruptions may act indirectly by affecting host immunity (as in HIV and SIVcpz), indirectly by altering the microbiome (for example, by displacing protective mutualists like antibiotic producers), or through a combination of both mechanisms.

## Anna Karenina principle of perturbations inducing microbiome destabilization



From: Zaneveld *et al*, Nat. Microbiol., 2017

# EFFECTS OF GUT MICROBIOTA ON HOST HEALTH

- Barrier effect
- **Immunocompetence/Tolerance**
- **Protection against infections**
- Synthesis
- Metabolic/Trophic function
- Drug metabolism
- Behavior conditioning

# ***Gut microbiome precision medicine***

***Moving to a **Microbiota**  
**signature** for any disease or  
pathological status?***



OPEN ACCESS

ORIGINAL ARTICLE

## A microbial signature for Crohn's disease

### How might it impact on clinical practice in the foreseeable future?

- ▶ Considering CD and UC as two distinct subtypes of IBD at the microbiome level could help designing specific therapeutic targets.
- ▶ The microbial signature specific to CD combined with either imaging techniques or calprotectin data could help decision-making when the diagnosis is initially uncertain among CD, UC and IBS.

### What are the new findings?

- ▶ Dysbiosis is greater in CD than in UC, with a lower microbial diversity, a more altered microbiome composition and a more unstable microbial community.
- ▶ Different microbial groups are associated with smoking habit and localisation of the disease in CD and UC.
- ▶ Eight groups of microorganisms including *Faecalibacterium*, an unknown Peptostreptococcaceae, *Anaerostipes*, *Methanobrevibacter*, an unknown Christensenellaceae, *Collinsella* and *Fusobacterium*, *Escherichia* could be used to discriminate CD from non-CD; the six first groups being in lower relative abundance and the last two groups in higher relative abundance in CD.



ORIGINAL ARTICLE

## A microbial signature for Crohn's disease

Victoria Pascal,<sup>1</sup> Marta Pozuelo,<sup>1</sup> Natalia Borrue, <sup>1,2</sup> Francesc Casellas,<sup>1,2</sup>  
David Campos,<sup>1</sup> Alba Santiago,<sup>1</sup> Xavier Martinez,<sup>1</sup> Encarna Varela,<sup>1</sup>  
Guillaume Sarrabayrouse,<sup>1</sup> Kathleen Machiels,<sup>3</sup> Severine Vermeire,<sup>3</sup> Harry Sokol,<sup>4</sup>  
Francisco Guamer,<sup>1,2</sup> Chaysavanh Manichanh<sup>1,2</sup>

- We analysed a cohort of 2045 non-IBD and IBD faecal samples from four countries (Spain, Belgium, the UK and Germany), applied a 16S rRNA sequencing approach and analysed a total dataset of 115 million sequences.
- In the Spanish cohort, dysbiosis was found significantly greater in patients with CD than with UC, as shown by a more reduced diversity, a less stable microbial community and eight microbial groups were proposed as a specific microbial signature for CD.
- Tested against the whole cohort, the signature achieved an overall sensitivity of 80% and a specificity of 94%, 94%, 89% and 91% for the detection of CD versus healthy controls, patients with anorexia, IBS and UC, respectively.
- Although UC and CD share many epidemiologic, immunologic, therapeutic and clinical features, these results showed that they are two distinct subtypes of IBD at the microbiome level.
- For the first time, microbiomarkers were proposed to discriminate between CD and non-CD independently of geographical regions.

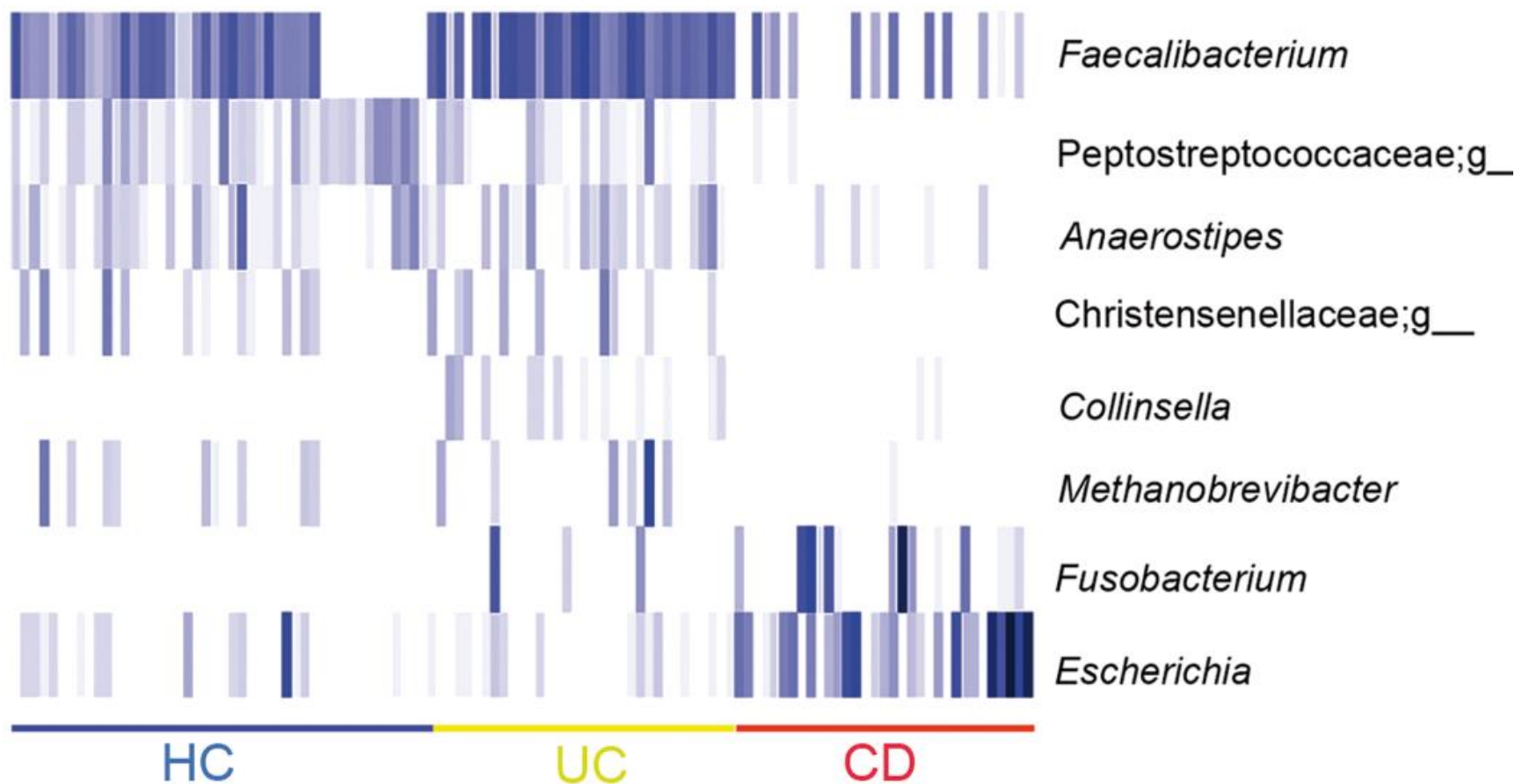


ORIGINAL ARTICLE

## A microbial signature for Crohn's disease

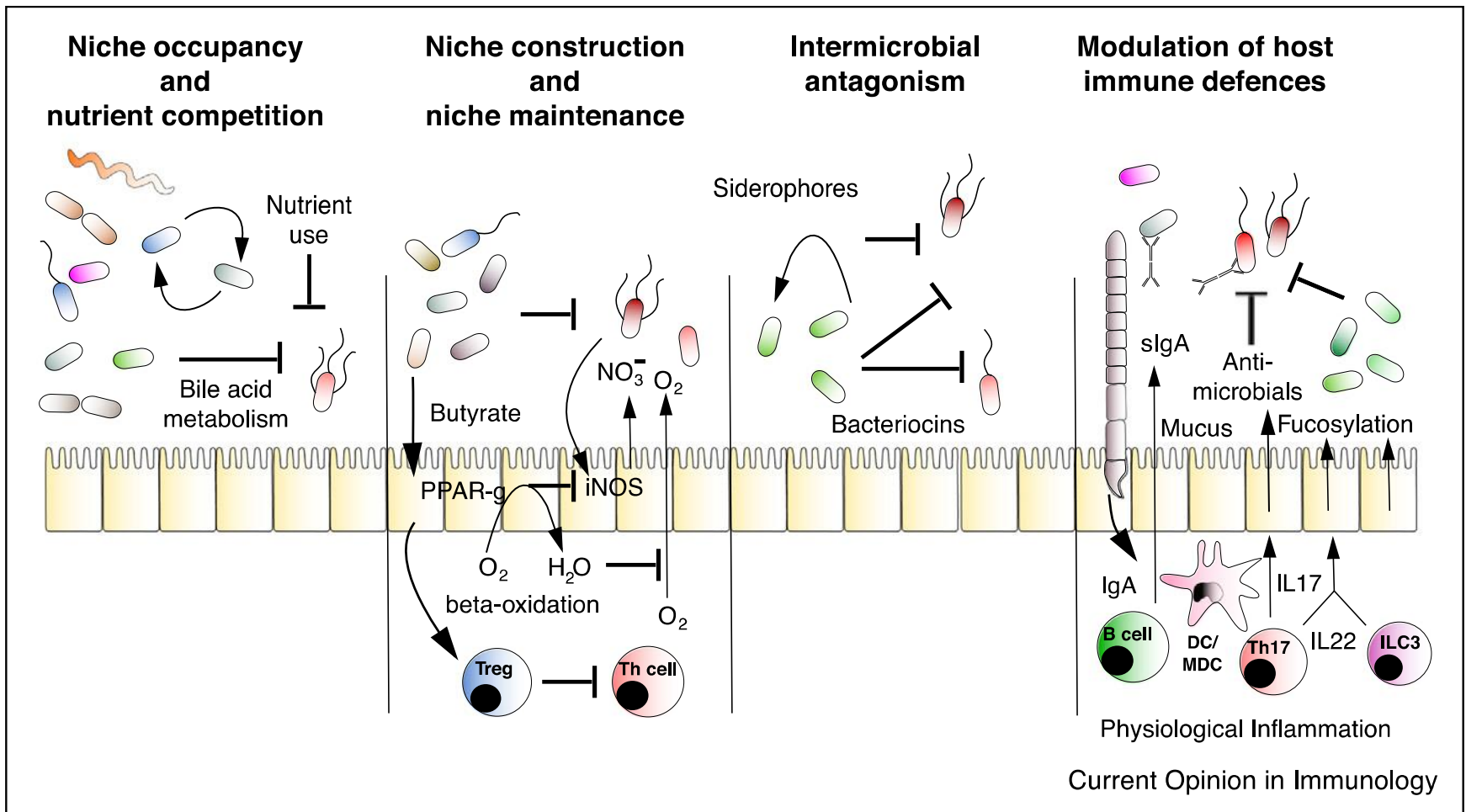
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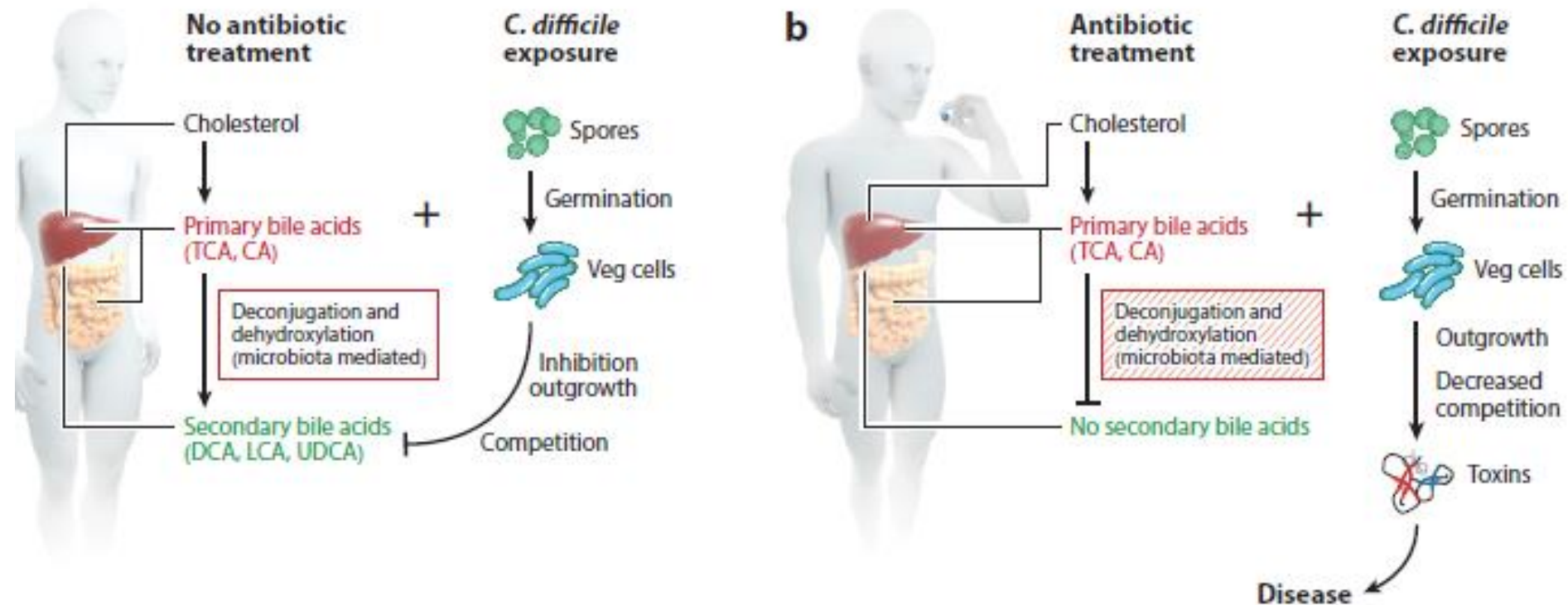




# Mechanisms involved in gut microbiota-mediated resistance against enteropathogen infection



# Antibiotic-induced alterations in gut microbial metabolism decrease colonization resistance against *C. difficile*



Antibiotic treatment alters the gut microbiota structure, specifically decreasing bacteria that are able to deconjugate and dehydroxylate primary bile acids into secondary bile acids. The loss of secondary bile acid metabolism and competition from the gut microbiota allow for *C. difficile* outgrowth, toxin production, and disease.

## Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent *Clostridium difficile* infection

G. Cammarota\*, L. Masucci<sup>†</sup>, G. Ianaro\*, S. Bibbò\*, G. Dinoi\*, G. Costamagna<sup>‡</sup>, M. Sanguinetti<sup>†</sup> & A. Gasbarrini\*

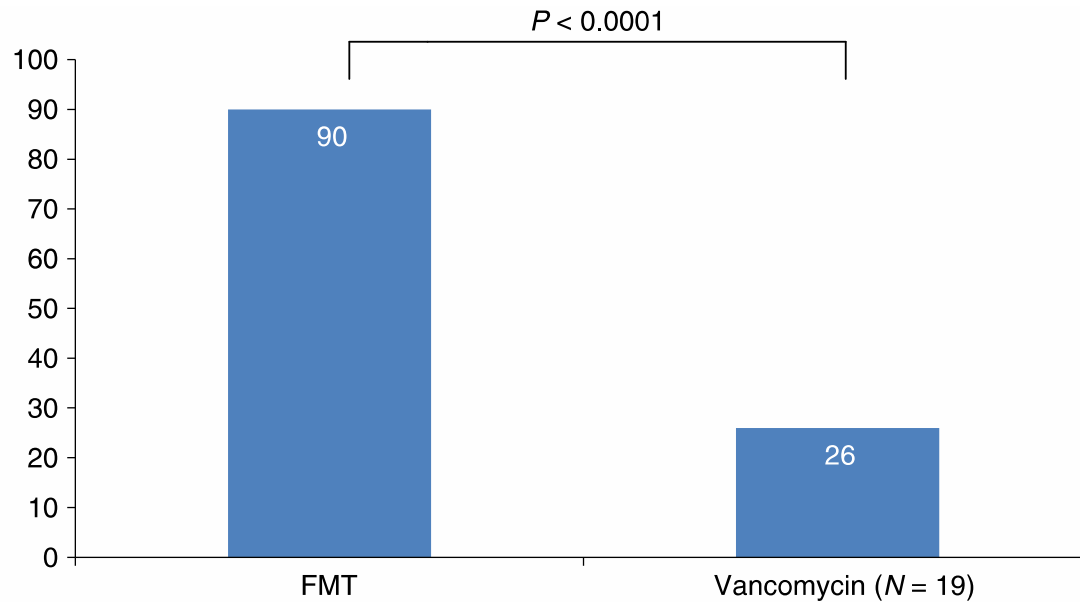
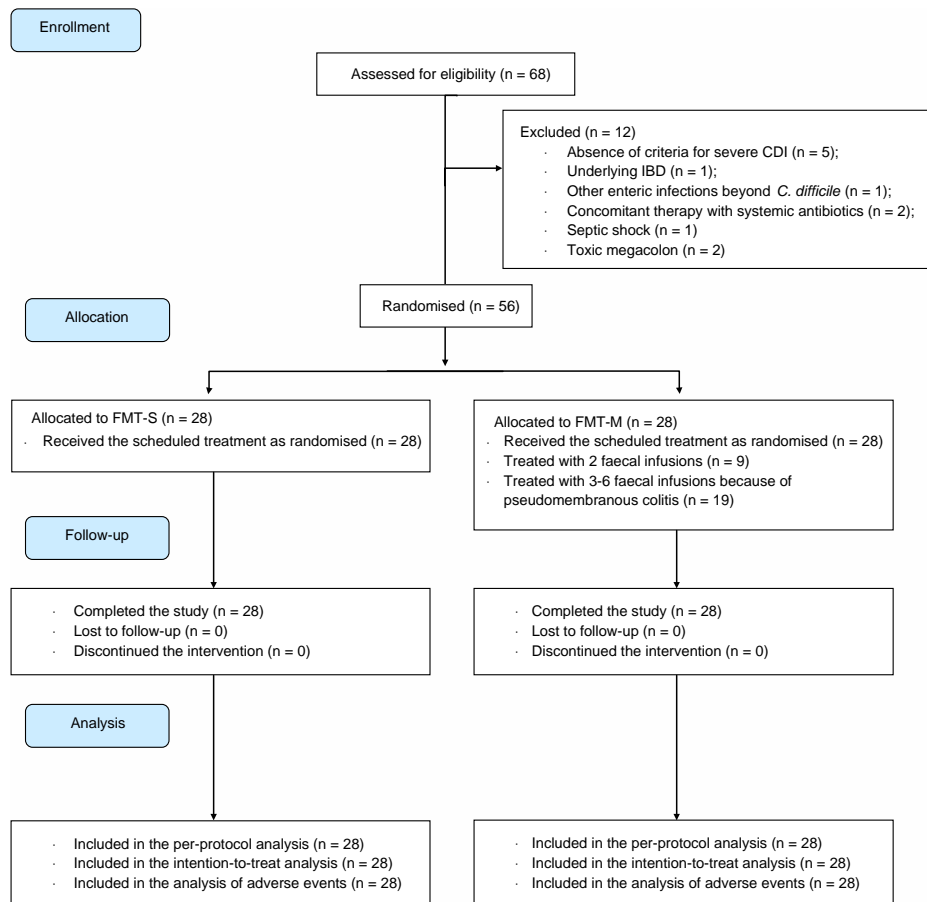


Figure 2 | Percentage of patients cured.

# Randomised clinical trial: faecal microbiota transplantation by colonoscopy plus vancomycin for the treatment of severe refractory *Clostridium difficile* infection—single versus multiple infusions

G. Ianiro<sup>1</sup>  | L. Masucci<sup>2</sup> | G. Quaranta<sup>2</sup> | C. Simonelli<sup>1</sup> | L. R. Lopetuso<sup>1</sup>  |  
M. Sanguinetti<sup>2</sup> | A. Gasbarrini<sup>1</sup> | G. Cammarota<sup>1</sup> 



- Subjects with severe *Clostridioides difficile* infection refractory to antibiotics were randomly assigned to one of the two following treatment arms:
  - FMT-S, including a single faecal infusion via colonoscopy followed by a 14-day vancomycin course,
  - FMT-M, including multiple faecal infusions plus a 14-day vancomycin course. In the FMT-M group, all subjects received at least two infusions.
- The primary outcome was the cure of refractory severe *Clostridioides difficile* infection.
- Fifty six subjects, 28 in each treatment arm, were enrolled.
- Twenty one patients in the FMT-S group and 28 patients in the FMT-M group were cured (75% vs 100%, respectively, both in per protocol and intention-to-treat analyses;  $P = 0.01$ ).
- No serious adverse events associated with any of the two treatment protocols were observed.

# Fecal Microbiota Transplantation at the “A. Gemelli” Hospital - Overall Data (years 2013–2018)

- ❑ **150 patients treated** (mean age, 73 years; range, 29–94 years)
- ❑ Mean number of recurrences of *Clostridium difficile* infection (rCDI) = **3 (range = 2–11)**
- ❑ Mean Charlson Comorbidity Index score = 3
- ❑ Inpatients/Outpatients = 100/50
- ❑ **Overall 223 infusion procedures** (91 patients received one infusion, 79 multiple infusions)
- ❑ Fresh material/Frozen material= 126/87

## FMT procedures/year

- June–December 2013 = 9 (7 pts)
- 2014 = 24 (15 pts)
- 2015 = 36 (25 pts)
- 2016 = 49 (30 pts)
- 2017 = 58 (40 pts)
- Jan-Jul 2018= 47 (33 pts)

## RESULTS:

- Severe colitis (PMC) in 65 patients
- Follow-up range = 1–60 months
- **Resolution of rCDI in 147/150 treated patients (98%)**
- **No patients experienced further FMT recurrences**
- **No more surgery for CDI in our hospital since 2014**

# L'infezione da *Clostridium difficile* è uno dei temi caldi in ospedale

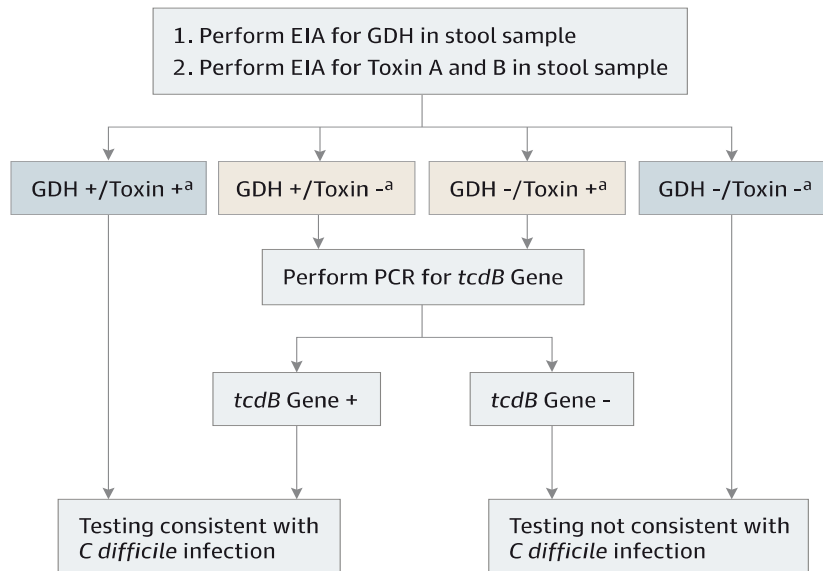
PROCEDURA	Rev.: 0
PERCORSO CLINICO ASSISTENZIALE DEL PAZIENTE CON INFEZIONE DA CLOSTRIDIUM DIFFICILE (CDI)	PRO.POL.RMA. 188

Adopted procedure for the management of *C. difficile* infection at the “A. Gemelli” Hospital

Procedura  
Percorso Clinico Assistenziale del paziente  
con Infezione da *Clostridium difficile* (CDI)

Questo PCA è stato **attivato nel 2015** per rispondere all'esigenza di tenere sotto controllo il fenomeno dell'infezione da *Clostridium difficile* identificando una procedura standard. Questa procedura prevedeva l'introduzione di algoritmi e test diagnostici innovativi

## Detailed Diagnostic Steps



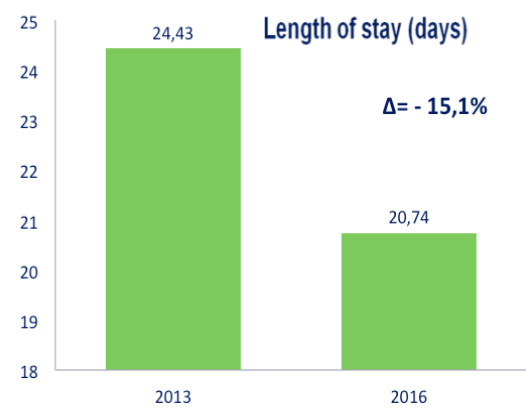
Il PCA prevede tre passi principali già in PS:

1. Identificazione del paziente con sospetta infezione da *Clostridium difficile*
2. Valutazione microbiologica real-time del paziente
3. Gestione del paziente a seconda dell'esito degli approfondimenti diagnostici (**inclusa la possibilità di FMT**)

# The pre-post analysis of the procedure implementation revealed impacts on the length of stay, on the appropriateness of the treatment and on mortality

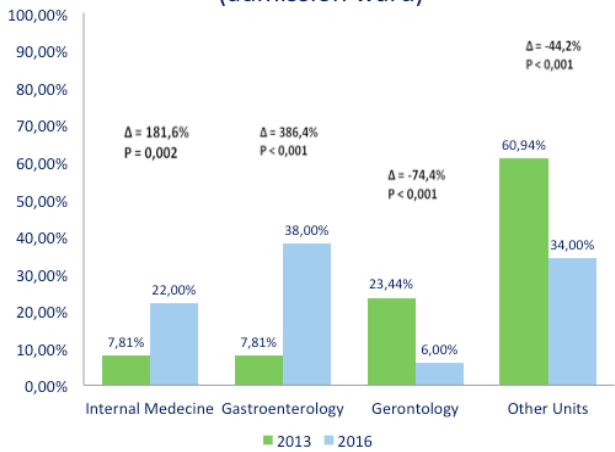
Lenght of stay (days)

*Clostridium difficile* ADMISSIONS– 2013 vs 2016 (lenght of stay)



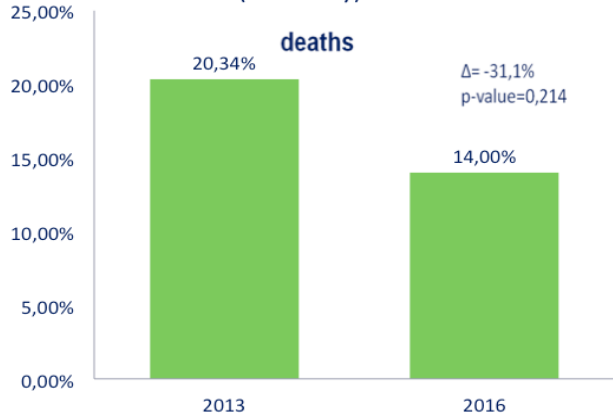
Ward of treatment

*Clostridium difficile* ADMISSIONS – 2013 vs 2016 (admission ward)

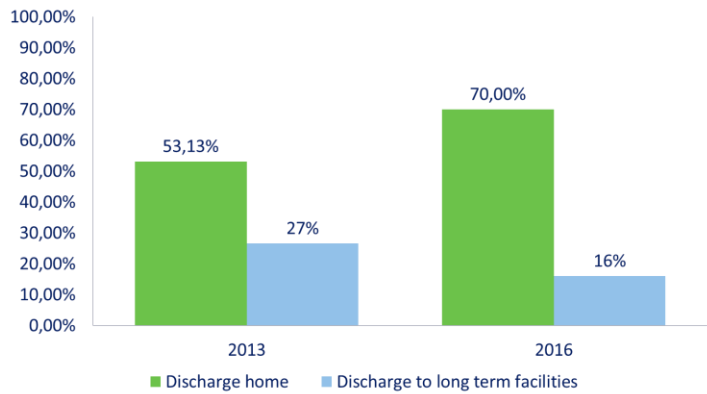


Hospital mortality (%)

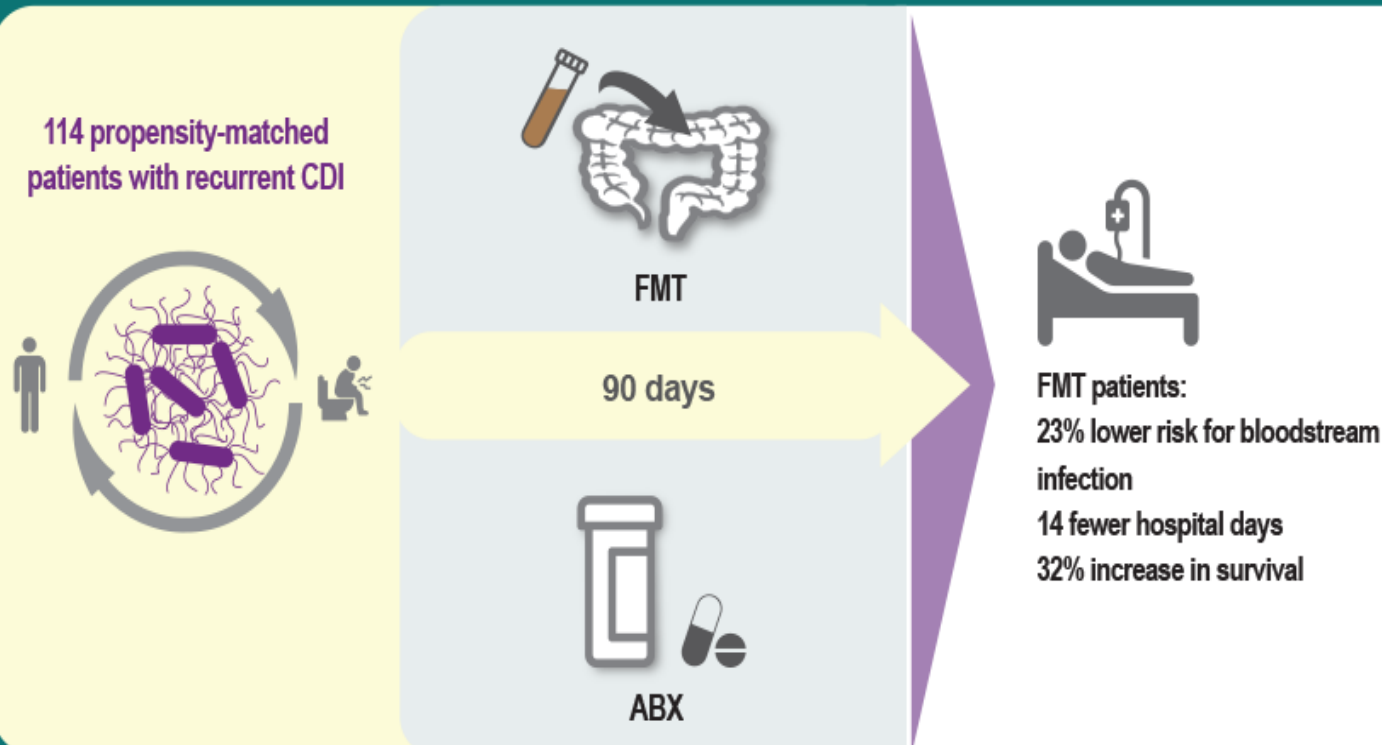
*Clostridium difficile* ADMISSIONS – 2013 vs 2016 (Mortality)



*Clostridium difficile* ADMISSIONS – 2013 vs 2016 (Discharge)



Is treatment of recurrent *Clostridioides difficile* infection (CDI) with fecal microbiota transplantation (FMT) versus antibiotics (ABX) associated with incidence of CDI-associated bloodstream infection?





# Incidence of Bloodstream Infections, Length of Hospital Stay, and Survival in Patients With Recurrent *Clostridioides difficile* Infection Treated With Fecal Microbiota Transplantation or Antibiotics

## A Prospective Cohort Study

Gianluca Ianiro, MD; Rita Murri, MD; Giusi Desirè Sciumè, MD; Michele Impagnatiello, MD; Luca Masucci, MD; Alexander C. Ford, MBChB, MD; Graham R. Law, PhD; Herbert Tilg, MD; Maurizio Sanguinetti, MD; Roberto Cauda, MD, PhD; Antonio Gasbarrini, MD; Massimo Fantoni, MD; and Giovanni Cammarota, MD

Table 2. Outcome Data in the Original Cohort and in the Propensity Score-Matched Cohort

Variable	Original Cohort			After Propensity Score Matching		
	Treated With FMT	Treated With Antibiotics	Difference, %	Treated With FMT	Treated With Antibiotics	Difference (95% CI), %
<b>Patients, n</b>	109	181	–	57	57	–
<b>Primary outcomes, n (%)</b>						
BSI	5 (5)	40 (22)	16	2 (4)	15 (26)	23 (10–35)
Polymicrobial*	1 (1)	11 (6)	–	0 (0)	0 (0)	–
Bacterial	5 (5)	28 (15)	–	2 (4)	8 (14)	–
Fungal	0 (0)	12 (7)	–	0 (0)	7 (12)	–
<b>Secondary outcomes</b>						
Length of hospitalization	–	–	24	–	–	14 (9–20)
Mean (SD), d	13.3 (14.8)	29.7 (22.6)	–	13.4 (13.7)	27.8 (17.6)	–
Median (interquartile range), d	8 (2–20)	22 (14–39)	–	9 (2–21)	22 (14–40)	–
Overall survival at 90 d	–	–	30	–	–	32 (16–47)
Alive after 90 d, n (%)	100 (92)	111 (61)	–	51 (89)	33 (58)	–
Total deaths within 90 d, n (%)	9 (8)	70 (39)	–	6 (11)	22 (39)	–
Deaths in days 0–30, n	5	53	–	3	15	–
Deaths in days 31–90, n	4	17	–	3	7	–

BSI = bloodstream infection; FMT = fecal microbiota transplantation.

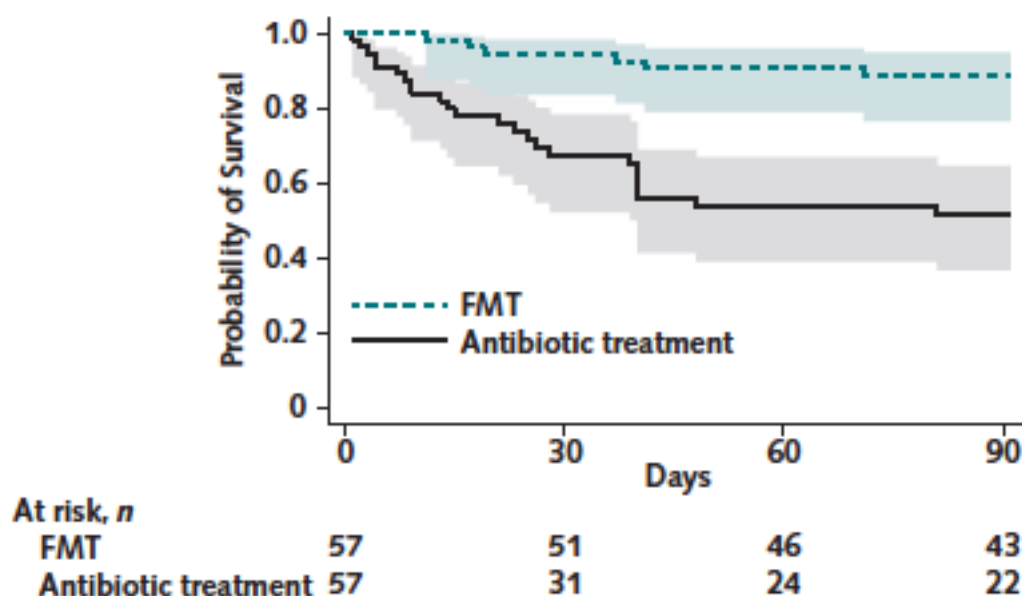
\* 12 of 45 patients developed a polymicrobial BSI (from multiple bacteria in 10 patients and from fungal and bacterial organisms in 2 patients).

# Incidence of Bloodstream Infections, Length of Hospital Stay, and Survival in Patients With Recurrent *Clostridioides difficile* Infection Treated With Fecal Microbiota Transplantation or Antibiotics

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**Figure 2.** Overall survival at 90 d in 57 patients treated with FMT compared with 57 patients treated with antibiotics matched by propensity score.



Shaded areas are 95% CIs. FMT = fecal microbiota transplantation.

BRIEF REPORT

## Drug-Resistant *E. coli* Bacteremia Transmitted by Fecal Microbiota Transplant

Zachariah DeFilipp, M.D., Patricia P. Bloom, M.D., Mariam Torres Soto, M.A.,  
Michael K. Mansour, M.D., Ph.D., Mohamad R.A. Sater, Ph.D.,  
Miriam H. Huntley, Ph.D., Sarah Turbett, M.D., Raymond T. Chung, M.D.,  
Yi-Bin Chen, M.D., and Elizabeth L. Hohmann, M.D.

### SUMMARY

Fecal microbiota transplantation (FMT) is an emerging therapy for recurrent or refractory *Clostridioides difficile* infection and is being actively investigated for other conditions. We describe two patients in whom extended-spectrum beta-lactamase (ESBL)-producing *Escherichia coli* bacteremia occurred after they had undergone FMT in two independent clinical trials; both cases were linked to the same stool donor by means of genomic sequencing. One of the patients died. Enhanced donor screening to limit the transmission of microorganisms that could lead to adverse infectious events and continued vigilance to define the benefits and risks of FMT across different patient populations are warranted.



## European consensus conference on faecal microbiota transplantation in clinical practice

Giovanni Cammarota,<sup>1</sup> Gianluca Ianiro,<sup>1</sup> Herbert Tilg,<sup>2</sup> Mirjana Rajilić-Stojanović,<sup>3</sup> Patrizia Kump,<sup>4</sup> Reetta Satokari,<sup>5</sup> Harry Sokol,<sup>6</sup> Perttu Arkkila,<sup>7</sup> Cristina Pintus,<sup>8</sup> Ailsa Hart,<sup>9</sup> Jonathan Segal,<sup>9</sup> Marina Aloï,<sup>10</sup> Luca Masucci,<sup>11</sup> Antonio Molinaro,<sup>12</sup> Franco Scaldaferri,<sup>1</sup> Giovanni Gasbarrini,<sup>1</sup> Antonio Lopez-Sanroman,<sup>13</sup> Alexander Link,<sup>14</sup> Pieter de Groot,<sup>15</sup> Willem M de Vos,<sup>5,16</sup> Christoph Högenauer,<sup>4</sup> Peter Malfertheiner,<sup>14</sup> Eero Mattila,<sup>17</sup> Tomica Milosavljević,<sup>18</sup> Max Nieuwdorp,<sup>12,15,19</sup> Maurizio Sanguinetti,<sup>11</sup> Magnus Simren,<sup>20</sup> Antonio Gasbarrini,<sup>1</sup> The European FMT Working Group

Cammarota G, *et al. Gut* 2017;**66**:569–580.

### Box 3 Blood and stool testing to check donors for any potentially transmittable disease

#### GENERAL BLOOD TESTING

- ▶ Cytomegalovirus
- ▶ Epstein-Barr virus
- ▶ Hepatitis A
- ▶ HBV
- ▶ HCV
- ▶ Hepatitis E virus
- ▶ Syphilis
- ▶ HIV-1 and HIV-2
- ▶ *Entamoeba histolytica*
- ▶ Complete blood cell count with differential
- ▶ C-reactive protein and erythrocyte sedimentation rate
- ▶ Albumin
- ▶ Creatinine and electrolytes
- ▶ Aminotransferases, bilirubin, gamma-glutamyltransferase, alkaline phosphatase

#### BLOOD TESTING IN SPECIFIC SITUATIONS

- ▶ Human T-lymphotropic virus types I and II antibodies
- ▶ *Strongyloides stercoralis*

#### GENERAL STOOL TESTING

- ▶ Detection of *Clostridium difficile*
- ▶ Detection of enteric pathogens, including *Salmonella*, *Shigella*
- ▶ *Campylobacter*, *Escherichia coli* O157 H7, *Yersinia*, vancomycin-resistant enterococci, methicillin-resistant *Staphylococcus aureus*, Gram-negative multidrug-resistant bacteria
- ▶ Norovirus
- ▶ Antigens and/or acid fast staining for *Giardia lamblia* and *Cryptosporidium parvum*
- ▶ Protozoa (including *Blastocystis hominis*) and helminths
- ▶ Faecal occult blood testing

#### STOOL TESTING IN SPECIFIC SITUATIONS

- ▶ Detection of *Vibrio cholera* and *Listeria monocytogenes*
- ▶ Antigens and/or acid fast staining for *Isospora* and *Microsporidia*
- ▶ Calprotectin
- ▶ *Helicobacter pylori* faecal antigen
- ▶ Rotavirus

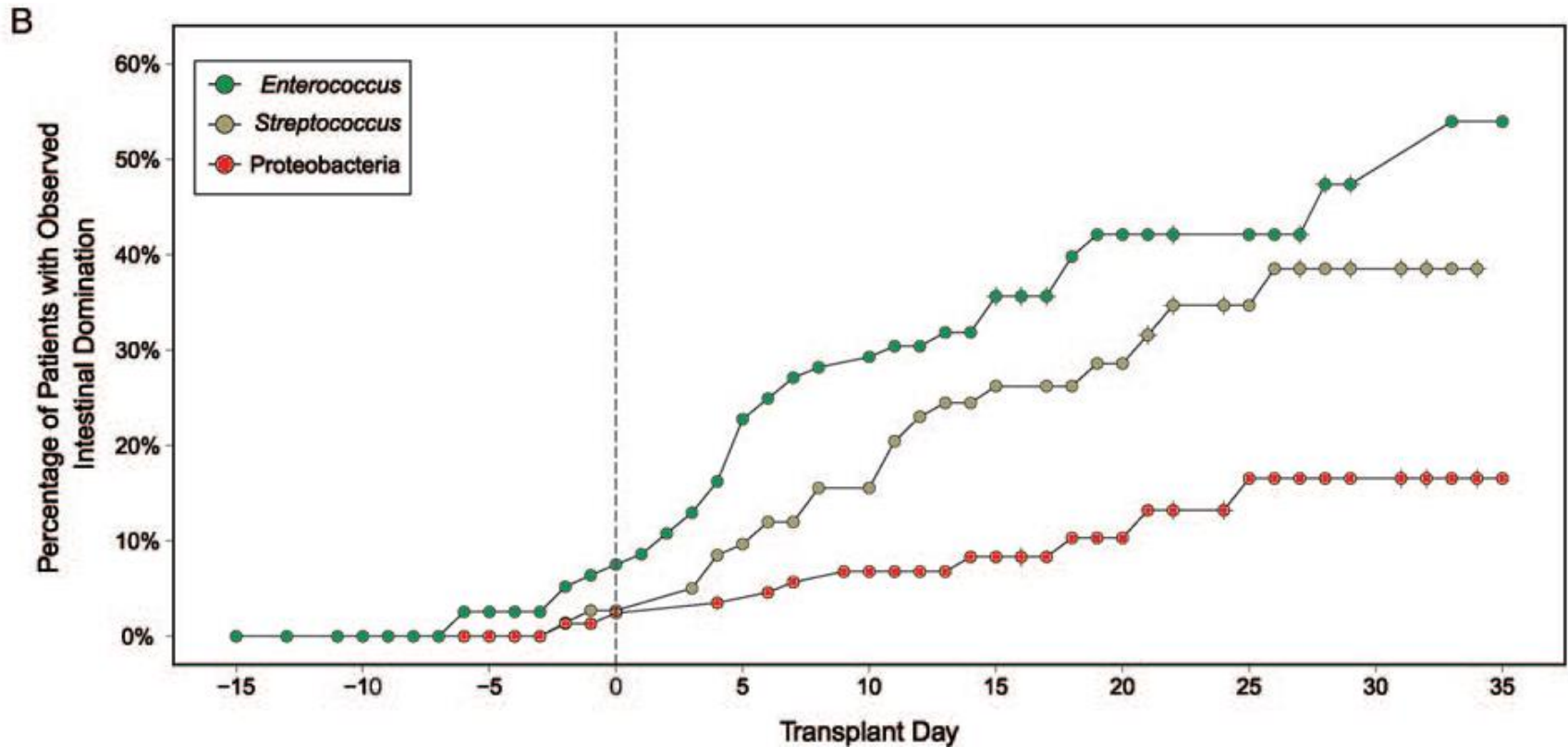
# Microbiota destruction and antibiotic-resistant infections

- Recommendations to combat the progression of antibiotic resistance included barrier and hygiene approaches to reduce transmission, limits on antibiotic use, and development of new and more effective antibiotics.
- A concern with these approaches, however, is that antibiotic resistance has grown despite their implementation.
- Recently, the White House provided a national action plan for combating antibiotic-resistant bacteria, which included specific milestones that introduced the potential role of the microbiome and the microbiota in combating antibiotic resistance ([www.whitehouse.gov/sites/default/files/docs/national\\_action\\_plan\\_for\\_combating\\_antibiotic-resistant\\_bacteria.pdf](http://www.whitehouse.gov/sites/default/files/docs/national_action_plan_for_combating_antibiotic-resistant_bacteria.pdf)).

# Intestinal Domination and the Risk of Bacteremia in Patients Undergoing Allogeneic Hematopoietic Stem Cell Transplantation

Ying Taur,<sup>1,2</sup> Joao B. Xavier,<sup>2,3</sup> Lauren Lipuma,<sup>2</sup> Carlos Ubeda,<sup>5</sup> Jenna Goldberg,<sup>4</sup> Asia Gobourne,<sup>2</sup> Yeon Joo Lee,<sup>1</sup> Krista A. Dubin,<sup>2</sup> Nicholas D. Socci,<sup>3</sup> Agnes Viale,<sup>6</sup> Miguel-Angel Perales,<sup>4</sup> Robert R. Jenq,<sup>4</sup> Marcel R. M. van den Brink,<sup>4,5</sup> and Eric G. Pamer<sup>1,2,5</sup>

CID 2012;55 (1 October)



Characterization of the intestinal microbiota during allogeneic hematopoietic stem cell transplantation

# Intestinal Domination and the Risk of Bacteremia in Patients Undergoing Allogeneic Hematopoietic Stem Cell Transplantation

Ying Taur,<sup>1,2</sup> Joao B. Xavier,<sup>2,3</sup> Lauren Lipuma,<sup>2</sup> Carles Ubeda,<sup>5</sup> Jenna Goldberg,<sup>4</sup> Asia Gobourne,<sup>2</sup> Yeon Joo Lee,<sup>1</sup> Krista A. Dubin,<sup>2</sup> Nicholas D. Socci,<sup>3</sup> Agnes Viale,<sup>6</sup> Miguel-Angel Perales,<sup>4</sup> Robert R. Jenq,<sup>4</sup> Marcel R. M. van den Brink,<sup>4,5</sup> and Eric G. Pamer<sup>1,2,5</sup>

CID 2012;55 (1 October)

**Table 3. Association of Intestinal Domination With Bacteremia<sup>a</sup>**

Dominating Taxon <sup>b</sup>	VRE Bacteremia		Gram-negative Bacteremia	
	HR (95% CI)	<i>P</i>	HR (95% CI)	<i>P</i>
<i>Enterococcus</i>	9.35 (2.43–45.44)	.001	1.35 (.25–5.08)	.690
<i>Streptococcus</i>	0.21 (.00–1.75)	.184	0.82 (.09–3.65)	.823
Proteobacteria	0.75 (.01–6.14)	.837	5.46 (1.03–19.91)	.047

Abbreviations: CI, confidence interval; HR, hazard ratio; VRE, Vancomycin-resistant *Enterococcus*.

<sup>a</sup> Bacteremia for each organism was defined as at least one positive blood culture within the study period.

<sup>b</sup> Intestinal domination was analyzed as a time-varying predictor.

# Gut Microbiome Composition Predicts Infection Risk During Chemotherapy in Children With Acute Lymphoblastic Leukemia

Hana Hakim,<sup>1</sup> Ronald Dallas,<sup>1</sup> Joshua Wolf,<sup>1</sup> Li Tang,<sup>2</sup> Stacey Schultz-Cherry,<sup>1</sup> Victoria Darling,<sup>1</sup> Cydney Johnson,<sup>1</sup> Erik A. Karlsson,<sup>1</sup> Ti-Cheng Chang,<sup>3</sup> Sima Jeha,<sup>4</sup> Ching-Hon Pui,<sup>4</sup> Yilun Sun,<sup>2</sup> Stanley Pounds,<sup>2</sup> Randall T. Hayden,<sup>5</sup> Elaine Tuomanen,<sup>1</sup> and Jason W. Rosch<sup>1</sup>

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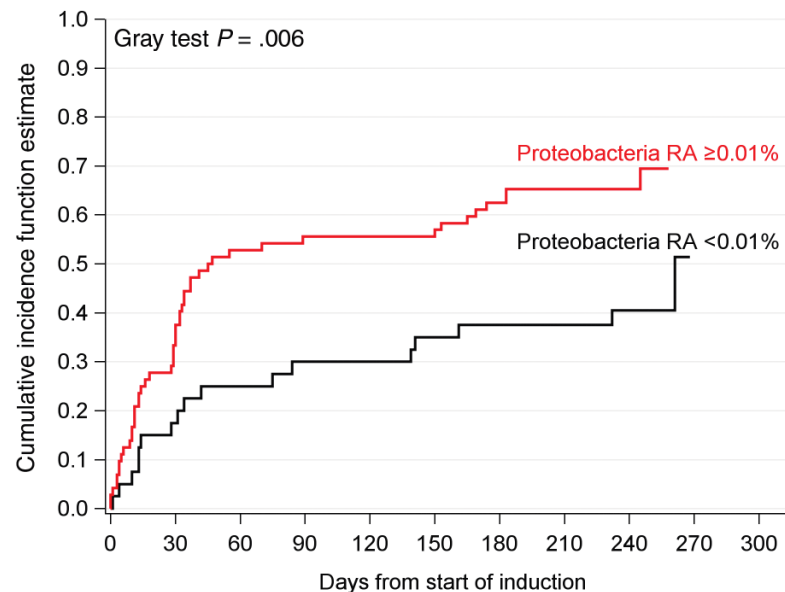
- After the induction and reinduction I phases of chemotherapy, microbial diversity decreased significantly relative to the prechemotherapy value.
- After chemotherapy, the relative abundance of certain bacterial taxa (eg, *Bacteroidetes*) decreased significantly, whereas that of other taxa (eg, *Clostridiaceae* and *Streptococcaceae*) increased.
- A baseline gut microbiome characterized by *Proteobacteria* predicted febrile neutropenia. Adjusting for the chemotherapy phase and ALL risk level, *Enterococcaceae* dominance (relative abundance  $\geq 30\%$ ) predicted significantly greater risk of subsequent febrile neutropenia and diarrheal illness, whereas *Streptococcaceae* dominance predicted significantly greater risk of subsequent diarrheal illness.
- In children undergoing therapy for newly diagnosed ALL, the relative abundance of *Proteobacteria* before chemotherapy initiation predicts development of febrile neutropenia, and domination of the gut microbiota by *Enterococcaceae* or *Streptococcaceae* at any time during chemotherapy predicts infection in subsequent phases of chemotherapy.



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**Figure 3.** The relative abundance (RA) of Proteobacteria in the baseline gut microbiota at acute lymphoblastic leukemia diagnosis is associated with increased cumulative incidence of first episodes of febrile neutropenia. The estimated hazard ratio was 2.12 (95% confidence interval, 1.22–3.69;  $P = .008$ ) for a Proteobacteria RA of  $\geq 0.01$  vs an RA of  $< 0.01\%$ , adjusting for gender.

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- Of 14 patients with dominant *Enterococcaceae*, 7 (50%) developed subsequent infection, as did 4 of 5 (80%) patients with dominant *Streptococcaceae*.
- Adjusting for chemotherapy phase and ALL risk level, *Enterococcaceae* dominance was significantly associated with increased risk of subsequent febrile neutropenia (HR, 2.97) and diarrheal illness (HR, 4.23) compared to nondominance of both *Streptococcaceae* and *Enterococcaceae*, while *Streptococcaceae* dominance carried a greater risk (HR, 7.94) of subsequent diarrheal illness.
- Thus, a gut microbiota dominated by *Enterococcaceae* or *Streptococcaceae* is associated with subsequent infections.

# Increased Relative Abundance of *Klebsiella pneumoniae* Carbapenemase-producing *Klebsiella pneumoniae* Within the Gut Microbiota Is Associated With Risk of Bloodstream Infection in Long-term Acute Care Hospital Patients

Teppei Shimasaki,<sup>1,✉</sup> Anna Seekatz,<sup>2</sup> Christine Bassis,<sup>2</sup> Yoona Rhee,<sup>1</sup> Rachel D. Yelin,<sup>1</sup> Louis Fogg,<sup>3</sup> Thelma Dangana,<sup>1</sup> Enrique Cornejo Cisneros,<sup>1,4</sup> Robert A. Weinstein,<sup>1</sup> Koh Okamoto,<sup>1,5</sup> Karen Lolans,<sup>1</sup> Michael Schoeny,<sup>3</sup> Michael Y. Lin,<sup>1</sup> Nicholas M. Moore,<sup>1</sup> Vincent B. Young,<sup>2</sup> and Mary K. Hayden<sup>1</sup>; for the Centers for Disease Control and Prevention Epicenters Program

- In this study were collected 2319 samples from 562 admissions (506 patients); KPC-Kp colonization was detected in 255 (45.4%) admissions and KPC-Kp bacteremia in 11 (4.3%).
- A relative abundance cutoff of 22% predicted KPC-Kp bacteremia with sensitivity 73%, specificity 72%, and **relative risk 4.2 (P = .01)**.
- In a multivariable Cox regression model adjusted for age, Charlson comorbidity index, and medical devices, carbapenem receipt was associated with achieving the 22% relative abundance threshold (P = .044).
- Carbapenem receipt was associated with increased hazard for high relative abundance of KPC-Kp in the gut microbiota. Increased relative abundance of KPC-Kp was associated with KPC-Kp bacteremia.

CASE REPORT

Open Access



# First bloodstream infection caused by *Prevotella copri* in a heart failure elderly patient with *Prevotella*-dominated gut microbiota: a case report

Patrizia Posteraro<sup>1†</sup>, Flavio De Maio<sup>2†</sup>, Giulia Menchinelli<sup>2</sup>, Ivana Palucci<sup>2</sup>, Federica Maria Errico<sup>1</sup>, Mariantonietta Carbone<sup>3</sup>, Maurizio Sanguinetti<sup>2,4\*</sup> , Antonio Gasbarrini<sup>5,6\*</sup> and Brunella Posteraro<sup>5,6</sup>

- We report the case of a 90-year-old heart failure (HF) patient who was admitted to the hospital for an

**Table 1** Most relatively abundant bacteria on order, family, genus, and species level identified in the patient's fecal sample

Taxa	% abundance	Taxa	% abundance
Order		Genus	
<i>Bacteroidales</i>	34.114	<i>Prevotella</i>	15.253
<i>Clostridiales</i>	27.590	<i>Faecalibacterium</i>	8.215
<i>Selenomonadales</i>	7.493	<i>Dialister</i>	6.372
		<i>Parabacteroides</i>	3.849
		<i>Bacteroides</i>	3.750
Family		Species	
<i>Prevotellaceae</i>	19.024	<i>Prevotella copri</i>	6.133
<i>Ruminococcaceae</i>	16.519	<i>Dialister succinatiphilus</i>	5.869
<i>Lachnospiraceae</i>	9.157	<i>Faecalibacterium prausnitzii</i>	3.180
<i>Porphyromonadaceae</i>	6.999	<i>Parabacteroides distasonis</i>	2.488
<i>Veillonellaceae</i>	6.758	<i>Prevotella</i> sp. (DJF RP53)	2.050
<i>Bacteroidaceae</i>	3.750		

- The listed taxa belong to the Firmicutes and Bacteroidetes phyla, whose relative abundances were 37.054% and 34.664%, respectively. Unlisted species include two other *Prevotella* sp. (BI-42 and DJF B112), whose relative abundances were 1.504% and 0.836%, respectively

# Fecal Microbiota Transplantation in Patients With Blood Disorders Inhibits Gut Colonization With Antibiotic-Resistant Bacteria: Results of a Prospective, Single-Center Study

Jarosław Bilinski,<sup>1</sup> Paweł Grzesiowski,<sup>2</sup> Nikolaj Sørensen,<sup>3</sup> Krzysztof Madry,<sup>1</sup> Jacek Muszyński,<sup>4</sup> Katarzyna Robak,<sup>1</sup> Marta Wroblewska,<sup>5,6</sup> Tomasz Dzieciatkowski,<sup>5</sup> Grażyna Dulny,<sup>7</sup> Jadwiga Dwilewicz-Trojaczek,<sup>1</sup> Wiesław Wiktor-Jedrzejczak,<sup>1</sup> and Grzegorz W. Basak<sup>1</sup>  
*Clinical Infectious Diseases*<sup>®</sup> 2017;65(3):364–70

**Table 2. Impact of Fecal Microbiota Transplantation on Complete and Partial Antibiotic-Resistant Bacteria (ARB) Decolonization, With or Without Antibiotics During the First Week After Transplantation**

Endpoint	All FMTs n = 25		With Antibiotics, n = 11		Without Antibiotics, n = 14		PValue
	No.	%	No.	%	No.	%	
Effect on all strains of ARB per FMT (complete ARB decolonization)							
At 1 month	15/25	60	4/11	36	11/14	79	.039
At 6 months	13/14	93	4/5	80	9/9	100	.037
Effect on at least 1 strain of ARB per FMT (partial ARB decolonization)							
At 1 month	20/25	80	7/11	64	13/14	93	NS
At 6 months	13/14	93	4/5	80	9/9	100	–

Abbreviations: ARB, antibiotic-resistant bacteria; FMT, fecal microbiota transplantation.

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**Table 3. Decolonization of Particular Strains of Antibiotic-Resistant Bacteria in All Participants and Those Without Antibiotics Use During the First Week After Fecal Microbiota Transplantation**

Pathogen	Negative Rectal Swab at 1 Week				Decolonization at 1 Month			
	All		Without Antibiotics		All		Without Antibiotics	
	No.	%	No.	%	No.	%	No.	%
<i>Klebsiella pneumoniae</i>								
New Delhi metallo- $\beta$ -lactamase 1	8/14	57	6/6	100	6/10	60 <sup>a</sup>	5/6	83 <sup>a</sup>
Other, carbapenem-resistant	2/3	67	2/2	100	3/3	100	2/2	100
ESBL+	1/2	50	0/1	0	1/2	50	1/1	100
<i>Escherichia coli</i>								
ESBL+	11/11	100	3/3	100	11/11	100	3/3	100
OXA-48 – extended-spectrum oxacillinase-48	1/1	100	1/1	100	1/1	100	1/1	100
<i>Pseudomonas aeruginosa</i>								
MBL+	2/2	100	2/2	100	2/2	100	2/2	100 <sup>a</sup>
Other, carbapenem resistant	1/2	50	1/2	50	2/2	100	2/2	100
Carbapenem-resistant <i>Enterobacter cloacae</i>	1/2	50	1/2	50	2/2	100	2/2	100
Vancomycin-resistant enterococci	2/2	100	1/1	100	2/2	100	1/1	100
<i>Acinetobacter ursingii</i> MBL+	1/1	100	1/1	100	1/1	100 <sup>a</sup>	1/1	100 <sup>a</sup>
<i>Stenotrophomonas maltophilia</i>	1/1	100	1/1	100	1/1	100	1/1	100

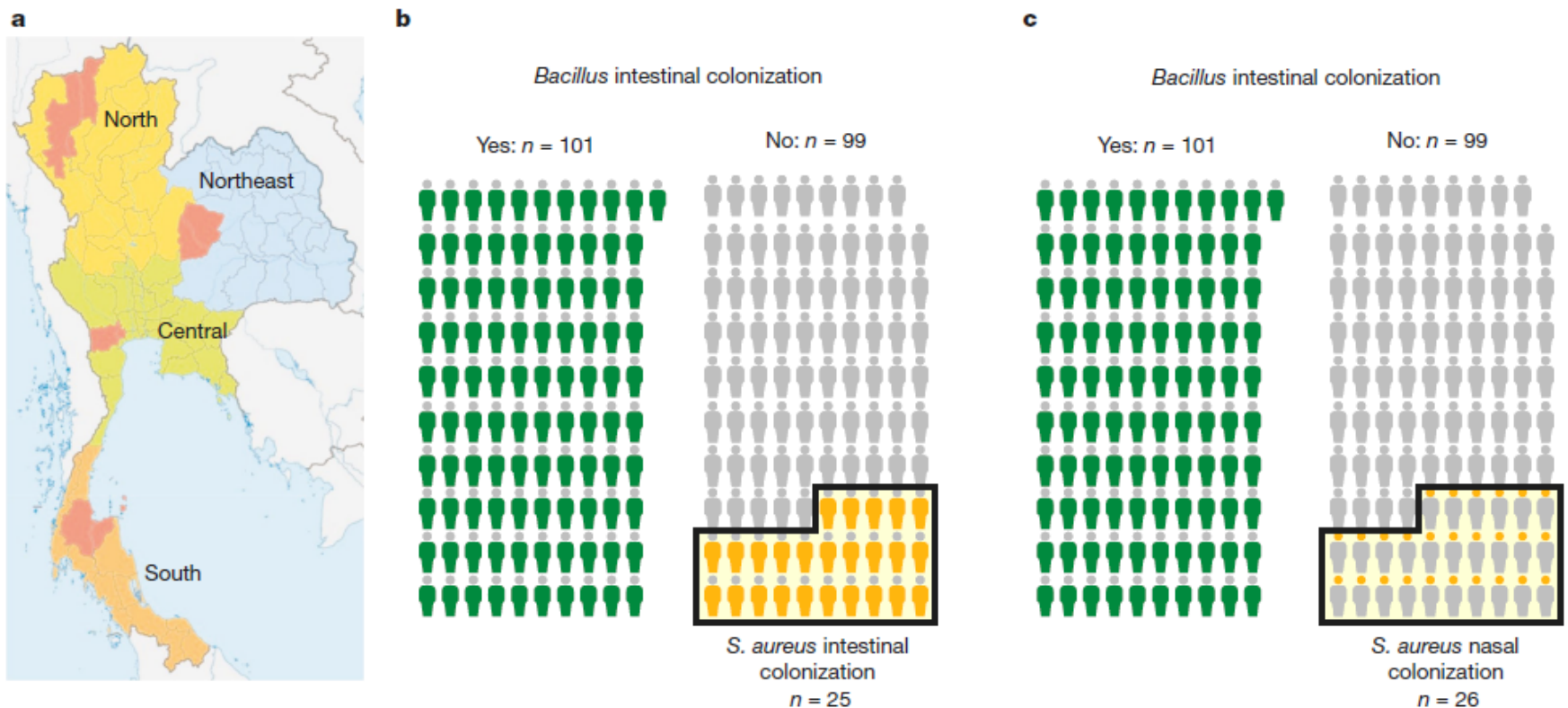
Abbreviations: ESBL, extended-spectrum  $\beta$ -lactamase; MBL, metallo- $\beta$ -lactamase.

<sup>a</sup>Decolonization confirmed by quantitative real-time polymerase chain reaction.

# Pathogen elimination by probiotic *Bacillus* via signalling interference

Pipat Piewngam<sup>1,2</sup>, Yue Zheng<sup>1,5</sup>, Thuan H. Nguyen<sup>1,5</sup>, Seth W. Dickey<sup>1</sup>, Hwang-Soo Joo<sup>1,4</sup>, Amer E. Villaruz<sup>1</sup>, Kyle A. Glose<sup>1</sup>, Emilie L. Fisher<sup>1</sup>, Rachelle L. Hunt<sup>1</sup>, Barry Li<sup>1</sup>, Janice Chiou<sup>1</sup>, Sujiraphong Pharkjaksu<sup>2</sup>, Sunisa Khongthong<sup>3</sup>, Gordon Y. C. Cheung<sup>1</sup>, Pattarachai Kiratisin<sup>2</sup> & Michael Otto<sup>\*</sup>

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**Fig. 1 | Exclusion of *S. aureus* colonization by dietary *Bacillus* in a human population. a, Areas (in red) from which faecal samples were collected in rural populations and analysed for the presence of *Bacillus***

**and *S. aureus*. b, c, Intestinal (b) and nasal (c) colonization with *S. aureus* (yellow) in individuals that showed (green) or did not show (grey) intestinal colonization with *Bacillus*.**



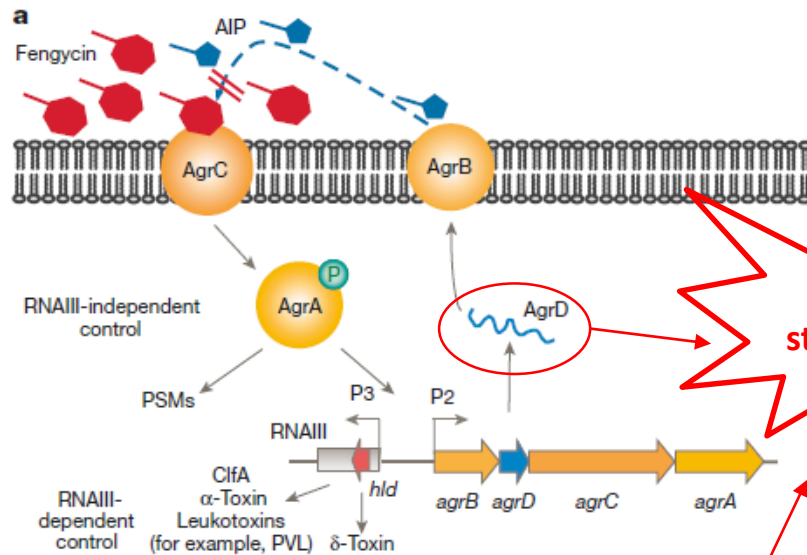
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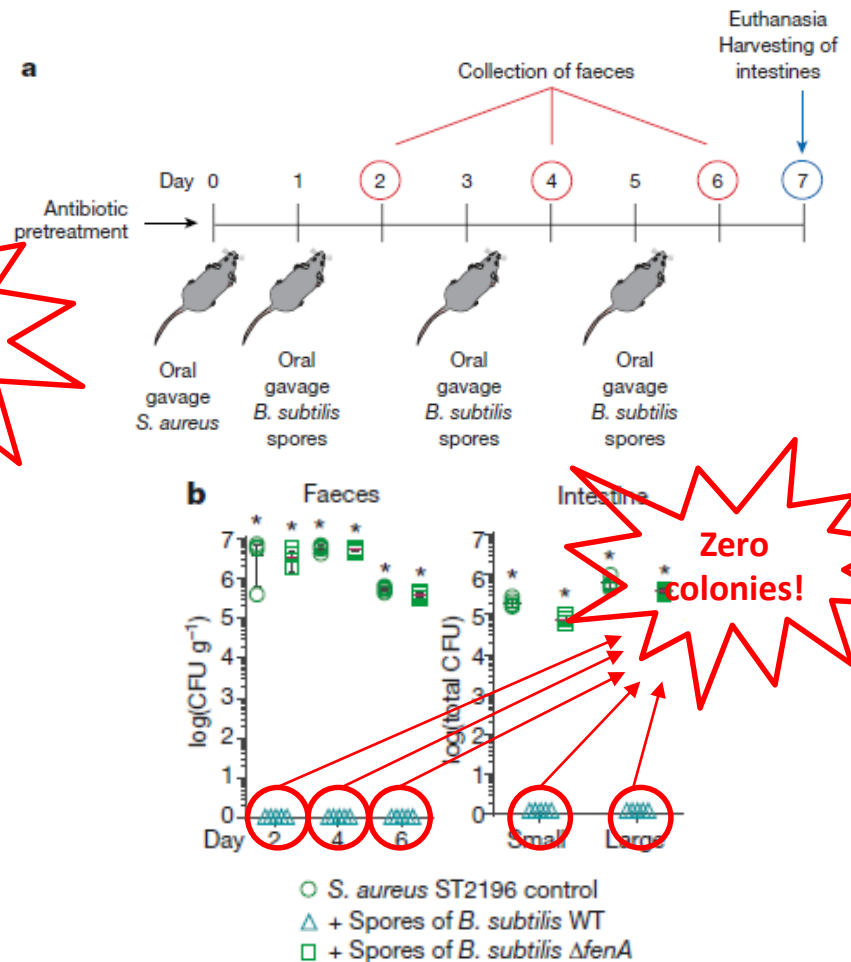
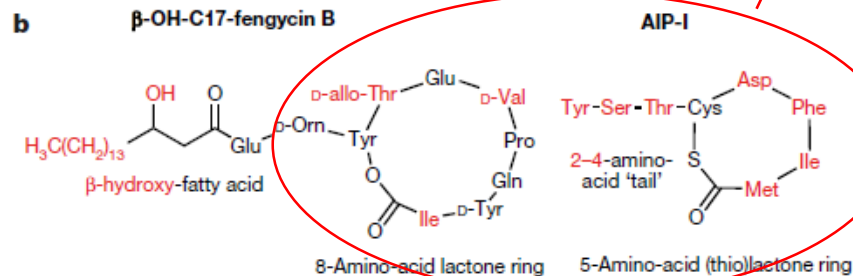
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Inhibition of *S. aureus* colonization by dietary fengycin producing *Bacillus* spores in a mouse model

Competitive inhibition of *S. aureus* AIP activity by fengycins



Similar structure!!!





# Conclusions

- Human microbiota study is one of the most intriguing and exciting research topics in the last years
- In the near future the association of specific microbiota types with specific diseases will permit to make treatment able to restore the “good microbiota” modified by therapeutic approaches
- The assessment of the microbiota composition that has to be used for FMT by -omics will permit to patient tailor this therapeutic procedure
- The use of “Bugs as Drugs”, by FMT, probiotics or phages, is supported by promising evidences derived from basic science and needs to be tested more extensively in randomized clinical trials.

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